Factors Influencing the Survival of Pathogenic Microbes in the Built (i.e. hospitals) and Natural Environments

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In the name of Allah, the Most Merciful, the Most Kind

Dedication

I dedicated this work to my parents (Nahlah Khawajh, Abdulrahim Alaeq) who continue to learn, grow and develop and who has been a source of encouragement and inspiration to me throughout my life. Also, I am grateful to my sisters: Roaa, Rola, Rawan, and Reman and my brothers Ali and Rakan and my friends for their spiritual support and encouragement.

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ABSTRACT

Healthcare associated infections, i.e. nosocomial infections, occur in patients under medical care. These infections occur during stays in hospital and cause prolonged hospitalisation, disability, and an economic burden. Nosocomial pathogens include bacteria, viruses and fungal parasites. Pathogens living in the healthcare environment and equipment can be an obvious source of pathogen transmission. These pathogens can be transmitted by person to person contact or via contaminated water and food, infected individuals, contaminated healthcare personnel's skin or contact via shared items and surfaces. Pathogens can survive in the hospital environment for long periods and, in some cases, resist disinfection. Bacteria are the most common pathogens responsible for nosocomial infections. The Thesis begins with a description of the aims of the work, followed by studies the distribution of bacteria in the environment and their survival in the healthcare settings, the determination of the number of bacteria on hands after washing and drying normally and following the use of a warm air dryer. The factors which influence the survival of bacteria and Candida in the built environment were also determined on ceramic tiles, copper and plastic plumbing surfaces, and on new toothbrushes. Pathogenic bacteria were isolated from used toothbrushes and the effect of toothpastes on the growth of pathogenic bacteria was determined, as was the effectiveness of antibacterial cloths in inhibiting the growth of bacteria and yeast. The metabolic diversity of the bacterial isolates was also determined. Bacteria were isolated from a range of surfaces commonly found in hospitals and health care settings. A wide variety of bacteria were isolated from sinks, computer keyboards and computer mice, taps and the surface of mobile phones and toilet mirrors. Species of *Bacillus* were exclusively isolated from vacuum cleaner dust and from books and dust obtained from a library. Bacteria other than just Bacillus were isolated from the soles of shoes. Lift buttons were found to be contaminated with bacteria, not surprisingly

with species which are typically skin commensals, with the number being highest on the ground floor-call button. Bacteria were found to be spread by hot-air hand dryers, both into the toilet environment and onto previously washed hands. It is provisionally recommended that disposable paper towels are used in preference to such machines. Bacteria and the yeast, Candida rugosa survived when inoculated onto both rough and smooth tile surfaces similar to those used in health care settings. A wide range of potentially pathogenic bacteria were isolated from used toothbrushes and the survival of inoculated bacteria on tooth brushes was determined. The bacteria were shown to survive for varying periods, a finding of some concern in relation to dental hygiene. A range of toothpastes were also shown to be antibacterial. The survival of inoculated bacteria on copper and plastic surfaces typically used as piping in health care settings was determined. Copper surfaces were shown to be antibacterial, while plastic surfaces were not. It is therefore suggested that in critical healthcare areas, copper piping should be given preference over the plastic variety. Proprietary antibacterial clothes were tested for their antibacterial properties. Despite being marketed for this purpose, the cloths showed no obvious, marked antibacterial activity.

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CHAPTER 1

DISTRIBUTION AND SURVIVAL OF BACTERIA IN HEALTHCARE SETTINGS

1.1 Introduction

A wide range of microorganisms can be isolated from the environment, including bacteria, viruses, filamentous fungi and yeasts. Particular emphasis has been placed on the role of bacteria in medicine, but fungi (including yeasts) are also causal agents of many plant and animal diseases although their role as human pathogens is often underplayed. In a recent study setting (Perlroth *et al.*, 2007), for example, fungi were estimated to kill at least as many people as tuberculosis or malaria, over the past decades. The pathogenicity of both bacteria and yeasts is particular potentially dangerous in the case of immunocompromised patients, in which they cause potentially life-threatening diseases (Sullivan *et al.*, 1997).

A hospital-acquired infections (HAIs), also known as a nosocomial infection, are acquired in health care facilities. Health care staff can spread infection, as can contaminated equipment, bed linens, or air droplets (McBryde *et al.*, 2004). The infection can originate from the outside environment, infected patients or staff or in some cases the source of the infection is unknown. The microorganism may originate from the patient's own skin microflora, becoming opportunistic after surgery or other procedures that compromise the protective barrier of the skin. It is estimated that in The US around 1.7 million hospital-associated infections occur

and contribute to 99,000 deaths each year. Nosocomial infections can cause severe pneumonia and infections of the bloodstream, urinary tract and other areas of the body. Many of these infections are difficult to treat with antibiotics (Klevens *et al.*, 2007).

1.2 Organisms involved

Methicillin resistant *Staphylococcus aureus*, *Candida albicans*, *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Stenotrophomonas maltophilia*, *Clostridium difficile*, *Escherichia coli*, and Vancomycin-resistant *Enterococcus*.

1.3 Sterilization

Sterilization kills all microbes on equipment and surfaces through exposure to chemicals, ionizing radiation, dry heat, or steam under pressure.

1.4 Isolation

Isolation precautions are designed to prevent transmission of microorganisms by common routes in hospitals. It involves the isolation of infectious cases in special hospitals and patients with infected wounds and joint transplantation patients in specific rooms. The following are important considerations regarding hygiene in health-care settings:

1.4.1 Handwashing

Handwashing is frequently the single most important act which reduces the risks of transmitting skin microorganisms between people or from one site to another on the same patient (Hugonnet et al., 2002). Prompt, thorough washing of hands between patient contacts and after contact with blood, body fluids, secretions, excretions, and equipment or articles contaminated by them is a major component of infection control and isolation (Katz, 2004). The spread of nosocomial infections, among immunocompromised patients is linked to health care workers' hand contamination in nearly half of all hands requires correct hand-hygiene procedures flora. The first relates to microbes picked up by workers from the environment which can survive on human skin and sometimes to grow (Katz, 2004). The second group is represented by the permanent microbes living on the skin surface which have low pathogenicity and infection rate, and can prevent colonization by more pathogenic bacteria; microbes making up the resident flora include: Staphylococcus epidermidis, S. hominis, and Micrococcus,

Propionibacterium, Corynebacterium, Dermobacterium, and *Pitosporum spp.*, while transient organisms comprise, *S. aureus*, and *Klebsiella pneumoniae*, and *Acinetobacter, Enterobacter* and *Candida* spp. Hand hygiene is aimed at eliminating the transient flora using a careful and dedicated hand washing with different kinds of soap, (normal and antiseptic), and alcohol-based gels. The lack of available sinks and time-consuming performance of hand washing presents problems related to its effectiveness (Langley, 2002).

1.4.2 Use of gloves

As well as hand washing, gloves play an major role in reducing the risks of microbial transmission. Gloves are worn to reduce the chance that microorganisms present on the hands of personnel can be transmitted to patients during invasive or other patient-care procedures and are also worn to limit the likelihood that the hands of personnel contaminated with microbes from a patient transmits the to another patient. Gloves must be changed between patient contacts, and hands should be washed after glove removal.

The wearing of gloves does not replace the need for handwashing, because gloves may have undetectable defects and hands can also become contaminated during glove removal; failure to change gloves between patient contacts is an obvious infection control hazard.

1.4.3 Antimicrobial surfaces

Microbes can survive on inanimate 'touch' surfaces for long periods, a fact which is troublesome in hospital environments in which patients with immunodeficiency are at increased risk of contracting nosocomial infections.

Common touch surfaces include bed rails, call buttons, touch plates, chairs, door handles, light switches, grab rails, intravenous poles, dispensers (alcohol gel, paper towel, soap), dressing trolleys, and counter and table tops are known to be contaminated with *Staphylococcus*, MRSA and vancomycin resistant *Enterococcus* (VRE) (Chemaly *et al.*, 2014). A number of compounds can

decrease the risk of bacteria growing on surfaces including: copper, silver, and germicides such as hydrogen peroxide vapour (Otter and French, 2009).

Nosocomial infections in the blood can cause serious medical problem and can result in major health care expenditure and death. The onset of infections in the bloodstream can be exacerbated by the previous use of antibiotics, corticosteroids, and a variety of chemotherapeutic agents. Major medical problems such cancers; neutropenia are also associated with increased problems from blood infections as is surgery and haemodialysis (Khan *et al.*, 2015).

The hospital environment is a potential reservoir of bacterial pathogens which are able to infect patients with a diverse variety of pathogenic microbes and thereby expose a large number of susceptible individuals to significant morbidity and mortality. Bacterial pathogens have an innate ability to survive for long periods on surfaces in the hospital environment (Dancer, 2009). Bacterial pathogens, isolated from hospital environment, are also beginning to develop resistance to multiple antimicrobial agents; these then cause difficulty in the treatment of nosocomial infections. The environment of patients is, as a result, heavily contaminated by infectious multidrug resistant organisms including methicillin-resistant *S. aureus* (MRSA), vancomycin-resistant Enterococci (VRE), *Clostridium difficile, Acinetobacter baumannii* and *Pseudomonas aeruginosa*, all of which are emerging to produce a major problem for healthcare systems (Dancer, 2009).

Infections in hospitals are, in the main, caused by factors such as crosscontamination between medical personnel, patients and visitors. Hospital surfaces

and equipment can also act as major reservoirs and sources of such cross infection (Cesar-Pastor et al., 2012). Ubiquitous pathogens such as bacteria (Escherichia coli, Enterococcus spp, Acinetobacter spp, species of Pseudomonas and Staphylococcus aureus), viruses (e.g. noroviruses) and fungi (notably Candida) are particularly troublesome in the hospital environment because of the way they can survive for long periods on surfaces or humans (Thomas et al., 2004). The uniforms of medical staff are often contaminated with multi-resistant Staphylococcus aureus and thereby provide a source of cross contamination (Boyce et al., 1997). Air conditioning systems also act as source of airborne fungi such as species of Aspergillus, which can cause lung problems, especially amongst immunocompromised patients (Thomas et al., 2004). Temperature is also a factor in the spread and development of hospital disease, especially in over-heated and under-ventilated environments which are often associated with the hospital environment, notably in winter (Ribera et al., 1994).

1.5 Pathogen transmission

The transmission of contaminated pathogens on environmental surfaces is dependent on many factors including: the ability of pathogens to remain viable on dry surfaces; their ability to remain virulent after environmental exposure; the ability to colonize patients; the ability to transiently colonize the hands of health care workers and finally, the ability to develop resistance to disinfectants used on environmental surfaces (Weber *et al.*, 2010). Bacteria are transmitted by direct

contact between hands, body fluids such as saliva and mucus droplets; dust contaminated by bacteria in the air; via patient carers and via contaminated objects or equipment (Ortega *et al.*, 2010)

1.6 Survival of bacteria

Pathogens are able to survive in the hospital environment for long periods and, in some cases, resist disinfection. The frequency of environmental contamination with multi-resistant bacteria is greater for patients with infected wounds or were supplied with urinary drainage systems. Gastmeier et al. (2006) showed a correlation between survival time and diversity of important nosocomial pathogens. Most Gram-positive bacteria can survive for months on dry surfaces; Enterococcus spp. (including VRE) for example, can survive for 5 days to 4 months, Staphylococcus aureus remains viable for from 7 days to 7 months and Streptococcus pyogens survives for 3 days to 6.5 months. Many Gram-negative species can also survive for several months. Acinetobacter spp. for example, can survive for 3 days to 5 months on environmental surfaces, Escherichia coli persist for 16 months, while *Klebsiella spp.* remain viable for more than 30 months (Kramer et al., 2006). Vancomycin-resistant Enterococcus (VRE) can remain viable on inanimate surfaces from seven days to two-four months (Burke, 2010). The duration of bacterial survival depends on the bacteria concerned and the nature of the contaminated surfaces. For example, Enterococci survive for more than 24h on bed-frames, 18h on cotton, 1h on telephones, 30 min on the diaphragm of a stethoscope, and 1 hour on gloves and hands (Talon, 1999).

Vancomycin-resistant *Enterococcus* have also been recovered from 3 of 10 seat cushions located in a room occupied by a VRE patient.

1.7 Responses of microorganisms to stress

Microbial cells sense stress in order to protect themselves against the deleterious effects of heat (56°C), freeze-thaw injury, chemical or osmotic shock, exposure to reactive oxygen and nitrogen and to antimicrobial peptides or proteins. These responses involve physiological adaptations that counterbalance damage and allow the cells to continue to survive and grow (Griffiths, 2005). Entry into the stationary phase produces a general stress response, resulting in microbial resistance to multiple stresses. For example, when *Escherichia coli* is exposed to a sub-lethal stress, some of the cells die while some can be recovered on growth media. Growth of the recovered cell is dependent on a number of factors, including the level of stress, the nature of the recovery conditions and the growth phase of cell when the stress was imposed (Griffiths, 2005). Stationary phase cells show a greater tolerance to stress than exponentially growing ones. E. coli cells become shorter and rounder, the mode of metabolism alters and changes in membrane and cell wall structure occur (Griffiths, 2005). This response is seen in facultative anaerobes such as Salmonella and Staphylococcus aureus but generally not in strictly fermentative organisms such as Streptococcus mutans. *Campylobacter jejuni* also shows a reduced resistance during the stationary phase (Griffiths, 2005).

1.8 Responses of microorganisms to stress include:

1- Synthesis of protective proteins that repair cell damage, cell maintenance and bring about the eradication of stress agents.

2- Increases in resistance to lethal factors.

3- Transformation of cells to a latent state, e.g. the formation of spores.

4- Evasion of the host's defence mechanisms.

5- The production of adaptive mutations.

1.8.1 Types of microbial stress adaptation

The mechanisms of bacterial defence against environmental conditions are divided into two classes:

1- Limited or specific adaptive response resulting from microbial exposure to physical, chemical or biological stress, which protects cells against the subsequent lethal effects following the same stress (Griffiths, 2005).

2- Multiple adaptive responses (cross-protection) which occur when bacterial cells adapt to an inherent physiological condition or to an environmental factor, which results in microbial protection against subsequent lethal treatments, including stresses to which the microorganism had not been previously exposed. This type of protection is produced by a variety of stress conditions such as cell starvation, exposure to high or low temperatures, high osmolality, and low pH (De Angelis and Gobbetti, 2004).

1.8.2 Starvation-stress response

When *E. coli*, *Salmonella*, and many other microorganisms are starved, they respond by inducing the expression of up to 50 new proteins or pre-existing proteins. The genetic and physiologic reprogramming that occurs is termed the starvation-stress response (SSR). The SSR allows for long term starvation survival of bacteria and provides generalized cross-resistance to a variety of other environmental stresses. In order to protect bacteria from damage, physiologic changes that occur during SSR include; the degradation of cellular RNA, proteins, and fatty acids; the reduction in the number of ribosomes; alterations in the amounts and type of nine lipids in the cytoplasmic membrane; an increase amounts of lipopolysaccharide in the outer membrane of Gram-negative bacteria; and increase the concentration of chromosomal DNA (Griffiths, 2005).

1.9 Epidemiologically significant pathogens

1.9.1 Bacteria

The main reservoirs for MRSA are infected patients and personnel in hospitals, and not surprisingly, the degree of pathogen contamination directly relates to the degree of localised infection in patients and on bandages, urine or blood. Both *Staphylococcus aureus* and MRSA can survive for up to nine weeks

(Popovich et al., 2008). Staphylococcus aureus causes a variety of infections including skin, soft tissue infections, BSIs (blood stream infections), pneumonia, meningitis, endocarditis, and toxic shock syndrome (Popovich et al., 2008). Coagulase-Negative Staphylococci (CoNS) are able to form biofilms on foreign devices located within patients, including prosthetic joints, pacemakers, intravenous catheters and shunts (Cervera et al., 2009). Hidron et al. (2008) found that CoNS are easily the most frequent cause of central line-linked blood system infections (CLABSIs) and the second most common cause of surgical site infections (SSIs). Less commonly, CoNS infections result in catheter associated urinary tract infections (CA-UTIs) and ventilator-associated pneumonias (VAPs) (Martin *et al.*, 1989). These types of infections are now seen as increasingly common their occurrence has increased with the use of inter-body devices. Wisplinghoff *et al.* (2004) concluded that 9% of CLABSIs are caused by Enterococcus species, of which 2% of E. faecalis isolates; 60% of E. faecium isolates were found to be vancomycin resistant. VRE contamination has been found in up to 37% of samples obtained from the environment associated with gowns, health care workers, medical equipment, and microsphere beds. *Clostridium difficile* spores are durable and resistant to unusual cleaning methods and contamination of the hospital environments by spores has been reported. Such spores are highly resistant and may survive for months in the environment. Hospital floors remain contaminated with these spores for up to five months and contamination density is increased by presence of patients infected with diarrhea

(Hidron *et al.*, 2008). The gastrointestinal tract of young people is a reservoir of *C. difficile* which is transmitted via the faecal-oral route, directly by hand carriage by health care workers (HCWs), or by patient-to-patient contact or indirectly from a contaminated environmental source (Burke, 2010). Best *et al.* (2010) sampled the air and environmental surfaces adjacent to patients with symptomatic CDI and found *C. difficile* near a majority of the patients. Clinicians around the world have noted an increase in disease caused by this bacterium in patients with CDIs. Hidron *et al.* (2008) demonstrated that *E. coli* and *P. aeruginosa* are the Gramnegative organisms which are most commonly isolated from health care-associated infections; less commonly isolated organisms include *Klebsiella pneumoniae, Enterobacter* species, *Acinetobacter baumannii*, and *Klebsiella oxytoca*.

1.9.2 Opportunistic mycoses

Opportunistic mycoses affect the skin, mucosa and the internal organs and are caused by both yeast and moulds. Such infections are associated with weakness in the host's immune defences, especially as the result of long-term and severe immunosuppressed. The most important pathogenic fungi are *Candida albicans*, *Aspergillus* spp., *Cryptococcus neoformans*, *Cryptosporidium*, *Malassezia spp., and Saccharomyces cerevisiae* (Badiee and Hashemizadeh, 2014). In addition to *Candida* and other yeasts, infections can also be caused by phaeohyphomycetes and hyalohyphomycetes. These organisms act as primary infection foci, usually affecting the upper or lower respiratory tract and can spread hematogenously and lymphogenously to infect additional organs (Badiee and Hashemizadeh, 2014). At

least 70% of all human *Candida* infections are caused by *C. albicans*, and the rest by *C. parapsilosis*, *C. tropicalis*, *C. guillermondii*, *C. kruzei*. Diba *et al.* (2012) identified pathogenic fungi from the environment including: *Candida albicans*, *C. krusei*, *C. glabrata*, *C. tropicalis* and *C. parapsillosis*. Of the isolates 35(31.5%) were shown to be *Candida* species, 48(43.2%), *Aspergillus* and 28 (20.3%) of other species.

1.10 Microorganisms used throughout the following

Staphylococci aureus is a non-motile small spherical cell (1µm) found in grapelike clusters. It is a Gram- positive coccus, catalase-producing bacteria, facultative anaerobe and can be cultured on normal nutrient media at 37° C. Extracellular enzymes and exotoxins produced by this bacterium, such as coagulase, alphatoxin, leukocidin, exfoliatins, enterotoxins, and toxic shock toxin are responsible for the resultant clinical symptoms and infections. *Staphylococcus aureus* is a frequent pathogen in nosocomial infections and it causes limited outbreaks in hospitals of problems including: furuncles, food poisoning, dermatitis exfoliativa, toxic shock syndrome, carbuncles, wound infections, sinusitis, otitis media, and mastitis puerperalis (Spaulding *et al.*, 2014).

Escherichia coli is a Gram-negative, flagellated rod whose natural habitat is the intestines of animals and humans. This bacterium is a major human pathogen that causes many infections including: lower urinary tract infection such as urethritis,

cystitis, urethrocystitis; infections of the renal pelvis and kidneys (cystopyelitis, pyelonephritis), wound infections, gallbladder and bile ducts infections, peritonitis, meningitis patients. *E. coli* also causes acute urinary tract infections in 70–80% of cases and in chronic, persistent infections in 40–50% of cases, as well as about 15% of all cases of nosocomial sepsis (*S. aureus* 20%).

Candida rugosa is a budding oval yeast Branched pseudohyphae are frequently seen and occasionally septate mycelia, with chains of elongated blastoconidia. *Candida rugose*-pseudohyphae differentiates it from *C. lusitaniae, C. parapsilosis. Candida rugosa* has recently been cited as a possible "emerging" fungal pathogen capable of causing invasive infection in immunocompromised patients especially following the use of catheters but also via other modes of nosocomial acquisition (Pfaller, 2006). Behera *et al.* (2010) identified *Candida rugosa* as a causative pathogen of candidaemia, thirteen out of the 19 patients (68.4%) with C. rugosa candidaemia died.

1.11 Molecular biology techniques

16S rRNA and 18S rRNA analysis has been applied here to identify both bacteria and the yeasts isolates studied here. DNA can be isolated from a variety of specimens including from tissues, blood, bones, sperm, plant, hair and bacteria (Lahiri, 1992). DNA can also be used for laboratory diagnosis and in forensics (Hill *et al.*, 2000). In DNA analysis tissues or cells are first broken and the cells are lysed using detergents or enzymes. This is followed by centrifugation to separate the DNA from other components and by DNA purification (Amann *et al.*, 1995).

1.11.1 Deoxyribonucleic acid (DNA)

Deoxyribonucleic acid (DNA) is the genetic material in living organisms. DNA plays an essential role for storing of the biological information due to its polymeric structure and because there are different nucleotides.

DNA is a polymer; a long, chainlike molecule consists of two long polynucleotide chains made up of four types of nucleotide sub-units called monomers. Each chain is called a "DNA chain" or "DNA strand", and both chains are linked together by hydrogen bonds. The Nucleotides are composed of five carbon sugars which are linked with one or more phosphate groups and nitrogen containing base. The sugar is a deoxyribose attached with a single phosphate and one of the nitrogen bases Adenine (A), cytosine (C), guanine (G) or thymine (T). These nucleotides are attached together in a chain with the sugars and the phosphate groups form a backbone. The nucleotide subunits are linked together in a specific way that gives the DNA molecule chemical polarity. The two ends of the chain will be differing One is a hole3[′] hydroxyl and the other is a knob5[′] phosphate (Madigan *et al.,* 2012).

1.11.2 Polymerase Chain Reaction (PCR) technique

PCR is a powerful method of amplifying specific DNA sequences invented by Kary Mullis in 1983 (Mullis, 1990). The PCR technique used in clinical microbiology and research laboratory for identify the microbial pathogens. It used for detect pathogens which has a slow growth rate, clarify the taxonomic positions of known pathogens and help genotyping for microbial characterization which is become more specific and easily quantified between different organisms (Valones et al., 2009). This technique depends on repeated synthesis of target DNA using DNA polymerase through three different steps under controlled temperatures in many thermal cycles. The steps are: **Denaturation** involves melting the double strand of the target DNA and separate them to single strands under high temperature 90-98°C for 1 minute. Annealing which allows the two added primers (oligodeoxyribonucletides) to anneal with the separated DNA to be amplified. The primers anneal to the opposite side of the DNA strands in their 3' end under low temperature 37-65°C for 45 seconds. In the **extension**, the DNA polymerase synthesis a new DNA strand which is complementary to the target DNA by adding the deoxynucletides (dNTPs) from the reaction mixture of the 5'-to-3' direction under optimum temperature for DNA polymerase 75–80°C for 2 minutes. These steps run in a single cycle so many cycles are needed to amplify the specific DNA to millions of copies. PCR reaction set of 30 cycles. Final elongation: this step is optional and requires a temperature of 70-74°C for 5-15 minutes after the final step of PCR to ensure that all single strand of DNA is elongated completely. Final

hold: in this step, the reaction mixture is cooled at 4-15°C for a short time to allow the products to be stored.

The aim of the work described here was to:

- Studies the distribution of bacteria in the environment and their survival in the healthcare settings including: sinks, lift buttons, computer keyboards and computer mice, mobile telephones, mirrors in toilets, dust obtained from vacuum cleaners, under the surfaces of shoes, books and shelves in libraries and upper surfaces of water taps.
- 2) Determine of the number of bacteria on hands after washing and drying normally and with warm air dryer and quantify of bacteria transferred from hand warm air dryers
- 3) Determine the factors which influence the survival of bacteria and to a lesser extent *Candida* in the built environment on ceramic tiles, on copper and plastic plumbing surfaces, and on new toothbrushes.
- Isolation pathogenic bacteria from used toothbrushes and determine the effect of toothpastes on growth of pathogenic bacteria.
- 5) Determine of effectiveness of antibacterial cloths in inhibiting the growth of bacteria and yeast.
- 6) Determine the survival and relevant metabolic diversity of bacterial isolates.

CHAPTER 2

THE DISTRIBUTION OF BACTERIA IN THE ENVIRONMENT AND THEIR SURVIVAL IN HEALTHCARE SETTINGS

2.1. Isolation of bacteria from sinks, toilets and other medical environments

Hospitals and other health care settings act as an obvious potential reservoir for pathogens (Dancer, 2009). (Bauer *et al.*, 1990, Kayabas *et al.*, 2008, Medina *et al.*, 1997, Sehulster *et al.*, 2003). Contaminated hand washing sinks are an obvious source of infection, especially where they are used for disposing of body fluids, where they become a focus for the survival of pathogens and especially biofilms; surprisingly perhaps, healthcare workers can become contaminated when washing their hands in a contaminated sink (Roux *et. al.*, 2013).

2.2. Isolation of bacteria from sinks

2.2.1. Materials and Methods

1) Isolation of samples and collection

Sterile cotton swabs wetted by dipping in normal saline were used to collect samples from sinks. All samples were labelled and streaked on Nutrient Agar (Oxoid) plates, followed by incubation at 25°C under aerobic conditions for 24hrs. After incubation the colonies were identified. The sinks examined were located as follows: Sheffield University: Firth Court Building, Disability and dyslexia support service, The Alfred Denny Building, Information Commons Building,
Students Union Building. Other areas in Sheffield: Sheffield Train Station, local supermarket, local hospital, various private dwellings.

2) Purification of isolates

All samples were isolated from sinks and then streaked on media in Petri dishes. The main medium used was Nutrient Agar, incubation was then at 37°C for 48 hours. The isolates were purified to single colonies and subjected to molecular identification.

3) Identification of bacterial isolates using 16s rRNA technique

A bacterial suspension in LB medium was prepared and incubated overnight at 37° C. After the incubation period, 1-3 ml of media was transferred to a sterile Eppendorf tube and centrifuged at 6000xg for 2 min at room temperature and the supernatant was decanted completely. A KeyPrep bacterial DNA extraction kit supplied by ANACHEM® was used and all steps were conducted as described in the instructions provided by the Company. Buffer (100µl) R1 was added to the pellet and the cells were re-suspended completely by pipetting up and down. After full cell homogenising, 20 µl of lysosyme was added, mixed and incubated at 37° C for 20 min. The mixture was then digested by centrifugation at 10,000xg for 3 min and the supernatant was completely decanted. The pellet was then re-suspended in 180 µl of buffer R2 and 20 µl of proteinase K was added and incubated at 65° C for 20 min in a water bath with occasional mixing every 5 min.

400 μ l of buffer. BG was added and mixed thoroughly by inverting the tube several times until a homogeneous solution was obtained and then incubated for 10 min at 65^o C. After the incubation period, 200 μ l of absolute ethanol was added and mixed thoroughly.

The sample was next transferred into a column assembled in a clean collection tube and centrifuged at 10,000 xg for 1 min while the flow was discarded. The column was washed by addition of 750 μ l of wash buffer and centrifuged at 10,000 xg for 1 min while the flow was discarded. Finally, the column was placed in a clean micro-centrifuge tube and 70 μ l of elution buffer was added and centrifuged at 10,000xg for 2 min to elute DNA. DNA was stored at -20^oC until the next step.

4) Gel electrophoresis

Gel electrophoresis was conducted to ensure that the bacterial DNA was wellextracted and purified. The following steps were used.

4.1 Agarose preparation

Powdered agarose was weighed carefully (0.5 g) into a conical flask and 50 ml of 1x TAE (Tris Acetate EDTA) buffer was added with 40ml of distilled water. The contents of the flask were then mixed and placed in a microwave plate until the contents just began to boil and all the powdered agarose is melted. After the contents cooled to 50-55 °C, ethidium bromide solution was added to give a final concentration of 5 pg/mL. The mixture was then be poured into a gel tray and left

to solidify. The gel was finally placed in an electrophoresis tank and submerged in 1x TAE buffer to ensure that the sample diffused into the wells.

4.2 Sample loading

DNA samples (10 μ l) were prepared by the addition of 5X (2 μ l) loading dye (blue or orange) to the samples and loaded into the wells of the gel. All samples were loaded at the same time. 6 μ l of hyper ladder was then added into an adjacent well as a reference. The voltage was set to the desired level at 80V for 40 minutes to initiate electrophoresis. The leads were then attached allowing DNA to migrate within the gel toward the anode. After electrophoresis the gel was removed from try, and the DNA fragment was viewed on a UV transilluminator; images were then captured using a connected digital camera.

4.3. Sample amplification

The Polymerase Chain Reaction (PCR) technique was used. A mixture in a sterile Eppendorf tube was prepared as follows: $12 \ \mu$ l of master mix, $1 \ \mu$ l of forward primer, $1 \ \mu$ l of reverse primer, $1 \ \mu$ l of the DNA sample, and $35 \ \mu$ l of sterile distilled water. The mixture was inserted in a PCR machine and the programme was adjusted as follows:

Time Number of Steps Temperature cycles (Min) Initialization (Initial 94°C 3 1 denature) 94°C 1 Denature 35 60°C 1 35 Annealing Extension/Elongation 72°C 35 1 Final elongation 72°C 5 1 Hold 4°C 1

Table 2.1 The Polymerase Chain Reaction (PCR) cycle.

5) 16S rRNA sequencing and phylogenetic analysis

After PCR, aliquots of 10 µl of each sample were allowed with 1µl of forward primer and 1µl of reverse primer in a sterile small size tube and sent to the Medical School Core Genetics Unit (University of Sheffield) to be sequenced. 16S rRNA gene sequences were adapted using the Finch TV software and then exported into the Basic Local Alignment Search Tool (BLAST), available from the website of the National Centre for Biotechnology Information (NCBI), to identify matches with existing characterized reference sequences. Partial sequences, generated in this experiment, were assembled and the errors of consensus sequences were corrected manually by using Finch TV software (version 1.4). In Finch TV software, the unknown nucleotide is represented as N, and it could be either A, or T, or G, or C, according to the different colours which appear (Mishra *et al.*, 2010).

2.2.2. Results

1) Isolation of bacteria from various sinks by cultivation on Nutrient Agar medium

Bacteria were isolated using Nutrient Agar which facilitates the rapid isolation of bacteria from mixed cultures and used for biochemical or serological tests. This medium is a basic culture medium used for maintaining microorganisms, cultivating fastidious organisms by enriching with serum or blood and are also used for purity checking prior to biochemical or serological testing. The presence of peptic digest of animal tissue, beef extract and yeast extract provides the necessary nitrogen compounds, carbon, vitamins and also some trace ingredients necessary for the growth of bacteria; sodium chloride maintains the osmotic equilibrium of the medium. The following bacteria were isolated from the surface of ceramic sinks.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.1) in order to confirm that they were bacteria



Kocuria rhizophila Gram stained. Magnification: 100x.



Micrococcaceae Bacterium Gram stained. Magnification: 100x.



Klebsiella oxytoca Gram stained. Magnification: 100x.



Bacillus subtilis Gram stained. Magnification: 100x.



Bacillus cereus Gram stained. Magnification: 100x.

Figure 2.1: Nutrient Agar plates showing cultured bacteria isolated from various sinks, also microscopy images shows the bacteria under the light microscope.

Table 2.2: Bacteria isolated from various sinks.

Representative	Closest matches	Sequence	NBCI (Accession
sequence	identification	identity	number)
S 1	Klebsiella oxytoca	100%	CP011618.1
S2	Bacillus subtilis	100%	KP340123.1
S4	Kocuria rhizophila	99%	KM978822.1
\$5	Bacillus cereus	99%	KC731425.1
\$6	Micrococcus luteus	99%	KT339390.1

2.2.3. Discussion

The bacteria shown in Table 2.2 were isolated from chosen sinks which are in general use and so represent the environments typical of health care settings. Five bacteria were isolated and identified, two of which were species of *Bacillus*.

Klebsiella oxytoca

Klebsiella is commonly isolated in the clinical laboratory from specimens of blood, urine and respiratory secretions. *Klebsiella* are non-motile, Gram-negative bacilli, glucose fermenters, and form red colonies on MacConkey agar; negative results for cytochrome oxidase activity, hydrogen sulfide production in triple sugar iron agar, arginine and ornithine, decarboxylation, phenylalanine deamination, and citrate utilization. *Klebsiella oxytoca* is a diazotroph, which colonises plants and

fixes atmospheric nitrogen in, for example, the barley rhizosphere. This organism causes colitis and sepsis (Hogenauer *et al.*, 2006).

Bacillus cereus

B. cereus group are Gram-positive, rod-shaped, spore formers, motile. *Bacillus* cereus is the most frequently isolated of *Bacillus* spp involved in nosocomial infections. It has been described an outbreak of invasive *B. cereus* infections in neonatal intensive care units. Kalpoe et al (2008) reported an outbreak of vancomycin resistant B. cereus respiratory tract colonization in patients in pediatric intensive care units. Bacillus cereus is also common in bloodstream infections in patients with hematologic malignancies due to contamination of reused towels. Bacillus cereus is a highly important eye pathogen because it can cause blindness rapidly; endophthalmitis associated with this bacterium often results in significant vision impairment. This bacterium is the most known foodborne pathogen associated with food-poisoning. It has also been found to be associated with fulminant pneumonia in immunocompetent patients and with oral diseases; meningoencephalitis and meningitis in infants, neonates and immunocompromised individuals; necrotizing fasciitis and myositis (Liu, 2011).

Bacillus subtilis

Bacillus species, including *B. subtilis*, can form endospores which allow them to survive heat treatment and disinfection procedures and as a result, present serious health problems as contaminants of food or in the general hospital settings as a cause of nosocomial infections, especially in immunocompromised patients. These bacteria produce toxins which have been linked as the causative agent of several human diseases including endophthalmitis, inflammation of intraocular tissues or fluids due to intraocular infection (Liu, 2011). They also contribute to foodborne infections by producing toxins with a cereulide-like mode of action (e.g. in oral disease, mainly gingivitis and periodontitis). Cryptic pyogenic infections of the central nervous system (CNS) following dental affections have also been implicated to *B. subtilis* and *B. circulans* (Liu, 2011).

Kocuria rhizophila

Kocuria rhizophila cells are Gram- positive, non-acid-fast, non-motile, nonendospore forming, aerobic, and occur in pairs, tetrads, and packets. *Kocuria rhizophila* is widely distributed in the natural environments such as soil, the rhizosphere, freshwater, marine sediments, and fermented foods. *Kocuria* spp are also found in the skin, mucosae, and oropharynx of humans as commensals, although several of them have been associated with human diseases (Liu, 2011). Becker *et al.*, (2008) reported a case of *Kocuria rhizophila* infection in a boy with methylmalonic aciduria. The patient developed sepsis and showed symptoms of acute pancreatitis and fever. A *Kocuria rhizophila* strain was also isolated from blood samples drawn through a port system and from peripheral veins during septic episodes. A number of *Kocuria* species are opportunistic human pathogens associated with catheter-related bacteraemia, pneumonia, intracranial abscesses, septic arthritis, meningitis, peritonitis, and endocarditis in immunocompromised patients (Liu, 2011).

Micrococcus luteus

Members of the genus *Micrococcus* are Gram-positive cocci, aerobic, non-motile, and non-endospore forming. *Micrococcus* spp. are commonly found as members of the normal skin flora of humans and other mammals. *Micrococcus* strains such as *M. luteus* are opportunistic human pathogens associated with catheter-related bacteraemia in patients undergoing heamodialysis or leukaemia treatment, pneumonia, endocarditis patients particularly immunocompromised individuals, intracranial abscesses, continuous ambulatory dialysis peritonitis, septic arthritis, and meningitis (Liu, 2011).

The results show that sinks are clearly a potential source of pathogens, both in the domestic and healthcare settings.

Hospital sinks are considered to be one of the most frequently implicatedreservoirs for MDR Gram-negative bacilli, including MDR coliforms (Roux *et al.*,2013). *K. pneumoniae* strains which have the ability to survive for long periods

within plumbing components are also more likely to contain bacteria exhibiting extended-spectrum β -lactamases (Yang and Zhang, 2008). Removal and replacement of the sink and associated pipe work together with upgrading practices for sink use and cleaning is often required to end an outbreak. Outbreaks of MDR *Klebsiella* are also often associated with the bad practise of tipping patient fluids down the nearest available sink instead of removing clinical waste to the designated waste sluice located at a distance (Roux et al., 2013). Not surprisingly, lower rates of sink contamination are significantly linked to daily bleach disinfection, in addition to restrictions being placed on sink use for hand washing only and not the routine disposal of fluid or clinical waste (Roux et al., 2013, Yang and Zhang, 2008). It is noteworthy in the finding reported in this Thesis that the bacteria isolated from sinks were essentially environmental species and not those capable of causing major life-threatening infections. It needs to be emphasised, however, that even these bacteria can cause major problems in immunocompromised patients.

2.3 Isolation of bacteria from lift buttons surfaces

2.3.1 Methods

Sterile cotton swabs wetted by dipping in normal saline were used to collect samples from environmental surfaces such as lifts buttons from different floors in the MBB building and Sheffield University Students' Union. All samples were labelled and streaked on to Nutrient Agar Plates which were incubated at 25°C under aerobic conditions for 24hrs. After incubation, the colonies were identified. Genomic DNA was extracted from each isolate and was identified using 16S rRNA. The extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit (supplied by ANACHEM). The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.3.2 Results and Discussion

1) Isolation of bacteria from various lift buttons by cultivation on Nutrient Agar medium

Bacteria were cultured using Nutrient Agar having been isolated from lift buttons. The bacteria shown in (Figure 2.2)



Figure 2.2: Bacteria contaminating lift buttons. From left to right, shows ground floor to upper floors from various locations.

Representative	Closest matches	Sequence	NCBI (Accession

Table 2.3. Bacteria isolated bacteria from lift buttons

sequence	identification	identity	number)
6W3	Staphylococcus warneri BCL-34	99%	KT582294.1
7W1	Staphylococcus epidermidis	98%	FR797804.1
7Y3	Micrococcus luteus MBS022	99%	KM378607.1

Staphylococcus

The characteristics of these bacteria include:

Staphylococcus is a genus of catalase positive Gram-positive bacteria.

Staphylococcus bacteria appear as round (cocci) and come together as grape-like clusters. *Staphylococci* consist of >40 species and are noted for their ability to cause disease, with *Staphylococcus aureus* being a most common pathogen of man and animals. Most *Staphylococcus* species are however, harmless and normally live on the skin and mucous membranes of humans and other organisms (Liu, 2011).

Staphylococcus aureus is mainly a commensal organism residing on the human epithelia of the mucosae and skin. It has its primary ecological site in the vestibulum nasi, located in the forefront region of the nose. None-nasal sites typically harbouring the organism include the skin, perineum, and pharynx; other

sites include the gastrointestinal tract, vaginas, and axillae. This bacterium causes superficial skin lesions like impetigo, styes and furuncules, cellulitis folliculitis, carbuncles, staphylococcal scalded skin syndrome (SSSS), and abscesses (Liu, 2011). More serious infections include pneumonia, mastitis, phlebitis, meningitis, urinary tract infections (UTIs), and septicaemia. *Staphylococcus aureus* infections including osteomyelitis, hospital-acquired (nosocomial) infections in surgical wounds and infections associated with indwelling medical devices. *S. aureus* also causes food poisoning by producing enterotoxins and toxic shock syndrome (TSS) by release of superantigens into the blood stream (Liu, 2011).

Staphylococcus epidermidis is a skin commensal which can be isolated from mucous membranes, such as the groin or axilla, exposed skin surfaces and saliva. It is the main human pathogen in intravascular catheter-related infections, nosocomial bacteraemia, endocarditis, urinary tract and surgical wounds infections and infections of the central nervous system, ophthalmologic infections, peritoneal dialysis-related infections and infections of prosthetic joints (Liu, 2011). Some isolates produce slime or biofilm which is the major virulence factor of *S. epidermidis*, enabling colonization and persistence on prosthetic material, the resistance to the effects of antibiotics, and the ability to evade the immune system (Liu, 2011).

Micrococcus are Gram-positive, non-motile, non-endospore forming, aerobic cocci. *Micrococcus luteus* have been associated with catheter-related bacteraemia in patients undergoing haemodialysis or leukaemia treatment, pneumonia,

endocarditis, intracranial abscesses, continuous ambulatory dialysis peritonitis, septic arthritis, and meningitis (Liu, 2011).

The number observation of bacterial contamination of lift buttons decreased from ground floor towards upper floors as shown in Figure 2.2. These results agree with the findings of Al-Ghamdi *et al.* (2011) who showed that the average rate of bacterial contamination of different objects within the hospital environment was 95.5% with elevator buttons showing the highest percentage (97%). Scott and Bloomfield (1990) suggested that when contaminated surfaces are contacted with the fingers, a significant number of organisms can be transferred which can be subsequently recovered on an agar surface. Inanimate objects can play a role in the transmission of human pathogens. These surfaces have been shown to carry both non-pathogenic and pathogenic bacteria and even a single hand contact with a contaminated surface results in a variable degree of pathogen transfer.

2.4 Isolation of bacteria from computer keyboards and computer mice

Multidrug-resistant strains of *S. aureus*, particularly methicillin resistant *S. aureus* (MRSA), present a major clinical and epidemiological problem in healthcare settings as they are easily transferred among hospital staff and patients, especially in intensive care units (ICUs). Hails *et al.* (2003) found that at least 16% of patients were colonised with MRSA. A significant factor contributing to the transmission of microorganisms is their ability to survive on environmental

surfaces. Also the microbial contamination of environmental surfaces in the hospital may contribute to the spread of potential pathogens without direct patient contact. Coagulase-negative Staphylococci have been cultured from all keyboards investigated in different studies conducted in the USA (Anastasiades *et al.*, 2009). In another study which focused on the isolation of MRSA, 65% of nurses, who had contact with MRSA-infected patients, bacteria contaminated their uniforms as well as the keyboards and computer mice (CM) in the hospital wards. These results confirmed that inanimate objects can serve as reservoirs for bacteria. Hartmann *et al.* (2004) also found that keyboards and CM might serve as a source for the transmission of microorganisms.

In a study investigating the presence of both Gram-positive and Gram-negative bacteria on computer keyboards, coagulase-negative staphylococci and *S. aureus* were isolated from 100% and 4% of keyboards, respectively (Rutala *et al.*, 2006). It was suggested that the use of plastic keyboard covers could reduce contamination. It was then recommended that the same infection prevention measures employed during direct contact with patients (i.e. hand washing and use of gloves), should be enforced when handling computer hardware in healthcare settings.

As has been mentioned in Chapter 1, medical surfaces present a major source of outbreaks of community-acquired and nosocomial infections. Such surfaces include frequently used computer keyboards and accompanying mice which

harbour skin and dust bacteria, such as species of *Staphylococcus* and *Streptococcus epidermidis* and fungi (yeasts), notably *Candida albicans*. Keyboards were also implicated in burns unit-associated nosocomial infections by *A. baumannii* (Neely *et al.*, 1999) MRSA and *Enterobacter* spp. (Bures *et al.*, 2000; Goldman, 2000). It goes without saying that hospital keyboards and computer mice should be routinely cleaned and covered with transparent plastic covers when not in use. Hand hygiene is also necessary to avoid cross contamination of keyboards and other computing devices. This is especially important since contaminated personal computers are often contaminated with Staphylococci and *Pseudomonas* spp. And can transmit MRSA (Isaacs *et al.*, 1998).

2.4.1 Materials and Methods

All samples are isolated from computer keyboards using swabs (70 samples) streaked on petri dishes containing Nutrient Agar and incubated at 25°C for 48 hours. Genomic DNA extracted from each isolate was identified using 16S rRNA. The extraction of genomic DNA by using (KeyPrep bacterial DNA extraction kit supplied by ANACHEM), The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.4.2 Results

1) Isolation of bacteria from computer keyboard

Bacteria were cultured using Nutrient Agar after isolation from computer

keyboards and mice.

2) Light microscope images. The isolates were examined under a light

microscope (Figure 2.3).



Bacillus amyloliquefaciens Gram stained. Magnification: 100x.



Bacillus subtilis Gram stained. Magnification: 100x.



Brevibacillus brostelensis Gram stained. Magnification: 100x.



Bacillus cereus Gram stained. Magnification: 100x.



Pantoea caldia Gram stained. Magnification: 100x.



Staphylococcus epidermidis Gram stained. Magnification: 100x.

Figure 2.3: Nutrient Agar plates showing cultured bacteria isolated from various computer keyboards, also microscopy images showing the bacteria under the light microscope.

Representative	Closest matches	Sequence	NBCI (Accession
sequence	identification	identity	number)
KB1	Bacillus subtilis	99%	KJ746466.1
KB4	Bacillus amyloliquefaciens	99%	AB301004.1
KB6	Brevibacillus brostelensis	98%	EU816699.1
KB11	Staphylococcus epidermidis	99%	KF575163.1
KB13	Pantoea caldia	99%	AB907785.1
KB14	Bacillus cereus	99%	DQ923480.1
KB21	Pseudomonas luteola	99%	KC429633.1

Table 2.4	Bacteria	isolated	from	various	computer	keyboards.
	Daeterra	10014004		10000	eomparer	nej couras.

Bacillus amyloliquefaciens

This bacterium typically presents as motile, Gram-positive rods, often forming chains, with peritrichous flagella. Optimal temperature for growth is 30 to 40°C, while no growth occurs below 15°C or above 50°C. *B. amyloliquefaciens* can be isolated from soil and brackish sediments (Priest *et al.*, 1987). *B. amyloliquefaciens* is not known to produce any mammalian toxins, and is not associated with food-borne disease. No problems have been reported, and none are expected from exposure to *B. amyloliquefaciens* via drinking water (Priest *et al.*, 1987).

Brevibacillus brostelensis

Brevibacillus brostelensis cells present as Gram-positive rods, motile with peritrichous flagella. It is a thermophilic, spore-forming rod with a growth optimum at 50°C. Colonies are flat, smooth, circular, and entire. This bacterium produces a soluble brown-red pigment(s) on nutrient agar and is strictly aerobic. The habitat of *Brevibacillus* overlaps with that of *Bacillus* and is associated with soils and dairy environments (Sanders, 2003; Shida *et al.*, 1995).

Pseudomonas luteola

Pseudomonads appear as straight or slightly curved Gram-negative rods. They are usually easily differentiated from Enterobacteriaceae by a lack of a bipolar

staining. Isolates often seen as clusters of rods encapsulated in a thick pink staining alginate coat. *Pseudomonas* spp. grow on several different nonselective agars, including nutrient, 5% blood, chocolate, and MacConkey agars (Liu, 2011). Most species are aerobic and grow at 37°C, and within 24h, form visible colonies. This bacterium is oxidase negative and produces a non-diffusible yellow pigment. *Pseudomonas* spp. act as opportunistic pathogens and *P. luteola* can cause community- or hospital acquired infection in humans especially patients in an intensive care unit. *Pseudomonas luteola* causes dialysis associated peritonitis; cellulitis; sepsis; prosthetic valve endocarditis; and postoperative endophthalmitis (Liu, 2011).

Pantoea calida

Pantoea calida cells are Gram-negative rods that are facultatively anaerobic and motile. Colonies are non-pigmented and convex, oxidase-negative and catalase-positive (Fritz *et al.*, 2014). *Pantoea calida*, a recently described environmental Enterobacteriaceae organism, has yet to be associated with human infection, although Fritz *et al.*, (2014) isolated this organism from a patient with postoperative meningitis.

2.5. Isolation of bacteria from mobile telephones

2.5.1 Materials and Methods

Isolates were obtained from a variety of mobile telephones (30 samples) using nutrient agar medium. Incubation was at 37°C for 48 hours and the isolates were identified using 16S rRNA. Extraction of genomic DNA was by using Norgen's Fungi/Yeast Genomic DNA Isolation Kit. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.5.2 Results

1) Isolation of bacteria from mobile phones

Bacteria were cultured using Nutrient Agar after isolation from mobile phones.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.4).



Staphylococcus epidermidis Gram stained. Magnification: 100x.



Staphylococcus warneri Gram stained. Magnification: 100x.



Bacillus cereus Gram stained. Magnification: 100x.



Bacillus subtilis Gram stained. Magnification: 100x.

Figure 2.4: Nutrient Agar plates showing cultured bacteria isolated from various mobile phones, and also microscopy images showing the bacteria under the light microscope.

Representative	Closest matches	Sequence	NBCI (Accession
sequence	identification	identity	number)
3P	Staphylococcus	99%	K1398217 1
51	epidermidis	<i>y y</i> / 0	13370217.1
15P1	Staphylococcus warneri	99%	KP771665.1
15P2	Bacillus subtilis	99%	HG764646.1
18P	Bacillus cereus	99%	KJ612539.1

2) Table 2.5: Bacteria isolated from various mobile phones.

Staphylococcus warneri a Gram-positive member of the microbiota,

normally found on the skin of humans and animals. *S. warneri* is a significant opportunistic nosocomial pathogen causing complications to the use of central venous catheters, prosthetic heart valves and joints, and neurosurgical ventricular shunts (Incani *et al.*, 2010). *Staphylococcus warneri* has been associated with bacteraemia in hospitalized patients; with intravascular catheter infections in immunocompromised patients (Incani *et al.*, 2010).

2.6 Isolation of bacteria from mirrors in toilets

2.6.1 Materials and Methods

Isolates were obtained from a variety of toilet mirror surfaces (30 samples) using Nutrient Agar medium. Incubation was at 25°C for 48 hours and the isolates were

identified using 16S rRNA. Extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.6.2 Results

1) Isolation of bacteria from mirrors

Bacteria were cultured using Nutrient Agar after isolation from mirror surfaces.

2) Light Microscope images. The isolates were examined under a light microscope (Figure 2.5).



Bacillus cereus KP100400.1 Gram stained. Magnification: 100x.



Staphylococcus haemolyticus Gram stained. Magnification: 100x.



Bacillus cereus Q659737.1 Gram stained. Magnification: 100x.



Bacillus subtilis DQ683077.1 Gram stained. Magnification: 100x.

Figure 2.5: Nutrient Agar plates showing cultured bacteria isolated from various mirror surfaces, and also microscopy images showing the bacteria under the light microscope.

Representative	Closest matches	Sequence	NBCI (Accession
Representative	Closest materies	bequeillee	
sequence	identification	identity	number)
M8-8	Racillus corous	99%	KP100400-1
1010-0	Bacillus cereus	<i>))/</i> 0	Ki 100400.1
M6-1	Racillus subtilis	99%	DO683077 1
	Ducillus subtilis	<i>JJ</i> /0	DQ003077.1
M15-1	Bacillus cereus	99%	IO659737 1
		7770	02003707.1
	Staphylococcus		
M14 1		000/	VC120455 1
10114-1		99%	KC139433.1
	haemolvticus		

Table 2.6: Bacteria	isolated	from	mirrors.
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Staphylococcus haemolyticus is a major cause of human disease in humans and has been implicated in native valve endocarditis (NVE), septicaemia, urinary tract infections, peritonitis, and wound, bone, and joint infections. It forms biofilms which allow it to colonize and persist on prosthetic material where it shows resistance to antibiotics and is able to evade the immune system (Liu, 2011).

2.7 Isolation of bacteria from vacuum cleaner dust samples obtained from carpets, textiles and upholstered furniture

2.7.1 Materials and methods

Dust samples were obtained from a variety of different vacuum cleaners (30 samples) for use in determining the bacterial content of the waste. The dust was placed on the surface of Nutrient Agar medium plates (NA) and then incubated at 25°C for 3 to 5 days. Extraction of genomic DNA was achieved using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.7.2 Results and Discussion

1) Isolation of bacteria from dust samples of various vacuum cleaners

Bacteria were cultured using Nutrient Agar which isolated from dust samples.

2) Light microscope images

In order to confirm that they were bacteria, the isolates were examined under a light microscope (Figure 2.6).



Bacillus thuringiensis FJ174596.1 Gram stained. Magnification: 100x.



Bacillus mycoides KR088435.1 Gram stained. Magnification: 100x.



Bacillus licheniformis DQ071560.1 Gram stained. Magnification: 100x.



Bacillus subtilis KF220577.1 Gram stained. Magnification: 100x.

Figure 2.6: Nutrient Agar plates showing cultured bacteria isolated from dust samples of various vacuum cleaners, and also microscopy images showing the bacteria under the light microscope.

Representative	Closest matches	Sequence	NBCI (Accession
sequence	identification	identity	number)
VC1	Bacillus thuringiensis	98%	FJ174596.1
VC2	Bacillus mycoides	99%	KR088435.1
VC3	Bacillus licheniformis	99%	DQ071560.1
VC4	Bacillus subtilis	98%	KF220577.1

Table 2.7: Bacteria isolated from dust samples of various vacuum cleaners.

Bacillus licheniformis

B. licheniformis has been shown to be the cause of bacteremic episodes in patients having hemotological malignancies. Only one case has been reported of *B. licheniformis* nosocomial infection in neonates. In immunocompetent individuals, *B. licheniformis* bacteraemia has been reported in association with central venous catheters and with peritonitis in patients undergoing peritoneal dialysis (Liu, 2011). *B. licheniformis* has also been shown to be involved in foodborne illnesses due to its ability to produce toxins with a cereulide-action mode. Infections caused by this bacterium include, ophthalmitis, septicaemia, bacteraemia, and peritonitis. *Bacillus licheniformis* was isolated from patients suffering from eye trauma, lymphoma, leukaemia and metastatic lung cancer (Liu, 2011).

Bacillus thuringiensis

Bacillus thuringiensis is an insect pathogen widely used as a biopesticide. Kuroki *et al.* (2009) demonstrated *B. thuringiensis* species can form biofilms in nosocomial bacteraemia due to catheter infection. There is also a report of fatal *B. thuringiensis* bacteraemia in a neutropenic patient suffering from severe pulmonary disease showing that this entomopathogenic bacterium may be an opportunistic pathogen in immunocompromised patients; it is also the causative agents of infectious endophthalmitis and oral diseases, particularly gingivitis and periodontitis (Kuroki *et al.*, 2009).

Bacillus mycoides

Bacillus mycoides is typically isolated from soil and plant rhizosphere but has also been shown to be a causative agent of infectious endophthalmitis (Liu, 2011).

2.8 Isolation of bacteria from soles of shoes

An obvious potential infection hazard is presented by the transfer of microorganisms from the soles of shoes to floors and other surfaces within health care environments and pathogens can be carried in this way from environmental sources, notably soils and cat and dog faeces. In order to prevent the contamination, shoe-cover-alls should be provided for all persons entering sensitive medical areas, such as where immunocompromised patients are being treated.

2.8 Isolation of bacteria from the under surface of shoes

2.8.1 Materials and Methods

Isolates were obtained from a variety of shoe surfaces (30 samples) using Nutrient Agar medium; incubation was at 37°C for 48 hours and the isolates were identified using 16S rRNA. Extraction of genomic DNA by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.8.2 Results

1) Isolation of bacteria from the under surface of shoes

Bacteria were cultured using Nutrient Agar after isolation from the under surfaces of shoes.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.7) in order to establish that they were bacteria.



Escherichia coli LN558643.1 Gram stained. Magnification: 100x.



Bacillus licheniformis KP772335.1 Gram stained. Magnification: 100x.



Enterococcus mundtii KR085796.1 Gram stained. Magnification: 100x.


Enterococcus hirae KT261200.1 Gram stained. Magnification: 100x.



Bacillus licheniformis DQ071560.1 Gram stained. Magnification: 100x.



Brevibacillus borstelensis KT239000.1 Gram stained. Magnification: 100x.



Lysinibacillus fusiformis KP872952.1 Gram stained. Magnification: 100x.



Aneurinibacillus migulanus NR_113764.1 Gram stained. Magnification: 100x.



Bacillus subtilis EF488088.1 Gram stained. Magnification: 100x.

Figure 2.7: Nutrient Agar plates showing cultured bacteria isolated from the under surface of shoes, and also microscopy images showing the bacteria under the light microscope.

			NBCI		
Representative	Closest matches	Sequence	(Appaggion		
sequence	identification	identity	(Accession		
			number)		
<u> </u>		000/	LN550642.1		
GI	Escherichia coli	99%	LN558643.1		
G2	Brevibacillus borstelensis	98%	KT239000.1		
G3	Bacillus licheniformis	99%	KP772335.1		
G6	Enterococcus mundtii	99%	KR085796.1		
G7	Enterococcus hirae	98%	KT261200.1		
G9	Bacillus licheniformis	99%	DQ071560.1		
G10	Lysinibacillus fusiformis	99%	KP872952.1		
G13	Aneurinibacillus migulanus	90%	NR 11376/ 1		
015	(Bacillus brevis)	JJ /0	111 <u>1</u> 11370 1 .1		
G14	Bacillus subtilis	99%	EF488088.1		

Table 2.8: Bacteria isolated from the under surfaces of shoes.

Aneurinibacillus migulanus (Bacillus brevis)

Aneurinibacillus migulanus (formerly *Bacillus brevis*) produces the antibiotic peptide gramicidin S. The cells are rod shaped, motile, and peritrichous, and ellipsoidal spores are formed in swollen sporangia (Berditsch *et al.*, 2007). Colonies are flat and smooth on nutrient agar. This organism is positive for catalase activity, development of an alkaline pH in Voges-Proskauer broth, and has the ability to reduce of nitrate to nitrite (Berditsch *et al.*, 2007; Takagi *et al.*, 1993). *Bacillus brevis* is a common environmental organism which is not associated with human disease.

Escherichia coli

Escherichia coli is a Gram-negative, facultative anaerobic, rod-shaped, coliform bacterium that is commonly found in the lower intestine of warm blooded organisms. Most *E. coli* strains are harmless, but some serotypes can cause serious food poisoning (Liu, 2011). *E. coli* are part of the normal flora of the gut, and can bring benefit to their hosts by producing vitamin K₂ and by preventing colonization of the intestine with pathogens. This bacterium is expelled into the environment within faecal matter. Faecal transfer is the main route through which pathogenic strains cause disease. Virulent strains can cause gastroenteritis, urinary tract infections, and neonatal meningitis. *E. coli* are a major cause of urinary tract infections (Liu, 2011).

Lysinibacillus fusiformis

The is a Gram-positive rod with motile spore arranged in 2-3 chains. It is frequently isolated from groundwater-derived drinking. Wenzler *et al.*, (2015) report a case of severe sepsis caused by persistent *Lysinibacillus fusiformis* and *Paenibacillus* bacteraemia in a patient with a history of intravenous drug abuse and splenectomy; report a rare case of bacteraemia due to *Lysinibacillus* and *Paenibacillus spp*. in an immunocompetent patient without the involvement of implanted prosthesis or intravascular catheters (Rajesh *et al.*, 2013; Ahmed *et al.*, 2007; Wenzler *et al.*, 2015).

Enterococcus mundtii

Members of the genus *Enterococcus* are Gram-positive ovoid cells that appear singly, in pairs, or in short chains and are catalase- and oxidase-negative; facultative anaerobes; and homofermentative, with lactic acid being the end product of glucose fermentation. Surface colonies on blood agar or nutrient agar are circular, smooth, and entire. Enterococcal infections are a major health concern, and as nosocomial pathogens, they often prolong hospital stays. *Enterococcus mundtii* can cause postoperative endophthalmitis, although it is rarely identified in endogenous endophthalmitis (Higashide *et al.*, 2005).

Enterococcus hirae

Enterococcal disease comprises upper and lower airway, wound, hepatobiliary, intra-abdominal and urinary tract infections, meningitis, infective endocarditis and bacteraemia (including neonatal sepsis) (Poyart *et al.*, 2002). *E. hirae* has been reported to cause wound infections, gastritis, and occasional bacteraemia. It is a zoonotic pathogen rarely isolated from human infections. Poyart *et al.* (2002), however, found *E. hirae* colonized the native aortic-valve endocarditis in a 72-year-old man (Bourafa *et al.*, 2015; Poyart *et al.*, 2002; Savini *et al.*, 2013).

The results show that a wide range of bacteria could be isolated from the soles of shoes worn in normal use. This reality is reflected by the fact that in critical care facilities, visitors and staff are made to wear flexible plastic overshoes to prevent microbial contamination from outside (Falvey and Streifel 2007; Lai, 2001; Mehta, 1990).

2.9 Isolation of bacteria from books and shelves in libraries and archive storerooms

Libraries and archives act as collections for the storage of large amounts of books and documents which are affected by a variety of different environmental factors including: microclimate in store rooms (i.e. temperature and humidity of the air), type and the amount of light (i.e. type of light waves on the electromagnetic spectrum and radiation intensity), the type and amount of chemicals used for hygiene purposes and the air quality present in storage rooms. Books or other archival materials are a rich reservoir of many nutritional substances, such as cellulose, which is the main constituent of paper, and proteins, which are present in leather books. These nutritional substances stimulate the growth of a specific group of microorganisms (Kalwasińska *et al.*, 2012). Fungi are particularly strong cellulose decomposers, with species being represented, including members of *Botrytis, Chaetomium, Trichoderma, Penicillium*, while those fungi which exhibit strong proteolytic activity include species of the genera *Mucor, Aureobasidium, Chaetomium, Trichoderma, Verticillium and Epicoccum*. Bacteria however, are only infrequently found on paper, except where it is very damp when species of *Cellulomonas, Cellfalciculata, Cellvibrio* and *Cytophaga* tend to predominate (Kalwasińska *et al.*, 2012).

2.9.1 Materials and Methods

Sterile cotton swabs were wetted by dipping them in saline and used to collect samples from books and shelves in libraries and archive storerooms (Western Bank Library and the Information Commons, University of Sheffield). All samples were labelled and streaked on to nutrient agar plates; the plates were then incubated at 25°C under aerobic conditions for 24 hrs. After incubation the colonies were identified using 16S rRNA. Extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.9.2 Results

1) Isolation of bacteria from books and shelves in libraries and archive storerooms

Bacteria were cultured using Nutrient Agar after isolation from books and shelves.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.8).



Pseudomonas jessenii LN774645.1 Gram stained. Magnification: 100x.



Bacillus cereus KP192930.1 Gram stained. Magnification: 100x.



Bacillus altitudinis KT758615.1 Gram stained. Magnification: 100x.



Bacillus pumilus KP322017.1 Gram stained. Magnification: 100x.



Acinetobacter lofwii KT387352.1 Gram stained. Magnification: 100x.



Bacillus licheniformis DQ071560.1 Gram stained. Magnification: 100x.



Bacillus megaterium KU550043.1 Gram stained. Magnification: 100x.



Staphylococcus succinus KJ888125.1 Gram stained. Magnification: 100x.



Bacillus stratosphericus KJ672335.1 Gram stained. Magnification: 100x.



Bacillus licheniformis KT200463.1 Gram stained. Magnification: 100x.



Bacillus pumilus KU230023.1 Gram stained. Magnification: 100x.



Bacillus weihenstephanensis KC527665.1 Gram stained. Magnification: 100x.

Figure 2.8: Nutrient Agar plates showing cultured bacteria isolated from various from books and shelves in libraries and archive storerooms, and also microscopy images showing the bacteria under the light microscope.

Table 2.9: Bacteria present in dust settled on the surfaces of books and archive

 materials in the studied storerooms.

Representative	Closest matches	Sequence	NBCI (Accession
sequence	identification	identity	number)
9B3	Pseudomonas jessenii	99%	LN774645.1
3 SH11	Bacillus cereus	99%	KP192930.1
4 SH8	Bacillus altitudinis	98%	KT758615.1
7 SH11	Bacillus pumilus	98%	KP322017.1
1 SH6	Bacillus stratosphericus	98%	KJ672335.1
4 SH11	Bacillus weithenstephanensis	98%	KC527665.1
16 SH3	Acinetobacter lofwii	99%	KT387352.1
10B2	Bacillus licheniformis	99%	DQ071560.1
12SH4	Bacillus megaterium	99%	KU550043.1
14SH2	Staphylococcus succinus	99%	KJ888125.1
15SH3	Bacillus pumilus	99%	KU230023.1
2SH11	Bacillus licheniformis	99%	KT200463.1

Acinetobacter lwoffii

The genus *Acinetobacter* are Gram-negative, strictly aerobic, non-fermenting, nonfastidious, non-motile, catalase-positive, oxidase-negative, rod-shaped bacteria. *Acinetobacter* spp. are widely distributed in soil, water, vegetables, and are frequently isolated from animals and humans *Acinetobacter lwoffii* is part of the communal microflora on human skin and mucous membranes and causes nosocomial infections like septicaemia, pneumonia, meningitis, urinary tract infections, skin and wound infections (Regalado *et al.*, 2009) and is associated with polluted water systems (Liu, 2011).

Staphylococcus succinus

This is a Gram-positive, spherical bacterium which forms characteristic rosettes with one central cell surrounded by two to five peripheral cells; strains can be isolated from clinical specimens such as blood culture, pus, cerebrospinal fluid, exudate, eye and wound swabs (Liu, 2011).

Bacillus weihenstephanensis

Bacillus weihenstephanensis is a psychrotolerant species capable of growing at temperatures as low as $4^{\circ}C-6^{\circ}C$ is implicated in food spoilage and in foodborne illnesses (Liu, 2011).

Bacillus altitudinis

Colonies are Gram-positive, rod-shaped, endospore-forming and catalase-positive bacteria. Colonies on nutrient agar are white, convex with a regular margin and 2–3 mm in diameter (Shivaji *et al.*, 2006). Growth occurs between 8°C and 45°C and at pH 5–8. *B. altitudinis* was isolated from cryogenic tubes used for collecting stratospheric air samples from high altitudes (Shivaji *et al.*, 2006). Its pathogenicity is unknown.

Bacillus pumilus

Bacillus pumilus is one of many species within the genus *Bacillus* which have emerged as novel foodborne human pathogens that cause severe fatal infections (Kimouli *et al.*, 2011). This bacterium has also been reported to cause infectious endophthalmitis and a case of central venous catheter infection with *Bacillus pumilus* in immunocompetent patients. *Bacillus pumilus* is a member of the intestinal flora of humans and can produce exotoxins that are cytotoxic for cultured mammalian cells (Kimouli *et al.*, 2011).

Bacillus stratosphericus

Colonies on nutrient agar are white, irregular, and raised. Growth occurs between 8°C and 37°C, but not at 40°C (Shivaji *et al.*, 2006). *Bacillus stratosphericus* were isolated from cryogenic tubes used to collect air samples at altitudes of 24, 28 and

41km. Also it was isolated from cryogenic tubes used for collecting air samples from high altitudes (Shivaji *et al.*, 2006). Its pathogenicity is unknown.

Bacillus megaterium

Bacillus megaterium is a Gram-positive, mainly aerobic spore-forming bacterium found in soil, seawater, sediments, rice paddies, honey, fish, and dried food. It is not regarded as an important human pathogen (Scholle *et al.*, 2003)

Pseudomonas jessenii

Cells are Gram-negative, asporogeneous and rod shaped motile by means of single polar flagellum. Colonies are smooth on nutrient agar circular non-pigmented and non-haemolytic when they are grown on blood agar. It is not regarded as human pathogen (Verhille *et al.*, 1999).

2.9.3 Discussion

The most frequently isolated bacteria belong to the genus *Bacillus* which is a Gram-positive rod (Table 2.9), while Gram-positive cocci were represented by *Staphylococcus* spp. Gram-negative rods, like *Pseudomonas jessenii* were also isolated, as was *Acinetobacter lofwii*, a non-fermentative Gram-negative a member of the genus *Acinetobacter*. Most of the identified microorganisms have been

previously isolated from a wide variety of different materials such as leather, paintings and books (Karbowska-Berent *et al.*, 2011). *Bacillus* may cause the decomposition of adhesives in paper and book bindings. *Bacillus* has the ability to transform into spores allows them to preserve viability in unfavourable, dry conditions for a long period of time. Owing to that, these bacteria were the most numerous species in settled dust (Karbowska-Berent *et al.*, 2011).

2.10 Isolation of bacteria from the upper surface of water taps

Hospital water systems often act as a source of nosocomial infection, particularly among immunocompromised and high-dependency patients (notably in intensive care units) (Boyles *et al.*, 2012). Patients are exposed to waterborne microorganisms in hospitals when bathing, showering or washing hands following direct contact with contaminated fixtures (e.g. wash-basins and taps); by ingestion of water; via indirect contact (e.g. medical equipment rinsed with water); and by staff-transfer via medical equipment rinsed with water; and finally, by inhalation of aerosols produced by a water source, and the aspiration of contaminated water. Such microbes often originate from biofilms and sediments in supply water, water storage tanks, and water distribution network pipes as well as associated equipment (Boyles *et al.*, 2012). Water quality can become corrupted rapidly due to the formation of biofilms by bacteria in the supply water. Taps are frequently contaminated with biofilm-containing opportunistic pathogens, notably *P*.

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aeruginosa, and numerous cases of cross-infection from hospital taps have been reported (Boyles *et al.*, 2012). Immunocompromised patients are particularly susceptible to infection by such microorganisms, which can cause bacteraemia, pneumopathy, meningitis, and other conditions. *Pseudomonas aeruginosa* is particularly often isolated from these sources and has been reported to be lethal in 50%, 70%, and 20% of bacteraemia, pneumopathy, and meningitis cases, respectively (Boyles *et al.*, 2012).

2.10.1 Materials and Methods

Sterile cotton swabs were wetted by dipping them in saline and used to collect samples from tops of water taps in toilets in Firth Court and the Student Union Building (University of Sheffield), and from volunteer house owners. All samples were labelled and streaked on to Nutrient Agar plates; the plates were then incubated at 25°C under aerobic conditions for 24 hrs. After incubation, the colonies were identified. Genomic DNA extracted from each isolate was identified using 16S rRNA. The extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.10.2 Results

1) Isolation of bacteria from the upper surface of water taps

Bacteria were cultured using Nutrient Agar after isolation from the upper surface of water taps.

2) Light microscope images. The isolates were examined under a light

microscope (Figure 2.9).



Delftia lacustris KT958881.1 Gram stained. Magnification: 100x.



Rothia amarae NR_029045.1 Gram stained. Magnification: 100x.



Delftia acidovorans (comamonas acid) JX090199.1. Gram stained. Magnification: 100x.



Arthrobacter sanguinis NR_044399.1 Gram stained. Magnification: 100x.



Pseudomonas aeruginosa KF680991.1 Gram stained. Magnification: 100x.



Bacillus cereus GQ344804.1 Gram stained. Magnification: 100x.

Figure 2.9: Nutrient Agar plates showing cultured bacteria isolated from the topsurface of water taps, and also microscopy images showing the bacteria under the light microscope.

Representative	Closest matches	Sequence	NBCI (Accession			
sequence	identification	identity	number)			
2tt	Staphylococcus epidermidis	98%	KT887972.1			
3tt	Rothia amarae	99%	NR_029045.1			
4tt	Acinetobacter johnsonii	99%	KP763485.1			
6tt	Delftia lacustris	99%	KT958881.1			
7tt	Pseudomonas aeruginosa	98%	KF680991.1			
8tt	Delftia acidovorans (comamonas acid)	99%	JX090199.1			
9tt	Arthrobacter sanguinis	99%	NR_044399.1			
10tt	Delftia acidovorans (comamonas acid)	98%	KJ781879.1			
11tt	Bacillus cereus	94%	GQ344804.1			

Table 2.10: Bacteria isolated from the upper surface of water taps.

Pseudomonas aeruginosa

Pseudomonas spp. act as opportunistic pathogens with *P. aeruginosa* being the most clinically significant as a major cause of lower respiratory tract, bloodstream, and urinary tract infections, particularly among patients in intensive care units or with compromised immunity (Liu, 2011). This bacterium is often isolated from

moist environments within health care settings, such as taps, toilets, showers, respiratory equipment, and cleaning solutions. *Psuedomonas aeruginosa* is a major pathogen in chronic lung diseases, most notably in cystic fibrosis. It causes infections in critically ill patients in intensive care units, burn patients, and particularly when immunocompromised (Liu, 2011). Such infections include ventilator-associated pneumonia, catheter-related bloodstream or urinary tract infections; other infections are derived from medical devices or penetrating trauma, sepsis in neutropenic cancer and transplant patients, skin and soft-tissue infections in burn and diabetic patients, postoperative endophthalmitis and necrotizing otitis externa in diabetics (Liu, 2011).

Delftia acidovorans (Comamonas acidoverans)

Cells of this bacterium are Gram-negative straight to slightly curved rods, which occur singly or in pairs. They are motile and are oxidase and catalase positive. This organism has been isolated from serious infections such as central venous catheter-associated bacteraemia, corneal ulcers, otitis media exist (Khan *et al.*, 2012). *Delftia acidovorans* infections have also been reported from cases of endocarditis, ocular infections, acute suppurative otitis media, bacteraemia, urinary tract infections, intravascular catheter related infections and empyema; serious infections have been found amongst immunocompromised patients. Patients with indwelling devices are at particular high risk of acquiring *Delftia acidovorans* infections (Khan *et al.*, 2012).

Acinetobacter johnsonii

Acinetobacter spp. have been isolated from a variety of opportunistic infections, including septicaemia, pneumonia, endocarditis, meningitis, skin and wound infection, and urinary tract infection. *Acinetobacter spp*. (e.g., *A. johnsonii*) have been shown to form an important component of the communal microflora on human skin and mucous membranes (Bergogne-Bérézin *et al.*, 1996)

Rothia amarae

All *Rothia* species are Gram-positive and are isolated from humans or animals as well as from soil and water. They are particularly associated with dental caries and periodontal disease notably in immunocompromised hosts, but rarely in healthy hosts. Clinical syndromes linked to *Rothia* infection include bacteraemia, endocarditis, meningitis, peritonitis, bone and joint infections, pneumonia, skin and soft tissue infection, endophthalmitis, and prosthetic device infection (Fan *et al.*, 2002; Xion *et al.*, 2013; Ramanan *et al.*, 2014).

Arthrobacter sanguinis

Arthrobacter are Gram-positive aerobic bacilli commonly found in the environment, although they can also cause endophthalmitis after intra-ocular implantation, infective endocardites in intravenous drug- abuser, featal demise and disseminated intravascular coagulation in pregnant women and catheter-related bacteraemia in leukemic patients (Mages *et al.*, 2008; Yap *et al.*, 2015). *Arthrobacter* infections are often difficult to diagnose using conventional

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biochemical assays. *Arthrobacter* related peritonitis can be successfully treated with appropriate parenteral antibiotics without the need for catheter removal (Mages *et al.*, 2008, Yap *et al.*, 2015).

2.11 Determination of the number of bacteria on hands after washing and drying normally and with warm air hand dryer

2.11.1 Introduction

Hand drying represents the final part of the hygiene procedure in a public toilets and if the toilet is designed well, the number of surfaces which the user subsequently touches will be limited or reduced to near zero. This is essential because wet hands can spread up to one thousand times more bacteria than can dry hands (Smith, 2009). This is essentially because water transfers easily between surfaces and because bacteria do best in moist environments (Redway and Fawdar, 2008). As a result, it is critical that hands are not contaminated with bacteria due to the drying process (Harrison et al., 2003). The most frequently used means of hand drying are paper towels, hot air dryers, jet air dryers and cloth towels. It has been suggested that air dryers should be avoided as they accumulate aerosols from toilets which then contaminate hands (Snyder, 1998). This last mentioned author concludes that the use of paper towels decreases the number of bacteria on hands, while hot-air dryers conversely increased contamination by some bacteria; a finding suggestion that has been widely debated (Holah, 2011); many contend

however, that hot air dryers are often slow and inefficient, leaving the hands of users moist and possibly still contaminated. Cloth roller towels are similarly not recommended essentially because they are generally of low capacity, and when a roll comes to an end it becomes available for common use and is therefore likely to increase the spread of pathogens (Snyder, 1998).

This work was undertaken with the aim of evaluating the performance of warm air hand driers, in toilets relative to bacterial contamination. First, the ability of warm air driers to dry hands hygienically was evaluated by measuring the number of microorganisms on different working days. Secondly, it was determined if warm air driers influence the level of air-borne microorganisms in the washroom environment, as was suggested by Knights *et al.* (1993). Finally, the surfaces of warm air driers and other washroom areas were examined for total viable counts in order to determine if the use of air driers affects bacterial distribution.

2.11.2 Materials and Methods

General purpose, non-antimicrobial bar soap was used for hand washing. Hands were wetted with tap water, and the soap applied and lathered together vigorously for 5 minutes, covering all surfaces of hands followed by rinsing with tap water. Finally, the hands were shaken five times in order to remove of excess water before drying. Areas of the washed hands were applied to the surface of Nutrient Agar plate; the plates were then incubated at 37°C for 48h. Warm air hand-dryers were next used for hand drying; the air temperature applied was 0°C -40°C at a distance of 15 cm. When hands were placed in the 15 cm drying space, warm air flowing automatically was turned on. During drying, hands were either rubbed or had their palms turned upward and were held stationary. Then dried hands and fingers top were touched onto the surface of nutrient agar plate. The plates were incubated at 37°C for 48 h and scored for the presence or absence of growing bacteria.

2.11.3 Results

The results show that no bacteria were isolated from hands dried in the normal way, but were isolated from hands when dried using an air dryer.



Figure 2.10: Agar plates used to for the assessment of bacterial contamination on hands. (Left) hand washed and dried with a warm air hand dryer, (Right) hand washed and dried without warm air dryer.

2.12. Quantification of bacteria transferred from hand warm air dryers

2.12.1 Materials and methods

Nutrient Agar plates were used in this experiment to isolate the transferred bacteria from hand warm air dryers. Any bacteria transferred to the plates from the hot air driers were counted and the effect of drying times was determined.

2.12.2 Results

Table 2.11: Showing the mean of the bacteria count using three different types warm air dryer (WAD,1-3) for different drying times.

	5	10	15	20	25	30	35	40	45	50	55	60
	Sec											
WAD1	0	2	5	6	3	9	12	10	23	41	31	30
WAD2	12	17	18	22	18	16	50	37	15	49	44	63
WAD3	19	20	15	24	10	8	11	20	18	19	21	30

Table shows that there was a correlation between the drying time and the resultant bacterial counts, with the counts generally increasing with the drying time exceeding 30 seconds. Redway and Fawdar (2008) investigated the spread of

contamination from different drying methods and concluded that microorganisms were spread significantly further when an air blade dryer was used instead of paper towels. While not significant if only non-pathogenic microorganisms are spread, the risk of pathogen contamination of the environment will result when pathogens are present on hands and are liberated into the air (Blackmore, 1989).

2.12.3 Discussion

From the results presented in this Thesis, it is clear that hand dryers distribute bacteria into the surrounding air. As has been mentioned elsewhere in this Thesis, many of these bacteria, even when not direct pathogens, pose a potential problem for immunocompromised patients here. Hand dryers have recently become increasingly popular and for reasons of cost, these machines are often supplied in place of traditional cloth and paper towels (Huang *et al.*, 2012). This does not present a health problem for most hospital patients and visitors (except where immunocompromised). The use of hand dryers in medical settings should therefore be reviewed and, if necessary, they should be replacing by traditional approaches, the consensus being that paper towels are the preferred method of hand drying in such settings (Best *et al.*, 2014; Huang *et al.*, 2012).

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2.13. General Discussion regarding potential pathogens in the environment

Boyce et al., (1997) have shown that surfaces in hospitals are often contaminated with MRSA, as are staff. This points out the necessity of using detergents and antibacterial wipes to reduce bacterial loads, especially postoperative MRSA. Vancomycin-resistant enterococci (VRE) can achieve long-term survival in medical environment, even following extensive cleaning and disinfection. Pathogens can, in fact, often be spread when cleaning cloths are a) re-used on surfaces, b) when little contact time between a surface and the applied disinfectant, and c) when surface are casually sprayed and wiped instead of being aggressively scrubbed. VRE appears to be particularly efficient at surviving successive disinfection, regimes, even so-called "double bleach-based cleaning". Disinfectants can be effective if hospital surfaces are scrubbed daily and when this is associated with a hand cleaning programme. The surface screening of cultures has also proved effective, as has the use of 5% sodium hypochlorite to clean all surfaces at least three times daily. Both MRSA and VRE can survive for long periods which presents an obvious risk in relation to the movement of patients into rooms which have been previously occupied by infected patients.

Hospital sinks are an obvious source of pathogenic bacteria, with antibiotic resistant *Klebsiella pneumoniae* strains showing particularly extended survival within plumbing thereby, in extreme cases, necessitating the replacement of sinks and related pipe-work. Pathogen survival is also linked to biofilms adhering to

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surfaces on sinks and associated plumbing and unfortunately the use of chlorinecontaining products is often ineffective and even prolonged use fails to destroy all biofilms thereby necessitating the physical disruption of any biofilm which is found to be growing in contaminated plumbing systems.

2.13.1 Antimicrobial surfaces

Some high-tech solutions are currently available to control surface pathogen contamination, by so-called, "self-sanitizing surfaces" which include hard metals (e.g. copper and silver) and novel materials such as light-activated titanium dioxide-containing surfaces which prevent the accumulation of surface microbes (Dancer, 2014). Several types of antimicrobial surfaces including antiadhesive coatings, and antimicrobial coatings exist including Triclosan (Microban triclosan): which is used on surfaces and in many antibacterial liquid and bar soaps and 30% of bar soaps, although evidence for resistance is developing. Silver: Silver ions (Ag⁺) bind to thiol (-SH) groups in microbial enzymes and proteins and lead to microbial-cell inactivation; such coatings are not, however, permanently active and resistance can develop (Dancer, 2014). Copper: Copper (as will be discussed below) is also toxic to pathogens and coatings can be used to reduce microbial contamination. Polycationic antimicrobial surfaces: these are surfaces which are treated with hydrophobic, negatively charged poly-cations which kill bacteria by bringing about physical damage to the cell-envelope. Lightactivated antimicrobial surfaces: These produce unselective, reactive radicals

which kill a range of microorganism and reduce the chances of resistance developing. Two types are available, the first is based on a photosensitizer immobilized within a coating, while the second is a coating which contains a titanium dioxide (TiO_2)-based catalyst (Dancer, 2014).

CHAPTER 3

STUDIES ON SURVIVAL OF BACTERIA ON VARIOUS SURFACES

3.1 Studies on the survival of bacteria on smooth and rough unglazed ceramic tiles under ambient conditions

3.1.1 Introduction

Ceramic tiles are frequently used in the built environment, notably in hospitals and provide an obvious survival environment for potentially pathogenic microorganisms. Ceramic tiles are widely used in domestic and health care settings. They are obviously advantageous in being durable, water repellent, easy to keep clean, and are readily exposed to bacteria-killing sunlight. The grouting between tiles may however, act as a reservoir for microbes, including pathogens. Tile surfaces differ, between smooth and rough. Logically, one would assume that rough, unglazed tiles would provide a more suitable environment for microbes and as a result, these are generally preferred for use in health-care settings. The aim of this work was to determine the survival (in the ambient environment) of a variety of bacteria on smooth (glazed) and rough (unglazed) tiles.

3.1.2 Materials and Methods

Ceramic tiles having either smooth or rough surfaces (9 cm²) were firstly sterilized by autoclaving (Figure 3.1). The test bacteria were then grown on Nutrient Agar medium as a pure culture, while yeasts (*Candida albicans* and *C. rugosa* were grown on Sabouraud Dextrose agar). Bacterial and yeast suspensions from colonies were prepared in sterile saline equal to McFarland 0.5 turbidity (the density of a bacterial suspension equal 1.5x 10⁸ colony forming units CFU/ml). The tiles were inoculated by placing on their uppermost chosen surface approximately 10 microliters of either a bacterial or yeast suspension, which was allowed to outspread and dry under sterile conditions and left at room temperature (18-23°C). The tiles were transferred to sterile Falcon tubes containing sterile water and serial dilution counts were made.

In the second series of experiments, the tiles were inoculated with *Candia rugosa* and at the end of the same exposure period were washed and serial dilution plated onto Sabouraud Dextrose agar.



Figure 3.1: The two types of ceramic tiles used: A) smooth ceramic tile, B) rough ceramic tile.

3.1.3. Results

3.1.3.1 Bacterial survival on ceramic tiles

Figure (3.2) shows the number of S. aureus and E. coli bacteria isolated from the two tile types. Numbers of S. aureus on the rough tiles increased over the incubation period, while number on the smooth tiles decreased. In both cases however, bacteria were still present after 65h. In the case of *E. coli*, the number decreased over the incubation period, while there was a slight increase in the numbers of bacteria on smooth tiles (Figure 3.3). The results show that both S. aureus and E. coli can survive on both tile types over a 65 h period, a finding which is obviously relevant to pathogen survival and transfer in healthcare settings. Staphylococci are among the most resistant non-spore forming bacteria, and they can survive at various environmental conditions. They can be cultured from dried clinical material after several months. Yazgi et al. (2009) found that S. aureus survived on laminate for 75.4 days and inox sheet for 68.2 days, whereas these survival days were 73.8 days on ceramic tiles and 63.4 days on vinyl flooring. The length of survival of these organisms on the various materials may have significance for infection control in hospitals. For example, the ceramic tiles tested in this study are the material used for floors and walls of hospitals that are handled by both patients and staff when they are touched or cleaned. Staphylococci and E. coli survived for days on this covering materials, suggesting

that they could act as reservoirs for such pathogenic bacteria.



Figure 3.2: Survival of *S. aureus* on ceramic tiles during 65h.



Figure 3.3: Survival of *E. coli* on ceramic tiles during 65h.
3.1.3.2 Yeast survival on ceramic tiles

The results (Figure 3.4) show that *Candida rugosa* survived for a period of 72h on both smooth and rough tiles with numbers generally increasing for 53-56h when they declined rapidly but the yeast was still present in a viable state after 72h. The yeast is associated with nosocomial infections in patients. Where comparisons are possible for the yeast data, this result agrees with Traoré et al. (2002) who found C. albicans and C. parapsilosis remained viable for at least 3 and 14 days when dried on surfaces materials. The results of the ability of *C. albicans* and *C. parapsilosis* to survive on glass and metal carriers over a 14-day period are summarized in a study by Traoré et al. (2002). Candida parapsilosis was found to survive much better than C. albicans on both types of non-porous surface. Also, C. albicans was undetectable at the end of the third day, while C. parapsilosis remained detectable even after 14 days under ambient conditions. There was a significant difference between the survival of C. parapsilosis and C. albicans after 14 days on glass carriers and stainless steel carriers. These results indicate that ceramic tiles play a role as reservoirs or vectors for fungi because those tested generally remained viable on these surfaces for many days. In this age of increasing antifungal resistance, these survival data indicate that the appropriate disinfection of the environment and control procedures should be used to control infections in hospitals.



Figure 3.4: Survival of *Candida rugosa* on ceramic tiles.

3.1.3.4 Discussion

The most important factor in transmission of microorganisms in the environment is the ability of microorganisms to survive on environmental surfaces. The contamination and colonization of surfaces is important in relation to the transmission of nosocomial infections pathogens, such as methicillin-resistant *S. aureus* and vancomycin resistant Enterococci (VRE). For example, a range of floor covering materials are used in intensive care units, operating rooms, clinics, and laboratories of hospitals due to which influence the rate of microbial survival and as a result, need to be carefully chosen in order to minimize microbial colonization and survival. The preferred materials are smooth-surfaced and include ceramic tiles, laminates, inox sheets, and vinyl which are easy to disinfect

during cleaning. Neely and Maley (2000) studied the survival of several clinical and environmental *staphylococci* and *enterococci* on fabrics and plastic that are used in hospitals. They found that the survival period of *S. aureus* was more than 90 days on various surfaces, whereas it was 18 to more than 80 days for *enterococci*. It is clear from the results for both tiles and the study discussed in this Thesis that covering materials have a significant influence on the survival periods of bacteria. The Thesis results show that ceramic tiles play a role as reservoirs or vectors for the yeast, *C. rugose*, because this organism generally remained viable on these surfaces for many days. Strategies to reduce the rates of nosocomial infection with these pathogens should conform to established guidelines, with an emphasis on thorough environmental cleaning and use of Environmental Protection Agency–approved detergent-disinfectants.

3.1.4 Discussion

Bacterial and yeast survival was tested on both smooth and rough ceramic tiles. The results show that ceramic tiles provide reservoirs for microbes because the tested species generally remained viable on these surfaces for a number of days; the length of survival on both smooth and rough ceramic tiles being related to both the genus and the species. Clearly cleaning strategies aimed at maintaining low pathogen populations on hospital tiles, whether rough or smooth need to take such apparently anomalous findings into consideration.

3.2. Studies on the survival of bacteria on toothbrushes under ambient conditions

3.2.1 Introduction

The environment of the toothbrush is affected by many conditions whether it is the architecture of the toothbrush itself regarding bristles or by adjusting the pH level. These conditions alter the population of bacteria on the toothbrush. While the toothbrush is not the ideal niche for a microbe, the toothbrush is capable of supporting microbial life (Downes *et al.*, 2008).

3.2.2 Materials and methods

3.2.2.1 Bacterial survival on toothbrushes during a month

The new toothbrushes were prepared and were sterilized by 10% ethanol spray. The strains of the test bacteria were grown on the Nutrient Agar medium as a pure culture. Bacterial suspensions from the colonies were prepared in sterile saline equal to McFarland 0.5 turbidity. The toothbrushes were contaminated by placing on them on approximately 10 microliter volume of the bacterial suspension, and the inoculum allowed to outspread and dry itself under sterile conditions. Then the toothbrushes were kept at room temperature (18-23°C) (Figure 3.5). Beginning from the third day, toothbrushes were taken and placed in a sterile Falcon tubes contained sterile water. The numbers of viable organisms in the resultant suspension were determined by serial dilution and plating on to Nutrient Agar. The step was repeated after 5, 15, and 20 days and then the plates were incubated at 37°C for 48 h.



Figure 3.5: New toothbrushes inoculated with the bacterial suspension of *S. aureus* and *E. coli*.

3.2.3. Results and discussion

Numbers of inoculated *S. aureus* and *E. coli* on toothbrushes decreased after 5 days of storage (Figure 3.6 A, B) and then numbers remained constant (except for small increase in numbers of *S. aureus* at 37 days. *Escherichia coli* also survived for 5 days on tested toothbrushes.



Figure 3.6: Population density of *Staphyolcoccus aureus* (A) and *E. coli* (B) when inoculated onto toothbrushes (analysis of variance).

* All values after 5 days were significantly different from control (p=0.05)

3.2.4 Discussion

The aim of this study was to compare bacterial survival on new conventional toothbrushes in the laboratory. The survival times of *Staphylococcus aureus* and *E*. coli were compared on toothbrushes stored in the laboratory. Toothbrushes can become contaminated through contact with the environment, and bacterial survival is affected by toothbrush storage containers. Dayoub et al. (1977) found that toothbrushes placed in closed containers and exposure to contaminated surfaces give higher bacterial counts than those left open to air. Mehta et al. (1990) found that the use of a cap for toothbrush storage increased bacteria survival. Increased humidity in the environment increases bacterial survival on toothbrushes and bacteria can survive for more than 24 hours in the presence of moisture. The persistence of viable *Staphylococci* on a drying toothbrush in the humid atmosphere of a toothbrush holder is not surprising; interventions such as chlorhexidine, toothpaste, mouthwash, and ultraviolet sanitizers cab however, reduce bacterial survival (Downes et al., 2008).

Although sterile at birth, a great variety of microbes develops during the first day of life, including: *Streptococcus*, *Staphylococcus*, *Neisseria*, *Candida*, *Lactobacillus*, *Veillonella and* coliforms (McCarthy *et al.*, 1965; Socransky and Manganiello, 1971). Mutans *Streptococci* (MS), which is the primary etiological agent of human dental decay, is however, only found following dental eruption because it needs a hard surface on which to develop. Catalanotto (1975), Fujiwara (1991), Glass, (1992) and Glass and Lare (1996) showed that toothbrushes are particularly relevant to the transmission of pathogens amongst

immunocompromised patients, including coliforms picked up from the bathroom environment (Verran, 1996). While it is of course unlikely that toothbrushes will be intentionally shared amongst families and individual patients in hospitals, the potential still remains for accidental contamination-transfer. More importantly perhaps, is the fact that pathogens can re-infect patients from toothbrushes at the initial point when they are first immunocompromised and recycle pathogens, including MRSA. Toothbrushes are often exposed to sunlight and nearly always to drying –wetting cycles, all of which can reduce survivability. As a result, toothbrushes rapidly become contaminated with oral microbes, including caries causing pathogens like *Streptococcus mutans*, and opportunistic yeast pathogens like Candida albicans (Sammons et al., 2004). Additionally, organisms not usually considered to be components of the oral microflora can frequently be isolated from toothbrushes, including Enterobacteria and pseudomonads. Toothbrushes can therefore be seen as a potential source of both oral and systemic infection and re-infection (Sammons et al., 2004), particularly in immunocompromised patients and such organisms may also cause worrying medical problems in pregnant women (Bunete et al., 2000). The aim of the work described in this section was to therefore to determine if household toothbrushes are contaminated with bacteria.

3.3 Influence of copper and plastic surfaces on the survival of bacteria in relation to the health care environment

3.3.1 Introduction

Metallic copper (Cu) surfaces have antimicrobial properties against a variety of different microorganisms and copper touch surfaces are likely to be increasingly used in public places including hospitals (Espírito Santo *et al.*, 2011). In recent hospital trials, non-Cu surfaces in frequent contact with both patients and staff were replaced with their Cu equivalents. The use of metallic Cu resulted in diminishing bacterial surface-loads of up to 90% as compared to no-copper controls (Espírito Santo *et al.*, 2011). Molecular mechanisms that result in rapid killing of Cu surface-exposed bacteria and yeasts were studied and shown to result from a sharp shock of extreme and immediate Cu-ion overload combined with severe membrane and cell envelope damage, although similar low mutation rates were observed in cells obtained from both Cu and control surfaces (Espirito Santo *et al.*, 2011).

Mehtar *et al.*, (2008) conducted studies in which cells in buffer were applied to copper surfaces, incubated under ambient conditions, and were seen to be killed within hours, thereby mimicking contact of microbes to dry copper touch surfaces. Under such conditions, most microbes were found to be killed within minutes. Quaranta *et al.* (2011) found that copper ions are released from metallic copper upon contact with cells, a process which undoubtedly contributes to contact-

mediated killing. Extracellular supplementation with substances which are protective against oxidative stress (e.g. Catalase, superoxide dismutase, or mannitol, a hydroxyl radical quencher) increased the time needed to kill copper surface-exposed *E. coli* cells, indicating a role for this process in Cu-mediated killing.

3.3.2 Materials and Methods

Survival of bacteria on the surfaces of copper and plastic plumbing surfaces

The antibacterial activity of copper surfaces was determined by overlying suspensions of *Staphylococcus aureus* and *Escherichia coli* on copper surfaces. Two copper items were used; a copper pipe push-fit elbow 15mm, a copper pipe compression stop end, 15 mm and (as controls) a plastic pipe connector 22mm (all obtained locally from Wickes Building Supplies Ltd) (Figure 3.7). All copper pipes were sterilized by autoclaving (120°C for 20 min), while the plastic pipes were sterilized using a 10% ethanol spray. Bacterial suspensions from colonies were prepared in sterile saline equal to McFarland 0.5 turbidity. The pipes were contaminated by transferring a 10 microliter volume of the bacterial suspension, and the inoculum was allowed to out-spread and dry itself under sterile conditions. The experiments were performed at 18-23°C and each assay was carried out in duplicate. Results were calculated after 20 days. The numbers of viable organisms in the suspension were determined by serial dilution and plating on to Nutrient Agar plates; the plates being incubated at 37°C for 48h.



Figure 3.7: Copper and plastic pipes used to evaluate the antibacterial activity of copper and plastic surfaces.

3.3.3 Results



Figure 3.8: S. aureus and E. coli inoculated onto copper and plastic pipes.

Although copper and brass are traditionally used by UK plumbers for water pipes. in both hospitals and homes, they are gradually being replaced by plastic piping. It was of interest therefore to determine the survival of bacteria on both types of these surfaces. Copper is well known to be an antibacterial, and its use in medical environments is likely to lead to the continuous reduction of environmental microbial contamination, including MRSA. In viability assays, low counts of Staphylococcus aureus were seen on copper surfaces, as compared with those obtained on the plastic, control surfaces (Figure 3.8). The number of bacteria isolated from the plastic surfaces was consistently higher than the number isolated from copper surfaces. The survival rate of bacteria on the copper surfaces was low and none of the inoculated bacteria survived after 20 days of exposure. These results are similar to those found by Noyce et al. (2006) for copper, where survival was significantly lowered, with complete kill of MRSA being achieved after 90 min of exposure. Complete kills were produced on copper for all three strains of MRSA after 45, 60 and 90 min. The results presented here showed that E. coli failed to survive on copper pipes. These results are in agreement with those published by Wilks et al. (2005) who found that no viable Escherichia coli O157H7 were found on any of the copper pieces which they inoculated, even after only seventy-five to ninety minutes of exposure. Such results show that copper surfaces can remove cross contamination of pathogens in both households and health care settings and the continued replacement of other plumbing materials

with copper should minimize the risk of the survival and transmission of hospitalacquired infections.

3.3.4 Discussion

Copper has inadvertently been used as an antimicrobial agent for millennia, but its mode of action against microbes is still largely unknown. Today, nosocomial infections are a major health care threat and are responsible for large numbers of deaths and the resultant additional hospital costs, notably in the United States (Mehtar et al., 2008). The effectiveness of copper an antimicrobial material is not restricted to bacteria as it also kills *Candida albicans* (and other pathogenic yeasts) which have emerged as important agents of hospital-acquired infections (HAI) (Mehtar et al., 2008). Numerous studies have focused on the antimicrobial proprieties of copper (and its alloys) surfaces mainly in relation to the killing kinetics of a variety of microbial species exposed to different copper alloys. However, although it is becoming increasingly clear that metallic copper has excellent antimicrobial properties the molecular mode of action exerted by copper surfaces and the sensitivity of various cellular targets to its action are still largely unknown (Quaranta et al., 2011). Studies have suggested a role for copper in ion homeostasis a factor in the survival of bacteria on copper surfaces. Cells of Pseudomonas aeruginosa, Enterococcus hirae, or E. coli which had their copperion defence systems deleted have been shown to die more rapidly than wild-type cells when exposed on metallic copper, although the survival times of mutants were not dramatically different from those of wild-type cells (Quaranta et al.,

2011). Mehtar *et al.*, (2008) reported that the minimum concentration of Cu to be an effective antimicrobial agent is > 55% for bacteria excluding *Mycobacterium tuberculosis* (MTB). Copper and its alloys showed a marked inhibitory effect on MTB, despite the strains being drug resistant. Growth of both strains showed inhibition by Cu (88–98% inhibition). The above mentioned work shows that the incorporation of Cu in healthcare facilities may dramatically help reduce the environmental microbial burden and act as a useful adjunct to current infection prevention and control systems.

3.3.4.1 Copper pipes

Although copper is a nutrient, when present in large amounts it becomes highly toxic is to organisms including hospital pathogens like MRSA. This effect is due to its ability to bring about the rapid fragmentation of a cell's DNA which leads to death; as a result, it is an effective biocide (Warnes *et al.*, 2010). As a result, it is obvious that, wherever possible, plumbing materials in health care settings should be made of copper. As we have seen, hand to hand transfer is the most important means by which microbes, including pathogens, are spread in hospitals. More generalized microbial contamination of the hospital environment also takes place (Noyce *et al.*, 2006) with the result that inanimate surfaces can act as a source of nosocomial infections largely because most Gram-positive and Gram-negative bacteria are able to survive for months on dry surfaces (Hota, 2004), and the main factors influencing the survival of microbes on dry surfaces include exposure to sunlight, humidity, and temperature (Kramer *et al.*, 2006).

Recently, considerable attention has been focused on revaluating metallic copper as a biocide for use in killing bacteria on so-called "touch surfaces", which include door handles, bathroom materials, and metallic bed rails (Grass *et al.*, 2011). Weaver *et al.* (2010), for example, evaluated copper, as an alternative to alumium, as an antimicrobial surface for use in air-conditioning systems and this showed increased pathogen-killing by the former.

The copper content in copper alloys varies from 60% to 99.9%, and initial studies have shown that the survival rates of E. coli O157 vary considerably depending on the type of alloy used relative to its copper content (Noyce et al., 2006). These authors noted a gene-mediated response in *P. aeruginosa* to survival on copper alloys and in E. coli and E. faecium strains increased resistance to copper ions was due to the presence of additional plasmid-borne copper resistance genes. It was noted that the greatest differences in survival rates were related to moisture content in the sample, the type of medium the bacteria were suspended in, and whether it was a Gram-negative or a Gram-positive bacterium. Since copper ion toxicity had been reported previously as a possible driving force in the contactkilling of E. coli on copper alloys (Espirito Santo et al., 2008), an experiment was designed to block copper ions released from the copper surface by applying a corrosion inhibitor which would consequently prevent copper ions from entering the bacteria. This was correlated to electrochemical measurements and calculated concentrations of copper released from the surface, and an inverse relationship

was shown between the copper ion concentration released from the surface and survival rates of copper ion-resistant *E. coli*. Subsequently it has been shown that bacterial cells very quickly take up copper ions when exposed to copper surfaces which results in rapid cell death presumably related to increased oxidative stress (Grass *et al.*, 2010; Espirito Santo *et al.*, 2011). Copper surfaces are now being tested in hospitals where their efficacy is compared to stainless steel touch surfaces for long term observation, isolation, and characterization of surviving microbes. However, bacteria will continue to acquire resistance and will continue to proliferate in nature due to pollution -exposure. Here copper resistance can be quite useful, such as in the bioremediation of copper mining wastes (Grass *et al.*, 2011).

CHAPTER 4

ISOLATION OF BACTERIA FROM USED TOOTHBRUSHES AND DETERMINATION OF THE ANTI BACTERIAL POTENTIAL OF TOOTHPASTES

4.1.1 Introduction

The oral cavity can be colonized by more than 700 microbial species, including fungi, viruses and a variety of unclassified microorganisms, most being commensal species that are beneficial for oral health. However, some are pathogens and can overcome the host responses and cause serious oral diseases. The oral microflora can be divided into two groups:

- **Saprophytes:** These are the permanent microflora of the oral cavity whose presence is needed for the normal functioning of the dental system, as well as the body in general. The saprophytic microflora affects the conditions of the local immune system, prevents the development of pathological conditions and supports the overall bacterial balance.
- Pathogenic Microflora: These affect the organs and tissues of the mouth and entire body and bring about the emergence and development of a variety of diseases. This microflora ideally should not be present, or present in very low numbers that fail to substantially affect the oral cavity and the body in general (Otoikhian and Okoror, 2012).

An overload of pathogens in the oral microflora can result in caries, periodontitis and stomatitis, which are the most common oral microbial infections in humans. *Streptococcus spp.* and *Actinomyces spp.* are particularly common colonizers and cause supragingival plaque during the first stage of biofilm formation (Nascimento *et al.*, 2015). They also increase the acidity of the oral biofilm and thereby help to develop their cariogenic potential. After biofilm maturation, anaerobic and proteolytic bacteria, notably *Porphyromonas gingivalis*, *Tannerella forsythia, Treponema denticola* and *Fusobacterium spp.* are found to dominate the subgingival biofilm (Nascimento *et al.*, 2015).

Bacterial proteases and metabolic products often induce host responses like inflammation and immunoreactions which result in periodontitis/peri-implantitis. *Candida spp*. are the most common fungi in the oral cavity and often lead to denture stomatitis commonly isolated from infected root canals; additionally, they can act as opportunists in the formation of periodontal and peri-implantar lesions (Nascimento *et al.*, 2015).

Toothbrushes generally become highly contaminated with microorganisms which may arise from the oral cavity and from the environment in which toothbrushes are stored. As a result, toothbrushes may play a significant role in disease transmission and lead to an increase in infection risk (Frazelle and Munro, 2012). They act as a reservoir for microorganisms in both healthy and ill adults and contamination and survival of infectious organisms may occur on both animate and inanimate objects. Contamination of toothbrushes occurs after first use and increases with repeated use and they can become contaminated from a variety of routes including: the oral cavity, environment, hands, aerosol contamination, and storage containers. Bacteria may become attached to and accumulate and survive on toothbrushes which may transmit individual disease causing organisms (Frazelle and Munro, 2012).

There is a clear correlation between contaminated toothbrushes and the presence of diseases (Glass and Lare, 1986), and changing a toothbrush regularly has been linked to the elimination of symptoms and disease regardless of its nature. Moisture is an obvious factor in promoting the survival of bacteria on toothbrushes and immunocompromised patients are at particular risk from their use.

4.1.2. Materials and Methods

Fifty toothbrushes were collected from volunteers aged 5 to 45 years. The toothbrushes were transported to the laboratory in a sterile polythene bag sealed with a rubber band. Brushes were processed within 12h by a method modified from that described by Sammons *et al.* (2004). The handle was cut off using a rotary saw and the head of the brush was retained in the bag to avoid contamination. Each brush head was then subjected to soaking in 10 ml of sterile water, for 20 min, followed by vigorous vortex mixing for 1 min and manual swabbing to dislodge persistently adherent bacteria. The resulting bacterial suspension was serially diluted and 0.1 ml aliquots plated onto nutrient agar to

select for bacteria. Plates were incubated aerobically at 37°C for 24–48 h. Total viable counts were estimated from the numbers of colonies on nutrient agar plates. Colony colour, morphology and haemolysis were recorded and Gram's stain was performed on a representative of each colony morphotype. Extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. Test sample preparation for PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis, has been explained above.

4.1.3. Results

1) Isolation of bacteria from various used manual toothbrushes by cultivation on Nutrient Agar medium

Bacteria were cultured using Nutrient Agar after isolation from used toothbrushes.

Table 4.1: Bacteria isolated from various used toothbrushes.

Representative		Sequence	NBCI (Accession
sequence	Closest matches identification	identity	number)
1 TB	Roseomonas mucosa	99%	KF247232.1
2 TB	Stenotrophomonas maltophilia	99%	LN890169.1
3 TB	Pseudomonas aeruginosa	99%	KR815846.1
5 TB	Leclercia adecarboxylata	99%	KT899848.1
6 TB	Enterobacter asburiae	99%	EU239468.1
7 TB	Candidatus Roseomonas massiliae	99%	KT321690.1
8 TB	Pseudomonas parafulva	99%	KT758848.1
9 TB	Bacillus licheniformis	99%	KU314515.1
11 TB	Pseudomonas aeruginosa	99%	KF680991.1
13 TB	Agrobacterium larrymoorei	99%	EF178437.1
15 TB	Pantoea septica	99%	KF475883.1
16 TB	Stenotrophomomnas rhizophila	99%	KP050794.1
18 TB	Citrobacter freundii	87%	CP007557
20 TB	Pseudomonas frederiksbergensis	99%	EU373369.1

2) Light microscope images.

The isolates were examined under a light microscope after the bacteria were

identified. (Figure 4.1)



Roseomonas mucosa KF247232.1 Gram stained. Magnification: 100x.



Stenotrophomonas maltophilia LN890169.1 Gram stained. Magnification: 100x.



Pseudomonas aeruginosa KR815846.1 Gram stained. Magnification: 100x.



Leclercia adecarboxylata KT899848.1 Gram stained. Magnification: 100x.



Enterobacter asburiae EU239468.1 Gram stained. Magnification: 100x.



Candidatus roseomonas massiliae KT321690.1 Gram stained. Magnification: 100x.



Pseudomonas parafulva KT758848.1 Gram stained. Magnification: 100x.



Bacillus licheniformis KU314515.1 Gram stained. Magnification: 100x.



Pseudomonas aeruginosa KF680991.1 Gram stained. Magnification: 100x.



Agrobacterium larrymoorei EF178437.1 Gram stained. Magnification: 100x.



Pantoea septica KF475883.1 Gram stained. Magnification: 100x.



Stenotrophomomans rhizophila KP050794.1 Gram stained. Magnification: 100x.



Citrobacter freundii KP050794.1 Gram stained. Magnification: 100x.



Pseudomonas frederiksbergensis EU373369.1 Gram stained. Magnification: 100x.

Figure 4.1: Nutrient Agar plates showing cultured bacteria isolated from various used toothbrushes, and also microscopy images showing the bacteria under the light microscope.

Enterobacter asburiae

This bacterium is Gram-negative, non-motile and known to cause necrotizing fasciitis and pneumonia. Some *E. asburiae* isolates are identified as human pathogens which are an opportunistic pathogen and causes different human diseases such as community-acquired pneumonia, soft tissue infections, wound infection and other infections (Brenner *et al.*, 1986).

Pseudomonas aeruginosa

Pseudomonas spp., notably *P. aeruginosa*, act as opportunistic pathogens particularly of the lower respiratory tract, bloodstream, and urinary tract amongst patients in intensive care units or which are immunocompromised. It is often isolated from moist areas within health care environments, such as taps, toilets, showers, respiratory equipment, and cleaning solutions. Skin, throat, and faecal carriage amongst healthy individuals have also been reported (Liu, 2011).

Pseudomonas parafulva

It is a Gram-negative environmental bacterium, frequently isolated from rice paddies. Colonies on nutrient agar were smooth, entire, and flat to convex and had a water-insoluble yellow pigment after an incubation of 24h at 35°C. This bacterium rarely causes acute meningitis (Ramirez *et al.*, 2010).

Pseudomonas frederiksbergensis

This bacterium is an environmental organism of no known pathogenicity.

Roseomonas mucosa

Roseomonas species are a recently typified group of pink, slimy, waterborne, Gram-negative coccobacilli. They typically cause fever associated with persistent catheter colonization, and *R. mucosa* has been isolated from various clinical samples, including blood samples, samples obtained from wounds, and samples obtained from the aquatic environment (Bard *et al.*, 2010).

Stenotrophomomas maltophilia

This bacterium frequently colonizes damp surfaces such as tubes used in mechanical ventilation, endoscopes, indwelling urinary and suction catheters. In immunocompetent patients, *S. maltophilia* is a relatively rare cause of pneumonia, urinary tract infection, or bloodstream infection; however, is a growing source of latent pulmonary infections. *S. maltophilia* colonization of with cystic fibrosis patients has been seen to be increasing (Brooke, 2012).

Stenotrophomomas rhizophilia

This is a soil and rhizosphere organism of no known pathogenicity. Cells are straight or slightly curved yellowish rods. Strains are plant associated and isolated

from the rhizosphere of oilseed rape and from the rhizosphere and tubers of potato (Wolf *et al.*, 2002).

Leclercia adecarboylata

L. adecarboxylata is a motile, Gram-negative rod. It is considered as the sole pathogen causing infection in an immunocompromised woman (Hess *et al*, 2008). It is considered as normal flora in the gut of animals and it has been isolated from human stools, from the skin of an asymptomatic blood donor and from a variety of environmental sources and drinking water. *L. adecarboxylata* have the ability to infect a variety of bodily fluids; it has been cultured from blood, sputum, peritoneal fluid, urine, synovial fluid, gallbladder tissue, cardiac valve tissue and wounds often as a part of mixed microbial growth in immunocompetent hosts (Tam and Nayak, 2012).

Agrobacterium larrymoorei

This is a soil organism of no known pathogenicity.

Citrobacter freundii

Citrobacter freundii is a Gram-negative, motile, facultative anaerobic bacterium that appear as rods or coccobacilli. *C. freundii* is often the cause of significant opportunistic infections. Also, it has been associated with neonatal meningitis and brain abscesses and with nosocomial infections in the respiratory tract and causes

pancreatic pseudocyst after an acute necrotizing pancreatitis (Badger *et al.*, 1999). It has been distributed in water, soil, food and the intestinal tract of humans. Urinary tract infections (UTIs) caused by *Citrobacter* species have been described in bacterial urine isolates in adults (Metri *et al.*, 2013).

Candidatus roseomonas massiliae

This is an organism of no known pathogenicity.

Pantoea septica

Cells are Gram-negative, motile by peritrichous flagella. *Pantoea* species are recovered from humans and are opportunist pathogens associated with contaminated catheters and penetrating trauma. *Pantoea septica* referring to the septicaemia outbreak associated with these strains (Brady *et al.*, 2010).

4.1.4 Discussion

Tooth brushes can be heavily infected with microorganisms, especially Streptococci, within a day's use and a lack of toothbrush disinfection and care promotes the spread of such pathogens, thereby causing inflammation of the oral tissues (Badger *et al.*, 1999).

Out of the fifty toothbrushes used for this study, none was found to be uncontaminated with bacteria. *Pseudomonas* species were found on four of the fifty toothbrushes, *Stenotrophomomnas sp.* and *Bacillus sp.* were found also. A wide variety of bacteria belonging to different species were isolated from the investigated toothbrushes (Table 4.1).

Contaminated toothbrushes are recognized as a mode for microbial transport, and growth, and can result in the reinfection of a person with pathogenic bacteria or environmental microorganisms. The head area of tooth brush is prone to heavy contamination as fluids and food particles can be drawn by capillary action into the spaces between tufts, a process which may lead to bacterial growth (Frazelle and Munro, 2012).

4.2. Scanning electron microscope studies of the surface of a used toothbrush

4.2.1 Materials and Methods

Scanning electron microscope studies were conducted on the surface of bristles of tooth brushes undergoing normal use. A toothbrush used by the same individual for 6 weeks (alternating night and morning) was processed for scanning electron microscopy (SEM). Bacteria were fixed by immersing the brush head in 2.5% glutaraldehyde in 0.1 M sodium cacodylate buffer, pH7.3, for 60min. Brushes were rinsed in the same buffer, dehydrated in ethanol and critical point dried using liquid CO₂. A segment of brush head with one row of bristles was cut using a rotary saw, mounted on aluminium stubs, gold sputter coated and examined using a JEOL JSM-5300lv scanning electron microscope at an accelerating voltage of 10–30 kV (Sammons *et al.*, 2004).

4.2.2 Results and Discussion

Scanning electron microscopy of biofilms on toothbrushes used for approximately 3 months showed collections of cocci and the development of biofilms on the heads and bristles of both (Figure 4.2). The bristles were rough, providing ample sites for trapping organisms. Examination of a brush that had been used for more than 3 months revealed a biofilm on the brush head. The biofilm showed on the surface of the head to be made up of a compacted community of microorganisms, including cocci, bacilli and filamentous organisms, together with cellular and other debris.

SEM is showed biofilms on the heads and bristles of both conventional brushes, showing that bacteria colonized and grew on them. The rough surfaces of the bristles provide ideal sites for the entrapment of microorganisms and later development of biofilms, whilst nutrients will tend to accumulate at the base of the bristles thereby favouring the development of biofilms. Most dental practitioners recommend that brushes be changed after 2–3 months and following any bout of illness. The example shown here, of a brush that had been used in excess of five months, showed that after such a long period of use, extensive biofilms consisting of mixed communities of organisms developed at the base of the bristles and extended into the head of the brush. This process which could lead to the formation of a reservoir of organisms which can act as a potential source of infection for any vulnerable patient (notably where immunocompromised) or act as a source of cross-infection.



Acc.V Spot Magn Det WD Exp100 W 30 500xSE 160 1BMS Sheffield50 µmAcc.V Spot Magn Det WD Exp100 W 30 4000xSE 160 1BMS SheffieldFigure (4.2): Scanning electron microscopy of toothbrush biofilms on antibacterialbrushes used for approximately 3 months. Top left) showing debris and coccibacteria 8000 ×, bar represents 2µm; top right) showing debris and biofilm;

bottom left) image of tip of bristle, showing indentations and crevasses $500 \times$, bar represents 50 µm; bottom right) cocci embedded in the biofilm, $4000 \times$ bar represents 5 µm.

4.3. Effect of toothpastes on the bacterial growth

4.3.1 Introduction

Standard toothpastes are usually made up of a mix of fluoride and detergents both of which are generally thought to improve biofilm-control. Surprisingly, however, many toothpaste-based antimicrobials have yet to be effectively tested or proven to work in the oral environment.

Fluoride, in marketed dentifrice is usually comprised of sodium fluoride (NaF) which is a widely recognized anti-caries agent and NaF-containing oral hygiene products which significantly reduce dental caries (Barboza-Silva *et al.*, 2005). Fluoride inhibits a variety of bacterial processes which are meditated by enzyme binding (Barboza-Silva *et al.*, 2005). The main urease producers are *A. naeslundii* and *Staphylococcus epidermidis* while *S. epidermidis* is often the most ureolytically active organism present in plaque. Fluoride directly inhibits bacterial ureases (Barboza-Silva *et al.*, 2005), while chlorhexidine is bacteriostatic and bactericidal against both Gram-positive and Gram-negative microbes (based on its ability to damage cytoplasmic membranes and disrupt cell membrane integrity). Microbial contamination can be effectively removed by immersing them on chlorhexidine overnight.

4.3.2. Materials and Methods

In order to determine the antibacterial effect of toothpastes (fluoride and chlorhexidine toothpastes) a toothpaste agar was used containing 100 ml bacteriological agar and 4 g of a proprietary toothpaste in 9 laboratory bottles. The basal medium used for isolation and growth for bacteria was bacteriological agar which is composed of (g/l): 70% agarose and 30% agaropectin, final pH 5.7-7.0 at 33-36°C. Suspend 23 g of bacteriological agar in 1000 ml of distilled water. The bacteriological agar was boiled to dissolve the medium completely and sterilized by autoclaving at 15 lbs. pressure (121°C) for 15 minutes. The melted agar was poured in half of Petri plates and left them to be cool and set. For preparation the toothpaste agar: bacteriological agar 50 ml was dissolved in 9 Duran laboratory bottles and sterilized by autoclaving, then added 4 gram of Commercial toothpastes in the bottle (Figure 4.3). The bottles were shacked and vortexed at low speed until complete mixing was achieved. The mixed toothpaste agar was poured in the other half of plates and left them to be cool and set. The plates were streaked by model strains of Staphylococcus aureus and Escherichia coli in both sides of plates over the medium. All plates were incubated in the 37°C for 18-24 hours. After incubation, an ellipse of inhibition was used to determine the effect of toothpaste on growth of bacteria.


Figure 4.3: (left) The commercial toothpastes and (right) chlorhexidine toothpaste all used to prepare toothpaste agar plates.



Figure (4.4 A) Half agar plates containing fluoride-containing commercial toothpastes inoculated with bacteria.



Figure (4.4 B) Half agar plates containing chlorohexidine commercial toothpastes inoculated with bacteria.

4.3.3. Results and Discussion

The results showed that the bacteria isolated from toothbrushes can grow on the agar only side, but no bacteria grew on the toothpaste-containing agar. This result proves that toothpaste acts as an antibacterial agent and can inhibit bacterial growth. The main reason for inhibition is the presence in toothpaste of fluoride which is widely used as an effective anti-caries agent (i.e. antibacterial) agent. Marquis (1995) showed that fluoride can affect bacterial carbohydrate metabolism and that it inhibits the functioning of essential bacterial enzymes; it also reduces the acid tolerance of bacteria and thereby prevents bacteria growth. Fluoridated

toothpastes showed antibacterial activity against *S. mutans* and both aerobic and anaerobic oral flora and also inhibited Streptococci selectively, and inhibited caries formation by interfering with biofilm development by Streptococci (Randall *et al.*, 2015).

Chlorhexidine toothpastes also inhibited bacterial growth when added to toothpaste agar plates. De Rossi et al. (2014), showed that chlorhexidine toothpastes presented antimicrobial activity against Gram-positive bacteria and yeasts and that the chlorhexidine molecule has both bactericidal and bacteriostatic antimicrobial effects on the tooth surface. In the present study, chlorhexidine was found to be effective in disinfecting contaminated toothbrushes, although in other studies Listerine was shown to be more effective. The higher efficacy of chlorhexidine could be due to its extended action-spectrum. It is also relatively non-toxic, odourless and is a commonly used mouthwash, properties which make chlorhexidine a good choice for the disinfection of contaminated toothbrushes. In conclusion, the results presented in this Chapter show that all toothpastes exhibited effective antimicrobial activity against the tested bacteria and that these products were able to inhibit the growth of bacteria. In addition, adding chlorhexidine to dentifrices can result in effective antimicrobial activity against all the evaluated Gram-positive bacteria and yeasts. However, further studies are required to evaluate their clinical advantages in the treatment or prevention of biofilm-mediated diseases.

4.4 Discussion

Again, the importance of these isolates is not that they are major pathogens, but that they can infect immunocompromised patients (Ankola *et al.*, 2009). Toothbrushes get re-contaminated after each use (Frazelle and Muro, 2012) and such re-contamination of the oral cavity can result from the retention of microorganisms on toothbrushes (Filho *et al.*, 2000, Karibasappa *et al.*, 2011). Nascimento *et al.* (2012) showed that new toothbrushes may often carry bacteria even before their use, since no regulatory requirement for pre-use sterilization is generally required. This observation was confirmed in our study, where all (100%) of new tested brushes were contaminated positive for bacterial growth, a finding which explains the high incidence of *Bacillus cereus* and *Candida albicans* on toothbrushes and oral swabs.

The effectiveness of toothbrushes at cleaning, and bacterial plaque removal largely depends on their abrasive qualities, although some toothpastes also contain antibacterial agents (which are often general sterilants) such as hexachlorophene. Toothpastes also contain a combination of fluoride and detergents, compounds which increase their efficiancy (Davies, 2008; Marsh, 2010; Prasanth, 2011); triclosan for example, is a low-toxicity, non-ionic, chlorinated bisphenol that is compatible with toothpaste additives like fluoride and surfactants, and it enhances the inhibition of cyclooxygenase/lipoxygenase pathways and also exhibits anti-inflammatory properties (Davies, 2008; Davies *et al.*, 2010). Chlorhexidine is usually regard as the most effective antimicrobial agent for use in dentistry (Jones, 1997; Twetman, 2004). Its effectiveness is due to the dicationic nature of the

chlorhexidine molecule, which leaves a lasting antimicrobial effect on the tooth surface (Twetman, 2004). Compounds derived from plants also have useful antimicrobial properties which are relevant to toothpaste use (Verkaik, 2011; Pannuti, 2003). A toothpaste called Parodontax for example, is an herb-based product which contains sodium bicarbonate and a number of herbal extracts such as chamomile *Echinaceas* and *Mentha piperita* (Panuti, 2003). For obvious reasons, the use of easily utilizable sugars is avoided as additives to toothpastes, although carbon substances which can support microbial growth are always present in moist and dry sputum.

The results presented show that tooth brushes are contaminated with potentially pathogenic bacteria and that toothpastes act as a microbial nutrient source. As a result, it is highly desirable that only sterile, one time-use tooth brushes should be used by immunocompromised patients. Although modern toothpastes are generally free of readily utilizable sugars, they provide microbial substances in the form of dried sputum and food particles. The antibacterial effect of toothpastes was determined using a readily available family and thus the results showed that pathogenic bacteria failed to grow on bacteriological agar containing toothpastes. The most likely explanation for the inhibitory effect is that the bacteria were inhibited by fluorides which are added to toothpastes as anti-carries agents. Some toothpaste also contains more specific biocides which play a role, alongside

fluorides in inhibiting potentially pathogenic bacteria by inhibiting carbohydrate metabolism and inhibiting essential enzymes (Marquis, 1995).

CHAPTER 5

EFFECTIVENESS OF ANTIBACTERIAL CLOTHS IN INHIBITING THE GROWTH OF BACTERIA

5.1 Introduction

The silver ion has long been known to be effective in inhibiting or killing a broad range of microorganisms and silver is increasingly being used to control bacterial growth in a variety of medical applications, including dental work, catheters, and in the healing of burn wounds (Jung et al., 2008). Slow-release "nanosilver" linings are also used in laundry machines, dishwashers, refrigerators, and toilet seats. The mechanism of the antimicrobial action of the silver ion relates to its interaction with thiol (sulfhydryl) groups. Amino acids, such as cysteine, and other compounds containing thiol groups, such as sodium thioglycolate, neutralize the activity of silver against bacteria (Jung et al., 2008). Silver affects bacterial enzymes and brings about a marked inhibition of bacterial growth with elemental silver often being deposited in the vacuole and cell wall as granules. This element also inhibits cell division, damages the cell envelope, and can denature the contents of the bacterial cell (Jung et al., 2008). Under the influence of silver ions, bacterial cells increase in size, and the cytoplasmic membrane, cytoplasmic contents, and outer cell layers all exhibit structural abnormalities. Finally, silver ions interact with nucleic acids and interact preferentially with the bases in DNA rather than with the phosphate groups (Jung et al., 2008).

5.2 Materials and Methods

The aim of the following experiments was to determine the effect of antibacterial e-cloth (EnivroProducts Kent, TN3 8LE) and a control microfiber cleaning cloth on bacterial contamination (Figure 5.1).

Two types of silver impregnated cloths (antibacterial e-cloth "red colour" and a non-antibacterial microfiber cleaning cloth "yellow colour") were applied to environmental surfaces (desks, air conditioner, windows, medical equipment, laboratories). Pieces of cloth were moistened with sterile water and wiped firmly over the entire surfaces. Two approaches were used. In the first method, silver containing cloths were used for isolation of various bacterial contaminants from environmental surfaces. Sterile water moistened pieces of cloths were wiped firmly over the entire surfaces. Half the number of cloth was placed in 50 ml of Nutrient broth in sterile tubes, and vortexed for one minute and was left in the shaker for overnight. After 18-24 hours of inoculation, the Nutrient Broth medium became turbid, the turbidity ensuring the presence of certain bacteria in the samples. The other half was placed on the surface of nutrient agar plates and the plates left in the incubator for 18-24 hour. After incubation, the presence or absence of bacterial growth was noted. For morphological identification of bacterial colonies, different types of bacterial colonies appeared on the Nutrient agar plates. Pure colonies of isolates were identified and characterized using standard microbiological techniques. The second method involved using silver containing cloths to determine the effect of silver as an antimicrobial on growth of

bacteria. Sterile water moistened some cloths were placed on the centre of the medium and streaked with cultures of *Staphylococcus aureus* and *Escherichia coli* (i.e. on both sides adjacent to the cloth). These plates were incubated at 37°C for 18-24 hours. Sterile water moistened cloths were placed on the centre of the Sabouraud Dextrose agar plates and streaked with cultures of *Candida inconspicua* and *Candid rugose* then incubated at 25°C for 5 days. After incubation, the size of any resultant inhibition zone was measured.



A) e-cloth

B) microfiber cleaning cloths

Figure 5.1: A) The antibacterial e-cloths and B) non-antibacterial microfiber cleaning cloths.

5.3. Results

5.3.1: Isolation of various bacterial contaminants from environmental surfaces using silver containing cloths on nutrient agar plates and nutrient broth.



Figure 5.2: Bacterial growth around the antibacterial and control cloth pieces after isolation from different surfaces.



Figure 5.3: Growth of bacteria in the nutrient broth inoculated with the antibacterial and control cloths pieces after isolation from different surfaces.

5.3.2 Examination of the effect of silver as an antimicrobial on growth of bacteria



Figure 5.4: No inhibition zones appeared around the antibacterial cloths and the control. The bacteria strain used is *E. coli*; yellow is the control cloth and red is the tested antibacterial cloth.



Figure 5.5: No inhibition zones appeared around the antibacterial cloths and the control. The bacteria strain used is *S. aureus*; yellow is the control cloth and red is the tested antibacterial cloth.

5.3.3 Examination of the effect of silver as an antimicrobial on growth of Candida

sp.



Figure 5.6: No inhibition zones appeared around the antibacterial cloths and the control. The fungal strain used is *Candida inconspicua*; yellow is the control cloth and red is the tested antibacterial cloth.



Figure 5.7: No inhibition zones appeared around the antibacterial cloths and the control. The fungal strain used is *Candida rugose*; yellow is the control cloth and red is the tested antibacterial cloth.

5.4. Discussion

The antibacterial effects of silver containing cloths against bacteria was determined using two techniques with two types of cloths, one an antibacterial cloth containing silver and a non-antibacterial cloth as control. The first method indicated that the growth of bacteria was heavy and there was no effect of silver on bacteria growth. There was no significant difference between the growth resulting from cloths containing silver and control cloths. Both showed the same results (Figure 5.2, 5.3). The bacterial growth was not affected by bactericidal activity resulting from silver.

The silver impregnated cloth used here was wiped on various surfaces and, not surprisingly, became contaminated with dust and dirt and, when this was transferred to growth medium, it supported bacterial growth. The antibacterial cloth did not prevent the growth of bacteria when incubated adjacent to the organisms on a solid growth medium. This result is entirely unexpected, since the cloth used here is marketed as an antibacterial material based on its silver content and its claimed ability to act as a broad spectrum micro-biocide capable of inhibiting bacteria and fungi including MRSA, and other antibiotic resistant species (Gupta and Chauhan, 2016). It is likely that the concentration of silver present in the cloth is too low to be capable of inhibiting bacteria and it appears that the antibacterial cloth, used here, would be of little use for controlling pathogens on surfaces in household and hospital settings.

The second method showed that no inhibition zones appeared around the antibacterial cloths, so the growth of bacteria was not affected by the sliver on cloths as shown in (Fig 5.4 ,5.5). Also, the antibacterial cloth did not prevent the growth of two yeasts when they were incubated adjacent to the material on a solid medium (Figure 5.6, 5.7). This result is both surprising and worrying, since it shows that cloths which are advertised and sold to kill bacteria do not do so. The result may be due, however, to the fact that heavy bacterial loads were used in this study. Further work will continue using lower loads.

It is known that the enhanced antibacterial effect of nanoparticles is due to their large surface to volume ratio, and therefore the smaller the particle, the greater this effect would be. Yu *et al.* (2013) reported that nanocomposites of larger sized AgNPs were much less cytotoxic than the smaller ones, without sacrificing the antibacterial potency of smaller particles.

The mechanism behind the antimicrobial potency of Ag nanoparticles is still not clear. It is believed that their antibacterial efficacy could stem from their absorption by bacterial cells, resulting in the shrinkage of the cytoplasmic membrane (Pattabi *et al.*, 2013). An alternate mechanism proposes that due to the interaction of Ag ions with the S-H bonds in proteins, the DNA loses its capacity to replicate and proteins become deactivated (Pattabi *et al.*, 2013).

CHAPTER 6

SURVIVAL AND RELEVANT METABOLIC DIVERSITY OF ISOLATES 6.1 Introduction

Bacteria play a major role in the cycling of elements in the outdoor and built environments, being involved in all of the major cycles, including participating in transformations of C, N, P, S as well as modifying elements such as copper, manganese and iron (Killham, 1994). The aim of the work reported below was to determine the ability of some of the bacterial isolates to hydrolyse urea, nitrify, oxidize reduced form of sulphur, solubilize insoluble phosphate and oxidise copper and iron sulphides. Some of these transformations are particularly relevant to healthcare environments, such as the hydrolysis of urea (present in urine) or the oxidation of copper and iron sulphides which are found in corroded hospital, metal pipes. Other transformations, such as the formation of thiosulphate may also enable bacteria to better survive in the medical environment by protecting them from the toxic effects of, for example, heavy metals, or even possibly biocides. The following is a brief survey of the transformations which occur in mineral cycling which are relevant to the work described here. Further details on these transformations can be found in the following references: Alexander (1977), Maier et al. (2009), Killham (1994) and Paul and Clark (1989).

Urea hydrolysis: Urea is water soluble and has a high nitrogen content exceeding that of ammonium nitrate and ammonium sulphate. Microbes secrete

ureases which converted urea to carbon dioxide and ammonia. A wide range of bacteria can mediate this process including species of *Pseudomonas*, *Achromobacter, Bacillus* and *Micrococcus*.

Nitrification: Nitrification is of major importance for the N-cycle in aquatic and terrestrial environments. It involves the oxidation of ammonium (NH₄⁺) to nitrite (NO₂⁻) and then nitrite to nitrate (NO₃⁻) by chemoautotrophic bacteria and by some heterotrophic fungi and bacteria, which can also perform these oxidations. Two kinds of nitrification exist (Killham, 1994): The first involves the activity of chemoautotrophic nitrifying bacteria (*Nitrosomonas*) by which ammonia (NH₃) or ammonium (NH₄⁺) ions are oxidised to nitrite (NO₂⁻). The second part of the process involves chemoautotrophic Gram-negative bacteria which oxidize nitrite (NO₂⁻) to nitrate (NO₃⁻). *Nitrobacter* and some fungi such as species of *Penicillium* and most other *Deuteromycetes* can also perform these oxidations (Maier *et al.*, 2009).

Sulphur oxidation: Sulphur is an essential element for growth of all organisms, being a required element for the synthesis of the amino acids, cysteine and methionine, and vitamins such as vitamin B1 (thiamine), hormones such as biotin, coenzymes and lipoic acid. Filamentous fungi play a role in the S- cycle; for example, *Fusarium solani* (a soil fungus) oxidizes elemental sulphur to polythionates, thiosulphate and sulphate. Fungi oxidize sulphur to sulphate with the formation of tetrathionate and thiosulphate. These products, it has been

suggested, may protect fungi from the toxic effects of heavy metals (Alexander, 1977).

Phosphorus solubilisation: Bacteria and fungi are able to solubilise phosphate and release P when growing *in vitro* with calcium phosphate, apatite or other sources of insoluble phosphate; phosphate solubilizing fungi include species of *Aspergillus, Fusarium,* and *Penicillium* (Paul and Clark, 1989). A wide variety of heterotrophs solubilize insoluble phosphate including tricalcium phosphate, dicalcium phosphate, hydroxyapatite, and rock phosphate (Altomare, 1999). Bacteria which participate in the reactions include: *Microccocus, Aereobacter, Flavobacterium, Erwinia Pseudomonas, Bacillus, Rhizobium, Burkholderia, Achromobacter* and *Agrobacterium* (Kim *et al.,* 1997,1998). The mechanisms of solubilisation of insoluble phosphate are based on acidification of the medium following organic acid production.

Microbial oxidation of metal sulphides: A wide range of bacteria and fungi have the ability to oxidize metal sulphides, some being chemoautotrophs such as members of the genus *Thiobacillus*, while others are common heterotrophs. The final product of the oxidation reactions involved is sulphate which occurs as sulphuric acid (Killham, 1994). This sulphuric acid can be corrosive to metals, but on the positive side, can be used in ore leaching to obtain metals.

The biogeochemical processes mentioned above and studied here have generally been associated with chemolithotrophic bacteria and the potential role of

heterotrophs in their mediation has generally been underestimated, but are increasingly being shown to play a crucial role in environmental cycling (Killham, 1994). Heterotrophic nitrification and S-oxidation may not benefit organisms in relation to direct end product formation, but through the indirect benefits gained by a heterotroph from participating in these processes. The production of polythionates during S-oxidation by fungi, for example, may help protect these organisms from the toxic effects of heavy metals and other toxicants.

6.2 Materials and Methods

Growth of bacteria and analysis of ions

The bacteria were grown on nutrient agar plates and inoculated into 10 ml of Nutrient Broth containing 0.1% w/v substrate in sterile Falcon Tubes (100ml); the tops were loosened to allow for gas exchange and then incubated with vigorous shaking for 7 days at 37°C. The contents were then allowed to settle and an aliquot was transferred to a HP-1050 DAD HPLC SYSTEM for analysis of resultant ions, or dipped with the relevant ion dipstick (Figure 6.1).



Figure 6.1: (A). Dipsticks container with dipsticks used for ion determination showing concentration chart, (B), Dipsticks used for ion determination showing concentration chart set used for determination of phosphate.

6.3 Results and Discussion

A comment on Dipsticks as a method for ion analysis. The measurement of ions (which are important in environmental transformations) in this laboratory (such as nitrate, phosphate and sulphate) has generally involved the use of colorimetry. These approaches have worked extremely well and, as result have proved to be very useful; of late however, it has been turned to the use of analytical-ion Dipsticks which are less expensive and are generally less- influenced by interference. They also do not involve the preparation and use of dangerous chemicals (for example when they are used to replace corrosive chromotropic acid in the analysis of nitrate) and can be used to rapidly screen a wide number of samples. This replacement of chromotrophic acid with Nitrate-Dipsticks provides a good example of the advantages in changing this analytical approach. This analytical reagent is based on concentrated sulphuric acid (Simms and Jackson,1971) and is therefore highly corrosive and is dangerous to prepare and use. Its use as an analytical reagent is also hindered by the fact that, as well as nitrate, it reacts with sugars, when these are present to give a pink-purple colour which interferes with the reading of the normal yellow colour produced by the interaction of chromotropic acid with the nitrate ion. Because no acids are used when nitrate-ion Dipsticks are used, and because no colour interference occurs using this approach, being quicker and cheaper in the long run, is regarded as an improvement on the use of chromotropic acid.

The only disadvantage with using Dipsticks is that some accuracy is sacrificed. In most cases, however, all that is needed from a nitrate ion analysis is a rapid indication of whether or not the ion in question is present or a relatively accurate estimate of its concentration, both of which can be provided using Dipsticks. Tests, by others in this laboratory have shown that results from Dipsticks are between 5 percent (plus or minus) of the values achieve using colorimetry, as was previously used.

The results show that all of the bacteria hydrolysed urea to ammonium and were able to oxidize ammonium to nitrate, via nitrite (Table 6.1). The most active urea hydrolyzer was isolate 7, followed by 5 and 4, while isolate 10 showed the least activity in this respect (Table 6.1). All isolates also oxidized ammonium to nitrate

with the formation of small amounts of nitrite as intermediates. Since the oxidation of nitrite to nitrate is rapid, the former ion rarely appears at high concentration in the environment (Killham, 1994). In terms of nitrate production, bacterium 7 was the most active, followed by 10 and 5, while bacteria 1 and 2 were the least active (Table 6.2).

Substrate-Urea	Ammonium (µgml ⁻¹)		
	1 27.5 (1.9) 30		
	2 25.3 (2.7) 15		
	3 26.7 (0.9) 25		
	4 30.6 (3.6) 30		
	5 43.6 (5.2) 50		
	6 13.1 (2.1) 15		
	7 50.9 (2.3) 60		
	8 10.8 (8.6) 5		
	9 17.3 (1.9) 20		
	10 4.4 (0.2) 10		

Table 6.1. Hydrolysis of urea to ammonium.

 Bacillus licheniformis, 2) Bacillus subtilis, 3) Bacillus thuringiensis
Enterococcus mundtii, 5) Citrobacter freundii, 6) Pseudomonas luteola
Arthrobacter sanguinis, 8) Klebsiella oxytoca, 9) Kocuria rhizophila, 10) Rothia amarae.

Figures in **bold** relate to Dipstick analysis. Means of triplicate flasks (+/- standard

deviation), 7day incubation at 37°C, values in excess of control value.

Table 6.2.	Oxidation	of an	nmonium t	to nitrite	and nitrate.
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Substrate-Urea	Product (µg ml ⁻¹)		
Nitrite	Nitrate		
1. 1.8. (1.9) All- trace	5.2 (0.66) 8		
2. 0.7 (0.1)	3.2 (0.8) Trace		
3. 0.9 (0.9)	11.9 (1.2) 10		
4. 1.4 (0.60)	15.9 (2.0) 20		
5. 0.3 (0.0.2)	30.2 (0.9) 25		
6. 1.1 (0.1)	10.2 (0.1) 15		
7. 0.5 (0.2)	50.3 (07) 35		
8. 0 (0.1)	7.9 (1.3) 10		
9. 1.9 (0.1)	20.9 (1.7) 20		
10. 1.4 (0.2)	30.2 (5.4) 50		

 Bacillus licheniformis, 2) Bacillus subtilis, 3) Bacillus thuringiensis
Enterococcus mundtii, 5) Citrobacter freundii, 6) Pseudomonas luteola
Arthrobacter sanguinis, 8) Klebsiella oxytoca, 9) Kocuria rhizophila, 10) Rothia amarae.

Figures in **bold** relate to Dipstick analysis. Means of triplicate flasks (+/- standard deviation), 7day incubation at 37 ^oC, values in excess of control value.

All of the isolates were able to solubilize insoluble phosphate, notably isolate 7

followed by isolate 5, with isolate 10 being the least effective. Similarly, all

isolates were able to oxidize iron sulphide to release Fe^{2+} and oxidize copper to

sulphide to release Cu^2 + ions. All of the isolates, with the exception of isolate 2,

released similar amounts of Fe $^{2+}$, while isolate7 and 5 were particularly active at oxidizing copper sulphide (Table 6.3).

Substrate-		Product (µg ml ⁻¹)	
Insoluble-P to	Phosphate	FeS to Fe ² +	CuS to Cu ² +
1 26.5 (1.3)	30	3.1 (0.9) All- trace	4.2 (1.6) 4
2 5.3 (2.7)	15	1.7 (0.2)	3.6 (1.8) 4
3 4.7 (1.9)	20	4.9 (1.9)	5.9 (1.2) 7
4 25.6 (4.6)	30	4.4 (1.6)	14.6 (1.8) 10
5 45.6 (4.2)	40	3.0 (1.2)	23.2 (1.9) 20
6 18.1 (1.1)	10	5.0(0.5)	10.4 (1.1) 10
7 45.9 (1.3)	50	3.5 (0.2)	25.3 (2.7) 15
8 20.8 (8.5)	15	4.0 (0.9)	17.9 (1.5) 20
9 15.3 (1.6)	15	5.0 (0.2)	12.9 (1.7) 10
10 6.4 (1.2)	5	2.4 (2.2)	12.2 (1.4) 30

Table 6.3. Solubilization of insoluble P and oxidation of FeS and CuS.

 Bacillus licheniformis, 2) Bacillus subtilis, 3) Bacillus thuringiensis
Enterococcus mundtii, 5) Citrobacter freundii, 6) Pseudomonas luteola
Arthrobacter sanguinis, 8) Klebsiella oxytoca, 9) Kocuria rhizophila, 10) Rothia amarae.

Figures in **bold** relate to dipstick analysis. Means of triplicate flasks (+/- standard deviation), 7day incubation at 37°C, values in excess of control value.

The results obtained using proprietary Dipsticks were in most cases similar enough to this obtained using HPLC to point to their use in experiment where an exact, but reasonably close figure for an ion-concentration is required. As mentioned above, this loss of accuracy is acceptable for convenience, low cost and safety of using Dipsticks instead of the colorimetric assay. The results show that the bacteria isolated from healthcare environments are capable of participating in some important metabolic process, whether such participation is relevant to such environments, especially where carbon is limiting, is not immediately clear. Despite this, the experience of conducting these experiments was, in any case, highly valuable.

CHAPTER 7

FINAL DISCUSSION

The work provided in this Thesis clearly shows that bacteria and other microbes commonly contaminate everyday objects and surfaces, a fact which does not generally cause problems in normal life (except for the occasional food poisoning episode). Such contamination does, of course, have major implications in health care settings such as hospitals and in particular for immunocompromised patients. These patients have their immune systems inhibited either intestinally (e.g. during organ transplantation), or as the result of a natural intervention, such as AIDS. The microbes which cause these infections are often generally not pathogenic and the immune systems of non-immunocompromised patients can generally cope with them, even when the patient is otherwise ill. The stark fact is that any microbe can act as pathogen in people with a compromised immune system. It is therefore extremely important to make certain that all surfaces in health care settings are as clean and microbe-free as possible and that this hygiene status includes non-pathogens as well as the generally recognized pathogens. Such a relatively microbe-free environment can be achieved using biocides, but there is no substitute for thorough "deep cleaning".

The cleaning of hospital surfaces is the main defence against the threat of antibiotic resistant bacteria and nosocomial infections. While the importance of deep cleaning is universally recognized, this does not mean that such cleaning programmes are theoretically effective or always well-managed. Bleach is by far the most commonly used biocide in both hospital and domestic settings and, when used correctly, can be

extremely efficient and act as an effective standby for all disinfecting purposes. Although it is more expensive, hydrogen peroxide provides a useful alternative to bleach especially for use on so-called "non-critical surfaces" since it is fact acting and exhibits a broad spectrum antimicrobial activity against bacterial and fungal spores and viruses; steam vapour systems and microfiber- containing biocides can also be effectively employed for routine cleaning purposes.

An essential requirement for successful disinfection is the provision of an adequate contact time between a disinfectant and the object being disinfected. Such contact times vary in addition to a number of environmental factors, notably the degree of microbial loading. Pathogens, and other microbes, can be transmitted in a number of ways in the hospital environment, including:

Droplet contact transmission: A large number of diseases are spread inside droplets which make contact with the eye, nose or mouth. Examples of these droplet infections include SARS, the common cold, Legionnaires' disease and MRSA.

Such infected droplets are produced by infected persons when coughing, sneezing or talking.

Airborne Transmission: Suspended dust particles which contain microbes often remain in the air over long periods and from here can gain entry into the upper and lower respiratory tracts and thereby bring about infections like Aspergillosis, chickenpox and measles.

Faecal-oral transmission: Pathogenic and non-pathogenic microbes infect patients following the consumption of food and water which has been contaminated with

faeces. Bad hygiene and poor sanitation allow such microbes to contaminate food, water and environmental surfaces. These organisms then multiply inside the digestive system then released from the body in faeces and urine, so that the infection-cycle then begins again. Diseases which are disseminated in this way include: rotavirus, hepatitis A virus, *E. coli, Cryptosporidium, C. difficile, H. pylori* and *Candida*.

Direct contact transmission: This disease-transmission process involves the direct physical contact between an infected person and the new host; examples include, kissing and sexual intercourse, or merely close contact. Diseases which are spread by direct contact transmission include H1N1 virus and hepatitis A virus, *Acinetobacter*, *E. coli*, SARS, the common cold, ringworm and other yeast infections, scarlet fever, norovirus, foot and mouth disease, *H. pylori*, and MRSA.

Indirect contact transmission: This process of disease spread takes place when a susceptible person makes direct physical contact with the body surfaces of an infected person via their hands, and then touches their own face, eyes or mouth, allowing the pathogen to gain access into the body and initiate infection. Organisms which use this transmission route include: *rotavirus*, hepatitis A virus, influenza, the common cold, *H. pylori* and tuberculosis, norovirus, *C. difficile*, MRSA, SARS, *E. coli*,

Cryptosporidium, ringworm and other yeast infections and scarlet fever. The fact that these microbes can survive and be replenished on body surfaces for long periods demonstrates the obvious need for hand washing and other aspects of body hygiene amongst patients, staff and hospital visitors.

Use of disinfectants and detergents in healthcare settings

The above discussion demonstrates the clear need for effective sterilizing agents to reduce the pathogen load on surfaces in hospitals. The use of such agents varies based upon factors such as efficiency and cleaning ability, environmental impact, cost, and associated toxic effects.

Detergents

Detergents generally need effort in regard to rubbing and scrubbing to be fully effective but are good at removing chemicals, food spills, and other commercial wastes. They are not, however, generally suitable for use in cleaning blood or other body fluids from contaminated surfaces and are also usually not effective against bacterial spores, particularly those of *C. difficile*.

Sterilizing agents

Sterilizing agents are generally expansive and highly toxic and are therefore not used in routine household or day to day use. They are however, effective on so-called "critical surfaces" in hospital, industrial and laboratory settings. These agents are however, very effective at killing pathogens, including spores of *C. difficile*, and are particularly effective for use in disinfecting and removing blood and other contaminating body fluids; they include agents such as chemicals, heat, irradiation, filtration and high pressure sterilization.

Guidelines for use in the decontamination of healthcare associated surfaces

Consideration needs to be given to the sterilization of the numerous surfaces found in health care environments, simply because each site varies in its surface properties and the environmental conditions to which it is exposed. An obvious example is provided by the differing conditions found on the surface of an oven compared to that of a refrigerator.

Non-critical medical surfaces

These areas can be adequately served by routine, low maintenance, detergent-based cleaning which will prevent the transmission of microbes from non-critical surfaces, such as furniture and floors. The cleaning process should begin the removal of soil and other debris by wiping or deep scrubbing before a cleaning/disinfecting agent is applied. The survival of bacteria such as *Staphylococcus* spp., MRSA, *Acinetobacter*, and other airborne fungi is encouraged by the presence of dust on surfaces and dampdusting with cloths wetted with detergents recommended for use on such non-critical surfaces, followed by a period thorough drying. All objects within the ward or room including radiators, air conditioning units, fans, switches, sockets and of course computer areas should be thoroughly wiped with disinfectant-rich cloths. Doors, which include their edges, should be cleaned and special attention should be given to areas which are frequently touched such as handles and door-push plates. In order to control Legionnaires' disease, any infrequently used taps and shower heads should be run weekly for around 10 minutes and all air conditioning and ventilation grills and

associated extractors and inlets should be dusted at least weekly and should be fully cleaned on a yearly basis. Walls and ceilings however, need to be washed only every 6 months or so, by the use of hot water and detergent.

The following factors affect the choice and use of disinfectants. Attention should always be given to the most important factor in their use, namely contact–exposure time:

- The need to comply with the appropriate chemical safety regulations.
- Some disinfectants corrode or discolour surfaces, so compatibility should be considered and compatibility tests conducted.
- The required antimicrobial activity- this relates to the need to determine if the proposed has a wide antimicrobial spectrum?
- Contact times- the necessary contact time needed to kill nearly are hundred percent of the pathogens present needs to be determined by laboratory tests.
- Is the disinfecting agent effective when organic matter is present? This is a very important point because organic matter rapidly deactivates many otherwise potent disinfecting agents?
- Storage and disposal -Is the agent stable when stored for long periods? This is an important point because such agents are often bought in bulk to reduce costs and are stored for long periods; storage should of course be done at room temperature in the dark. Issues regarding the disposal of an agent and its environmental toxicity obviously need to be considered; some healthcare authorities may wish to impose ethical environmental tests on the sterilizing products they employ.

- Environmental factors- Do common environmental factors such as temperature and pH affect the disinfectant's effectiveness?
- Costs-Finally, cost is an essential factor especially since healthcare facilities worldwide are increasingly being effected by Government cost-cutting.

Disinfectants used in routine cleaning of healthcare environments

The following is a description of the most frequently used, EPA-registered disinfectants:

- Phenolic and iodophore-based germicides.
- Quaternary ammonium germicidal detergents.
- Hydrogen peroxide (3-7.5%).
- Ethyl or isopropyl alcohol (70-90%); sodium hypochlorite, i.e. bleach (5.25-6.15% household bleach diluted 1:500 gives >100 ppm available chlorine).

Environmentally safe disinfectants

So-called environmentally safe disinfectants include baking soda, vinegar, eucalyptus oil, grapefruit seed extract borax, liquid detergent, alcohol and tea tree oil. With the exception of undiluted vinegar and eucalyptus oil, most of these disinfectants do not kill *E. coli*.

Antibacterial coating of surfaces

Antimicrobial coatings are available for use on commonly used domestic fabrics, including, linen (curtains); furniture (chairs and tables); office equipment (computers and printers); hand-touch sites (handles and water taps) and generally used surfaces such as walls, floors, walls and doors. Any product that can be impregnated or coated with a micro-biocidal paint or chemical can referred to as being 'antibacterial'. Such bioactive surfaces or coatings generally provide heavy metals, antiseptics and biocide to the surface undergoing treatment. Nano-silver particles, like titanium dioxide, are particularly useful since they form an invisible protective nano-coating which can act as a very effective biocide for use on a wide range of surfaces, even under extreme temperatures.

CONCLUSIONS

A wide range of bacteria were isolated during the course of this study from computer keyboards, sinks, used toothbrushes and vacuum cleaner dust. Bacteria were also shown to be emitted by toilet hand dryers; these are becoming increasingly popular in public buildings because of reasons of cost and convenience. The results of work presented here shows, however, that paper towels should be used in healthcare environments instead of these dryers, simply because hand dryers can release potential pathogens into the surrounding air, into what is often a small, enclosed space, with little in the way of ventilation. Studies are also reported on the survivability of bacteria on smooth and rough tiles, essentially the same similar to those found in homes and

healthcare settings. Seemingly paradoxically, bacteria were shown to survive for longer on rough, compared to smooth tiles as well as on plastic plumbing fittings than on copper fittings. These findings confirm those of other works, by showing that copper fittings are desirable over plastic fittings because they dramatically reduce the survival of surface-growing, potential pathogens. The results presented here also show that bacteria are important nosocomial pathogens and, since they have been shown to be widely isolatable from the hospital and other indoor environments studied here, they will likely cause life-altering diseases in hospital specially for immunocompromised patients. Clearly the existence and survival in healthcare settings of these potentially pathogenic bacteria is a highly important research area which deserves continued investigation.

SUGGESTIONS FOR FURTHER STUDIES

The following important areas of research which have been examined in this Thesis need to be further studied: a) the survival of potentially pathogenic bacteria on tiles, b) the ways in which potentially pathogenic bacteria can survive on different types of piping used in plumbing in hospitals and c) the part played by hand dryers in distributing bacteria and other microbes inside the environment of hospitals and other healthcare facilities.

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APPENDIX 1

Publications Arising From This Thesis

Bacterial contamination of used manual toothbrushes and effects of toothpastes on isolated potential pathogens

Rana Abdulrahim Alaeq and Milton Wainwright

University of Sheffield, UK

Toothbrushes play an essential role in oral hygiene and are commonly found in community and hospital settings. The toothbrushes may act as a reservoir for potential pathogens transferred from the oral cavity and from the bathroom environment. The purpose of this study is to determine the bacterial contamination of used toothbrushes and determine the antibacterial effect of toothpastes. Scanning electron microscopy (SEM) was used to visualize biofilms on toothbrush bristles. 50 used toothbrushes obtained from volunteers were analysed bacteriologically using standard microbiological techniques. Bacteria present on all toothbrushes heads were cultured to determine the presence of bacteria and scanned by SEM. The antibacterial effect of toothpastes was determined using seven types of commercial toothpastes and chlorhexidine toothpaste by inoculation bacteria on the toothpaste plates. The result showed that all the toothbrushes were contaminated with the following bacteria: Roseomonas mucosa, Stenotrophomonas maltophilia, Pseudomonas aeruginosa, Leclercia adecarboxylata, Enterobacter asburiae, Candidatus Roseomonas massiliae, Pseudomonas parafulva, Bacillus licheniformis, Pseudomonas aeruginosa, Agrobacterium larrymoorei, Pantoea septica, Stenotrophomomnas rhizophila, Citrobacter freundii and Pseudomonas frederiksbergensis. The bristle surfaces, being rough, provided ample sites for trapping organisms. Examination of a brush revealed a biofilm on the brush head. The biofilm seen on the surface of the head to be composed of a compacted mixed community of microorganisms, including cocci, bacilli and filamentous organisms, together with cellular and debris. The toothpaste used proved antibacterial and inhibited bacterial growth, based mainly in the activity of fluoride which is widely used as an effective anticaries agent. In conclusion the isolated organisms are potentially pathogenic, particularly in relation to immunocompromised patients. The appropriate rinsing and drying of the toothbrushes before storage will however, likely reduce the incidence of these bacteria and the health risk associated with these pathogens.

Rana Abdulrahim Alaeq *et al.*, J Bacteriol Parasitol 2016, 7:6 http://dx.doi.org/10.4172/2155-9597.C1.026

7th World Congress on Microbiology, November 28-29, 2016 Valencia, Spain

Influence of copper and plastic surfaces on the survival of bacteria in relation to the health care environment

Rana Abdulrahim Alaeq and Milton Wainwright

University of Sheffield, UK

Metallic copper (Cu) surfaces have antimicrobial properties against a variety of different microorganisms and copper touch surfaces are likely to be increasingly used in public places including hospitals. Studies in the literature show that molecular mechanisms result in the rapid killing of Cu surface-exposed bacteria and yeasts result from a sharp shock of extreme and immediate Cu-ion overload combined with severe membrane and cell envelope damage, although similar low mutation rates have been observed in cells obtained from both Cu and control surfaces. The aim of this study was to determine the survival of bacteria on the surfaces of copper and plastic plumbing surfaces. The antibacterial activity of copper surfaces was determined by overlying suspensions of Staphylococcus aureus and Escherichia coli on copper and plastic surfaces. All pipes were sterilized and bacterial suspensions from colonies were prepared and then the pipes were contaminated by the bacterial suspension. The experiments were performed at 18-23°C and the results were assessed after a 20-day exposure. The numbers of viable bacteria in the suspension were determined by serial dilution and plating on Nutrient Agar plates; the plates being incubated at 37°C for 48 h. The results showed that low counts of Staphylococcus aureus were seen on copper surfaces, as compared with those obtained on the plastic, control surfaces, i.e., the results show that E. coli failed to survive on copper pipes. The number of bacteria isolated from the plastic surfaces was consistently higher than the number isolated from copper surfaces. The survival rate of bacteria on the copper surfaces was low and none of the inoculated bacteria survived after 20 days of exposure. Copper is well known to be an antibacterial, and its use in medical environments is likely to the lead to the continuous reduction of environmental microbial contamination, including MRSA. The studies presented here show that the incorporation of Cu in healthcare facilities may dramatically help reduce the environmental microbial burden and act as a useful adjunct to current infection prevention and control systems, despite the fact that bacteria will eventually acquire resistance to the ion.

Rana Abdulrahim Alaeq *et al.*, Clinical Microbiology 2017, 6:5 (Suppl) DOI: 10.4172/2327-5073-C1-030

8th CLINICAL MICROBIOLOGY CONFERENCE October 26-28, 2017 | Paris, France



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APPINDIX 2

The phylogenetic analysis of bacteria isolated from environment samples

1- The phylogenetic analysis of bacteria isolated from sink samples

Klebsiella oxytoca CP011618.1

Klebsiella oxytoca strain CAV1335, complete genome Sequence ID: <u>gb[CP011618.1]</u> Length: 6229565 Number of Matches: 8

Range 1: 2928343 to 2929103 GenBank Graphics			V Nex	t Match A Previous N	latch		
Score 1373 bits	s(1522)	Expect 0.0	Identities 761/761(100%)	Gaps 0/761(0%)	Strand Plus/Plus		
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Query	121	TCCAATCO	GGACTACGACATACT	TATGAGGTCCGC	TTGCTCTCGCGAG	GTCGCTTCTCT	180
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Sbjct	2928583	TCATCCCC	CACCTTCCTCCAGTTT	ATCACTGGCAGTC	CCTTTGAGTTCC	CGACCTAATCG	2928642
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Sbjct	2928643	CTGGCAA	CAAAGGATAAGGGTTGO	CGCTCGTTGCGGG	ACTTAACCCAACA	TTTCACAACAC	2928702
Query	361	GAGCTGA	CGACAGCCATGCAGCAC	CCTGTCTCAGAGT	CCCGAAGGCACC	AAAGCATCTCT	420
Sbjct	2928703	GAGCTGA	ACAGCCATGCAGCACCTGTCTCAGAGTTCCCGAAGGCA	CCCGAAGGCACC	CAAAGCATCTCT	2928762	
Query	421	GCTAAGT	CTCTGGATGTCAAGAG	GTAGGTAAGGTTC	TCGCGTTGCATC	GAATTAAACCA	480
Sbjct	2928763	GCTAAGT	CTCTGGATGTCAAGA	GTAGGTAAGGTTC	TCGCGTTGCATC	GAATTAAACCA	2928822
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Sbjct	2928823	CATGCTCO	CACCGCTTGTGCGGGGC	CCCCGTCAATTCA	TTTGAGTTTTTAAC	CTTGCGGCCGT	2928882
Query	541	ACTCCCCZ	AGGCGGTCGACTTAAC	GCGTTAGCTCCGGA	\AGCCACTCCTCA	AGGGAACAACC	600
Sbjct	2928883	ACTCCCC	AGGCGGTCGACTTAACO	GCGTTAGCTCCGGA	AGCCACTCCTCA	AGGGAACAACC	2928942
Query	601	TCCAAGT	CGACATCGTTTACAGCO	GTGGACTACCAGG(GTATCTAATCCTG	ITTGCTCCCCA	660
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Sbjct	2929063	CCAGATC	CTACGCATTTCACCG	CTACACCTGGAAT	CTAC 292910	3	

Klebsiella oxytoca strain MTJW-7 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. SCT-5 16S ribosomal RNA gene, partial sequence	
Bacterium CSR-MOB8 16S ribosomal RNA gene, partial sequence	
enterobacteria 3 leaves	
enterobacteria 2 leaves	
2 <u>2 [c]Query_38105</u>	
enterobacteria 10 leaves	
Klebsiella oxytoca strain FUA 1266 16S ribosomal RNA gene, partial sequence	
Victoria et al segunda et a	
Klebsiella sp. 514b 16S ribosomal RNA gene, partial sequence	
Uncultured Klebsiella sp. clone F4jun.44 16S ribosomal RNA gene, partial sequence	
Uncultured Klebsiella sp. clone 30D16E 93 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. 2392 16S ribosomal RNA gene, partial sequence	
Klebsiella michiganensis strain AG-52 16S ribosomal RNA gene, partial sequence	
Uncultured Klebsiella sp. clone XY2-43 16S ribosomal RNA gene, partial sequence	
Klebsiella oxytoca gene for 16S ribosomal RNA, partial sequence, strain: NGB-FR89	
enterobacteria 8 leaves	
Klebsiella oxytoca strain CAV1374, complete genome	
Uncultured Klebsiella sp. gene for 16S ribosomal RNA, partial sequence, isolate: HLNR03	
Klebsiella oxytoca HKOPL1, complete genome	
Klebsiella oxytoca strain MFM-01 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. Q11 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. ICB439 16S ribosomal RNA gene, partial sequence	
Uncultured Klebsiella sp. clone Fljun.37 16S ribosomal RNA gene, partial sequence	
Bacterium SV6XIII 16S ribosomal RNA gene, partial sequence	
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Klebsiella oxytoca gene for 16S ribosomal RNA, partial sequence, strain: DRM9	
Klebsiella oxvtoca strain YN201309 16S ribosomal RNA gene, partial sequence	
Klebsiella oxytoca strain NF903966 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. ICB414 16S ribosomal RNA gene, partial sequence	
Uncultured Klebsiella sp. clone F3jan.13 16S ribosomal RNA gene, partial sequence	
Uncultured bacterium clone (2h24 16S ribosomal RNA gene, partial sequence	
Enterobacter sp. SW75 16S ribosomal RNA gene, partial sequence	
Klebsiella oxytoca gene for 16S ribosomal RNA, partial sequence, strain: NGB-FR129	
Klebsiella michiganensis strain BAL29 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. SPC06 16S ribosomal RNA gene, partial sequence	
Klebsiella oxytoca strain BCB4 16S ribosomal RNA gene, partial sequence	
Klebsiella oxytoca KCTC 1686, complete genome	
Klebsiella oxytoca strain NFL28 16S ribosomal RNA gene, partial sequence	
Uncultured Klebsiella sp. clone F4aug.32 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. PAMU-1.2 gene for 16S rRNA, partial sequence	
Uncultured bacterium clone XXM_1_062 16S ribosomal RNA gene, partial sequence	
enterobacteria 18 leaves	
🐗 enterobacteria 12 leaves	
Vielessiella oxytoca gene for 16S ribosomal RNA, partial sequence, strain: NGB-FR100	
Klebsiella oxytoca gene for 16S ribosomal RNA, partial sequence, strain: NGB-FR67	
4 Klebsiella michiganensis strain AB-181 16S ribosomal RNA gene, partial sequence	
Klebsiella sp. SM1 eene for 16S ribosomal RNA narial sequence	

0.001 .

Klebsiella sp. SM1 gene for 16S ribosomal RNA, partial sequence
 Klebsiella oxytoca strain E_IC 16S ribosomal RNA gene, partial sequence
 Uncultured Klebsiella sp. clone 8d165f 16S ribosomal RNA gene, partial sequence

Bacillus subtilis KP340123.1

Bacillus subtilis strain W6 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|KP340123.1]</u> Length: 1431 Number of Matches: 1

Range 1: 467 to 1367 GenBank Graphics Vext Match 🛦 Previous Match						
Score 1626 bits	(1802)	Expect 0.0	Identities 901/901(100%)	Gaps 0/901(0%)	Strand Plus/Minus	
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Query	121	TTGTGGGATTG	GCTTAACCTCGCGGTT	TCGCTGCCCTTT	GTTCTGTCCATTGTAGCACGT	180
Sbjct	1247	TTGTGGGATTG	GCTTAACCTCGCGGTT	I I I I I I I I I I I I I I I I I I I	GTTCTGTCCATTGTAGCACGT	1188
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Sbjct	947	CTGGTAAGGTT	CTTCGCGTTGCTTCGA	ATTAAACCACAT	GCTCCACCGCTTGTGCGGGCC	888
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Query	901	A 901				
Sbjct	467	A 467				



Kocuria rhizophila KM978822.1

Kocuria rhizophila strain AHT-1 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KM978822.1</u>] Length: 1491 Number of Matches: 1

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Sbjct	974	GGTCCGGTGTA	TGTCAAGCCTTGGT	AAGGTTCTTCGCO	TTGCATCGAATTAATCCGCA	TG 915
Query	481	CTCCGCCGCTT	GTGCGGGGCCCCGT	CAATTCCTTTGAG	TTTTAGCCTTGCGGCCGTAC	TC 540
Sbjct	914	CTCCGCCGCTT	GTGCGGGCCCCCGT	CAATTCCTTTGAG	TTTTAGCCTTGCGGCCGTAC	TC 855
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Sbjct	794	CTAGTGCCCAA	CGTTTACGGCATGG	ACTACCAGGGTAI	CTAATCCTGTTCGCTCCCCA	TG 735
Query	661	CTTTCGCTCCT	CAGCGTCAGTAACA	GCCCAGAGACCTO	CCTTCGCCATCGGTGTTCCT	CC 720
Sbjct	734	CTTTCGCTCCT	CAGCGTCAGTAACA	GCCCAGAGACCTO	CCTTCGCCATCGGTGTTCCT	CC 675
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Sbjct	554	ACCGCCTACGA	GCTCTTTACGCCCA	 ATAATT-CCGGAC	AACGCTTGCGCCCTACGTAT	11 TA 496
Query	901	CCGCGGCTGCT	GGCACGTAGT 92	1		
Sbjct	495	CCGCGGCTGCT	GGCACGTAGT 47	5		

Bacteria	m PL41 16S ribosomal RNA gene martial sequence
Bacteria	m PL-37 165 ribosomal RNA gene partial sequence
high G	C Gram+14 leaves
Kocuria	rhizophila strain BCX-32 16S ribosomal RNA gene, partial sequence
Kocuria	rhizophila strain BCX-8 16S ribosomal RNA gene, partial sequence
Lysiniba	acillus sphaericus strain ACCC11096 16S ribosomal RNA gene, partial sequence
🐴 high GO	C Gram+ 4 leaves
₽ k	Cocuria rhizophila 16S ribosomal RNA gene, partial sequence
	Kocuria rhizophila partial I6S rRNA gene, isolate G5
	Kocuria rhizophila partial 16S rRNA gene, isolate G4
	Kocuria rhizophila partial 16S rRNA gene, isolate G2
	Bacterium PJ-25 16S ribosomal RNA gene, partial sequence
	Kocuria rhizophila strain 236-4A 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium partial 16S rRNA gene, clone MC03D12
	unclassified 2 leaves
	 Uncultured bacterium clone nbt27c07 16S ribosomal RNA gene, partial sequence
	high GC Gram+ 2 leaves
	A bacteria 10 leaves
	Kocuria rhizophila partial 16S rRNA gene, strain PE-LR-2
	Very Kocuria rhizophila strain TA68 16S ribosomal RNA gene, partial sequence
	g Uncultured bacterium clone 16slp101-3c05.p1k 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 16slp123-2b01.p1k 16S ribosomal RNA gene, partial sequence
	high GC Gram+ 2 leaves
	Kocurta sp. M1-56 16S ribosomal RNA gene, partial sequence Mixing COC Security of the security of
	Thigh GC Gram+ 4 leaves
	Kocuria sp. FXJ8.237 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 16stp101-1b03.w2k 16S ribosomal RNA gene, partial sequence
	high GC Gram+ 2 leaves
	Kocuria sp. SGb100 16S ribosomal RNA gene, partial sequence
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	Y Kocura sp. jca+ 105 mossinar KNA gene, partial sequence
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	Kocuria sp PNS 16S ribosomal RNA gene partial sequence
	Uncultured hacterium clone [6sin10] -3(04 n]k [6S ribosomal RNA sene, nartial sequence
	Kocuria rhizophila gene for 16S ribosomal RNA, partial sequence
	Uncultured bacterium clone 16slp101-2d10.w2k 16S ribosomal RNA gene, partial sequence
	Kocuria sp. 9851a 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 16slp123-1b06.w2k 16S ribosomal RNA gene, partial sequence
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	Recura mizopina stran XPD-XX tos noosomai RNX gene, partai sequence
	g che unureu bacierinani e tosipi101-2004.weki tos nibosomia Riva gene, partial sequence
	Incultured organism clone FL100177.7472-SNDPCRAMeAN 000607 small subunit ribosomal RNA gene nartial sequence
	Uncultured bacterium clone 16slp101-2g11.w2k 16S ribosomal RNA gene, partial sequence
	Kocuria sp. SA3 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium partial 16S rRNA gene, clone SA04C12
	Uncultured organism clone ELU0177-T472-S-NIPCRAMgANa_000082 small subunit ribosomal RNA gene, partial sequence
	Kocuria rhizophila strain ID4123 I6S ribosomal RNA gene, partial sequence
	 Kocuria sp. pca2 16S ribosomal RNA gene, partial sequence
	Kocurra fuzophila strain AH1-1 165 ribosomal KNA gene, partial sequence
	Victura sp. rivo tos nuosomai KivA gene, partial sequence
	King a discontanti con e ou crea - rob rob novona robe gene, partial sequence Konita discontante - 100 robe - 10
	Kestina minospina stain 7 L22 too movinin RCV gene, partial sequence Kestina minospina stain 7 L22 too movinin RCV gene, partial sequence
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	Psychrobacter sp. MAR705 partial 16S rRNA gene, isolate MAR705
	Kocuria sp. FXJ8.177 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 16slp101-3b01.p1k 16S ribosomal RNA gene, partial sequence
.0.001	Kocuria sp. MT4.1 16S ribosomal RNA gene, partial sequence
0.004	 Uncultured bacterium partial 16S rRNA gene, clone SA03E03
	Scouria rhizophila strain XFB-BG I6S ribosomal RNA gene, partial sequence

Uncultured bacterium partial 16S rRNA Kocuria rhizophila strain XFB-BG 16S ribosomal RNA gene, partial sequence unclassified |2 leaves

Bacillus cereus KC731425.1

Bacillus cereus strain VITSH1 16S ribosomal RNA gene, partial sequence sequence ID: $\underline{gb|KC731425.1|}$ Length: 1369 Number of Matches: 1

Range 1: 513 to 1353 GenBank Graphics Vext Match 🛦 Previous Match							
Score		Expect	Identities	Gaps	Strand		
1283 bits	s(1422)	0.0	789/841(94%)	0/841(0%)	Plus/Minus		
Query	1	ACGGGCGGTGT			CGGCATGCTGATCCGCG	ATTAC 60	
Sbjct	1353	ACGGGCGGGGT	GTACAAGGCCCGGGA	ACGTATTCACCG	CGGCATGCTGATCCGCG	ATTAC 12	94
Query	61	TAGCGATTCCA	GCTTCATGTATGCAA	AGTTGCAACCGAC	AATCCAAACTGAAAACG	GTTTT 12	0
Sbjct	1293	TAGCGATTCCA	GCTTCAGGTAGGCAZ	GTTGCAGCCTAC	AATCCAAACTGAAAACG	GTTTT 12	34
Query	121	ATGATATTAGC	TCCACCTCGCGGTC	TGCAGCTCTTTG	FACCGTCCATTGTAACA	.CGTGT 180	0
Sbjct	1233	ATGAAATTAGC	TCCACCTCGCGGTCI	TGCAGCTCTTTG	TACCGTCCATTGTAGCA	CGTGT 11	74
Query	181	GTACCCCAGGT	CATAGGGGGGCATGAI	GATTTGACGTCA	ICCCCACCTTCCTCCGG	TTTGT 24	0
Sbjct	1173	GTAGCCCAGGT	CATAAGGGGCATGAI	IGATTTGACGTCA:	FCCCCACCTTCCTCCGG	TTTGT 11	14
Query	241	CACCGGCAGTC		ACTTATGGATGG	CAAGTAAGATCAAGGGT	TGCGC 30	0
Sbjct	1113	CACCGGCAGTC	ACCTTAAAGTGCCCA	ACTAAATGATGG	CAACTAAAATCAAGGGT	TGCGC 10	54
Query	301	TCGTTGCGGGA	CTTAACCCAACATC	CACAACACAACC	IGACAACAACCATGCAC	CACCT 36	0
Sbjct	1053	TCGTTGCGGGA	CTTAACCCAACATCI	CACAACACAAGC	IGACAACAACCATGCAC	CACCT 99	4
Query	361	GTCACTCTGCT	CCCGAAGGAAAAGAC	CTATCTCTAGGG	TTTTCACAGGATGTCAA	GAGCT 42	0
Sbjct	993	GTCACTCTGCT	CCCGAAGGAAAAGCC	CTATCTCTAGGG	ITGTCAAAGGATGTCAA	GACCT 93	4
Query	421	GGTAAGGGTCT		ATTAAACCACATG			0
Sbjct	933	GGTAAGGTTCT	TCGCGTTGCTTCAAA	ATTAAACCACATG	CTCCACCGCTTGTGCGG	GCCCC 87	4
Query	481	CGTCAATTCCT	TTGAATTTCAACCTI	GCGGACGTACTC	CCCAGGGGGGAATGGTTA	ATGCG 54	0
Sbjct	873	CGTCAATTCCT	TTGAGTTTCAGCCTI	IGCGGCCGTACTC	CCCAGGCGGAGTGCTTA	ATGCG 81	4
Query	541	TTAACTTCCGC	ACTAAAGGACGGAA	ACCCTCTAACACT		CGGCGG 60	00
Sbjct	813	TTAACTTCAGC	ACTAAAGGGCGGAA	ACCCTCTAACACT	TAGCACTCATCGTTTA	CGGCGT 75	54
Query	601	GAACTACCAGG	GTATCTAATCCGGT	TTGCTCCCCACGC	TTTCGCGCCTCACTGT	CAGTTA 66	50
Sbjct	753	GAACTACCAGG	GTATCTAATCCGGT	ITGCTCCCCACGC	TTTCGCGCCTCATTGT	CAGTTA 69	94
Query	661	GAGACCAGAAA	GTCCCCTTCGCCAC	IGGGGTTCCTCCA	TATCTCTACACATTTCA	ACCGCT 72	20
Sbjct	693	CAGACCAAAAA	GTCGCCTTCGCCAC	IGGTGTTCCTCCA	TATCTCTACGCATTTC	ACCGCT 63	34
Query	721	ACCCATGAAAT		CTGCACTCAAATC		CCCTCC 78	30
Sbjct	033	ACACATGGAAT	TCCACTTTCCTCTTC	CTGCACTCAAGTC	TCCCAGTTTCCAATGA	Decree 57	/4
Query	/81	GGGGTTGACCC				PACCCC 84	ŧU
Sbjct	573	ACGGTTGAGCC	GTGGGCTTTCACATO	CAGACTTAAGAAA	CCACCTGCGCGCGCTT	FACGCC 51	14
Query	841	C 841					
Sbjct	513	c 513					

e IellOuery 213179	
Bacillus cere	us strain 7852 16S ribosomal RNA gene partial sequence
firmicutes 12 le	aves
↓ firmicutes 4	leaves
- Bacillus cer	eus strain ANY 16S ribosomal RNA, partial sequence
- Bacil	lus cereus strain BA6-12 16S ribosomal RNA gene, partial sequence
Bacillus cer	eus strain CB-10 16S ribosomal RNA gene, partial seguence
- firmicute	s 2 leaves
	cillus sp. DmBR 32 16S ribosomal RNA gene, partial sequence
9 Bacil	laceae bacterium 6 St 3 16S ribosomal RNA gene, partial sequence
	firmicutes 4 leaves
4 9 B	acillus cereus strain FORC 005, complete genome
↓ B	acillus subtilis strain E1-3 16S ribosomal RNA gene, partial sequence
A B	acillus cereus strain CLHDHF(2)B 1-1 16S ribosomal RNA gene, partial sequence
	acillaceae bacterium b6 i17 16S ribosomal RNA gene, partial sequence
• B	acillaceae bacterium b5 i15 16S ribosomal RNA gene, partial sequence
• B	acillaceae bacterium b1 i6 16S ribosomal RNA gene, partial sequence
φB	acillaceae bacterium b1 i5 16S ribosomal RNA gene, partial sequence
B	acillus sp. HLB-3 16S ribosomal RNA gene, partial sequence
0	 Unidentified bacterium clone MEB140 16S ribosomal RNA gene, partial sequence
🐴 fi	irmicutes 3 leaves
	firmicutes 5 leaves
h i i i i i i i i i i i i i i i i i i i	Bacillus thuringiensis strain GYL1 16S ribosomal RNA gene, partial sequence
La La	firmicutes 4 leaves
4	Bacillus toyonensis strain D26 16S ribosomal RNA gene, partial sequence
	Bacillus sp. INBio_4108D 16S ribosomal RNA gene, partial sequence
	Bacillus sp. FW11 16S ribosomal RNA gene, partial sequence
	firmicutes 6 leaves
	Uncultured bacterium clone 6_Am_42 16S ribosomal RNA gene, partial sequence
	Bacillus sp. RM1(2014) 16S ribosomal RNA gene, partial sequence
	Bacillus anthracis strain IHB B 15784 16S ribosomal RNA gene, partial sequence
	Uncultured Bacillus sp. clone C4A04 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain NM2E11 16S ribosomal RNA gene, partial sequence
	Bacillus thuringiensis strain KNUSC3015 16S ribosomal RNA gene, partial sequence
	firmicutes 7 leaves
	Bacillus sp. S13(2015) 16S ribosomal RNA gene, partial sequence
	firmicutes 6 leaves
	Uncultured Bacillus sp. clone YXY-9 16S ribosomal RNA gene, partial sequence
	Bacillus thuringiensis strain YC-10, complete genome
	Uncultured Bacillus sp. clone RHDTWG185 16S ribosomal RNA gene, partial sequence
	Choultured Bacilius sp. clone C5D04 165 ribosomal KNA gene, partial sequence
	Bacillus antifracis strain IHB B 18203 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone b5_51 16S ribosomal KNA gene, partial sequence
	Pacifius cereus surain BGZ-8 105 ribosomai KINA gene, partial sequence
	Bacillus en CH-L1116S ribosomal RNA gene, partial sequence
	Dacinus sp. CH-E11 105 fibosofiat KIVA gene, partial sequence
	Poncunarea dacinas sp. cione KHD1 w0181 105 mossomai KNA gene, partial sequence
	Bacillus anthracis strain IHB B 19120 105 ribosomal RNA gene, partial sequence
0.007	Bacillus en IS-20 I6S ribosomal RNA cene partial sequence
	Uncultured hastarium clone 6 K 45 16S ribosomal DNA cana institut sequence
	Uncutured bacterium clone 6_K_45 165 ribosomai KNA gene, partial sequence

Micrococcus luteus KT339390

Micrococcus luteus strain DYPSBB RPF YRP01 16S ribosomal RNA gene, partial sequence Sequence ID: <u>KT339390.1</u> Length: 1462 Number of Matches: 1

Range 1: 30 to 1424 GenBank Graphics Vext Next Next Next Next Next Next Next N					latch 🔺 Previous Match
Score 2560	bits(13	Expect 86) 0.0	Identities 1393/1396(99%)	Gaps 2/1396(0%)	Strand Plus/Plus
Query	2	ACATGCAAGTCGAACGA	TGAAGCCCAGCTTGCTGGGTGG	SATTAGTGGCGAACGGGTGAGT	61
Sbjct	30	ACATGCAAGTCGAACGA	téAAécccAécttéCtééétéé	SATTAGTGGCGAACGGGTGAGT	89
Query	62	AACACGTGAGTAACCTG			121
Ouerv	122	GATAGGAGCGTCCACCG	CATGGTGGGTGTTGGAAAGATT	TATCGGTTTTGGATGGACTCG	145
Sbjct	150	GATAGGAGCGTCCACCG	CATGGTGGGTGTTGGAAAGATT	TATCGGTTTTGGATGGACTCG	209
Query	182	CGGCCTATCAGCTTGTT	GGTGAGGTAATGGCTCACCAAG	3GCGACGACGGGTAGCCGGCCT	241
Sbjct	210	ceectatcaecttett	ddtdaddtaatddctcaccaad	beceletere a construction of the construction	269
Query	242	GAGAGGGTGACCGGCCA			301
Ouerv	302	GTGGGGGAATATTGCACA			329
Sbjct	330	GTGGGGGAATATTGCACA	ATGGGCGCAAGCCTGATGCAG		389
Query	362	ĢÇÇTTÇĞĞĞTTĞTAAAÇ	стстттсадтаддаадаасс	ҙ҄ѧѧѧӯҭӯѧҫӯӯҭѧҫҫҭӯҫѧӯѧѧ	421
Sbjct	390	GCCTTCGGGTTGTAAAC	CTCTTTCAGTAGGGAAGAAGC	SAAAGTGACGGTACCTGCAGAA	449
Query	422	GAAGCACCGGCTAACTA	CGTGCCAGCAGCCGCGGTAATA	ACGTAGGGTGCGAGCGTTATCC	481
Sbjct	450	GAAGCACCGGCTAACTA	CGTGCCAGCAGCCGCGGTAATA		509
Query	482		AAGAGCTCGTAGGCGGTTTGTC		541
Ouerv	542	GGCTTAACCCCGGATCT	GCGGTGGGTACGGGCAGACTAG	TAGTGCAGTAGGGGGGGGGGGCTGGA	601
Sbjct	569	GGCTTAACCCCGGATCT	GCGGTGGGTACGGGCAGACTAG	JAGTGCAGTAGGGGAGACTGGA	628
Query	602	ATTCCTGGTGTAGCGGT	GGAATGCGCAGATATCAGGAGG	SAACACCGATGGCGAAGGCAGG	661
Sbjct	629	ATTCCTGGTGTAGCGGT	GGAATGCGCAGATATCAGGAGG	GAACACCGATGGCGAAGGCAGG	688
Query	662	TCTCTGGGCTGTAACTG	ACGCTGAGGAGCGAAAGCATGO	3GGAGCGAACAGGATTAGATAC	721
Sbjct	689	TCTCTGGGCTGTAACTG	ACGCTGAGGAGCGAAAGCATGO	GGAGCGAACAGGATTAGATAC	748
Sbict	749				781 808
Query	782	CGCCGCAGCTAACGCAT	TAAGTGCCCCGCCTGGGGAGTA		841
Sbjct	809	CGCCGCAGCTAACGCAT	TAAGTGCCCCGCCTGGGGAGTA	ACGGCCGCAAGGCTAAAACTCA	868
Query	842	AAGGAATTGACGGGGGC	CCGCACAAGCGGCGGAGCATGC	GGATTAATTCGATGCAACGCG	901
Sbjct	869	AAGGAATTGACGGGGGGC	ccdcacaadcddcddddddddd	todattaattcoatocaacoco	928
Query	902				961
Ouerv	929	GGGCGGGTTCACAGGTG	TIGACATGITICICGATCGCCG	TAGAGATACGGTTTCCCCTTTG	988
Sbict	989	GGGCGGGTTCACAGGTG	GTGCATGGTTGTCGTCAGCTCG	TGTCGTGAGATGTTGGGTTAA	1048
Query	1022	GTCCCGCAACGAGCGCAA		ACGTAATGGTGGGGACTCATG	1081
Sbjct	1049	GTCCCGCAACGAGCGCAA	ACCCTCGTTCCATGTTGCCAGC/	ACGTAATGGTGGGGACTCATG	1108
Query	1082	GGAGACTGCCGGGGTCAA	ACT CGGAGGAAGGTGAGGACGA(CGTCAAATCATCATGCCCCTT	1141
Sbjct	1109	GGAGACTGCCGGGGTCA	ACTCGGAGGAAGGTGAGGACGA	cgtcaaatcatcatgcccctt	1168
Query	1142	ATGTCTTGGGCTTCACG	CATGCTACAATGGCCGGTACAA	TGGGTTGCGATACTGTGAGGT	1201
Sbjct	1169	Atététtéééééttékéé	catóctacaatóóccóótacaa	tögöttöcöAtActötöAööt	1228
Query	1202	GGAGCTAATCCCAAAAAG	GCCGGTCTCAGTTCGGATTGGG	GTCTGCAACTCGACCCCATGA	1261
Sbjct	1229	GGAGCTAATCCCAAAAAA	SCCGGTCTCAGTTCGGATTGGG	GTCTGCAACTCGACCCCATGA	1288
Query	1262				1321
Ouerv	1322	GTACACACCGCCGTCA			1381
Sbict	1349				1408
Query	1382	TGTGGGGG-AGCCGTC	1396		
Sbjct	1409	TGTGGGGGGGAGCCGTC	1424		


2- The phylogenetic analysis of bacteria isolated from mirror samples

Bacillus cereus KP100400.1

Bacillus cereus 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KP100400.1|</u> Length: 1179 Number of Matches: 1

Range 1:	Range 1: 55 to 1014 GenBank Graphics Vext Match 🔺 Previous Match					
Score 1683 bit	s(1866)	Expect 0.0	Identities 949/960(99%)	Gaps 0/960(0%)	Strand Plus/Plus	
Query	1	GGCGGACGGGTG	GAGTAACACGTGGGT	AACCTGCCCATA	AGACTGGGATAACTCCGGGAAA	60
Sbjct	55	GGCGGACGGGT	GAGTAACACGTGGGT	AACCTGCCCATA	AGACTGGGATAACTCCGGGAAA	114
Query	61	CCGGGGGCTAATA	ACCGGATAACATTTT	GAACCGCATGGT	CGAAATTGAAAGGCGGCTTCG	120
Sbjct	115	CCGGGGGCTAATA	ACCGGATAACATTTT	GAACCGCATGGT	CGAAATTGAAAGGCGGCTTCG	174
Query	121	GCTGTCACTTAT	GGATGGACCCGCGT	CGCATTAGCTAG	TGGTGAGGTAACGGCTCACCA	180
Sbjct	175	GCTGTCACTTAT	GGATGGACCCGCGT	CGCATTAGCTAG	TGGTGAGGTAACGGCTCACCA	234
Query	181	AGGCAACGATGC	GTAGCCGACCTGAG	AGGGTGATCGGC	CACACTGGGACTGAGACACGGC	240
Sbjct	235	AGGCAACGATGC	CGTAGCCGACCTGAG	AGGGTGATCGGC	CACACTGGGACTGAGACACGGC	294
Query	241	CCAGACTCCTAC	GGGAGGCAGCAGTA	GGAATCTTCCG	CAATGGACGAAAGTCTGACGGA	300
Sbjct	295	CCAAACTCCTAC	GGGAGGCAGCAGTA	GGAATCTTCCGC	CAATGGACGAAAGTCTGACGGA	354
Query	301	GCAACGCCGCGI	GAGTGATGAAGGCT		ACTCTGTTGTTAGGGAAGAAC	360
Sbjct	355	GCAACGCCGCGI	GAGTGATGAAGGCT	TTCGGGTCGTAA	ACTCTGTTGTTAGGGAAGAAC	414
Query	361	AAGTGCTAGTTG	SAATAAGCTGGCACC		ACCAGAAAGCCACGGCTAACT	420
Sbjct	415	AAGTGCTAGTTG	SAATAAGCTGGCACC	TTGACGGTACCT	ACCAGAAAGCCACGGCTAACT	474
Query	421	ACGTGCCAGCAG	CCGCGGTAATACGT	AGGTGGCAAGCG	TATCCGGAATTATTGGGCGTA	480
Sbjct	475	ACGTGCCAGCAG	SCCGCGGTAAAACGT	AGGTGGCAAGCG	TATCCGGAATTATTGGGCGTA	534
Query	481		GTGGTTTCTTAAGT	CTGATGTGAAAGO	CCACGGCTCANCCGTGGAGGG	540
Sbjct	535	AAGCGCGCGCAG	GTGGTTTCTTAAGT	CTGATGTGAAAGO	CCCACGGCTCAACCGTGGAGGG	594
Query	541	TCATTGGAAACT	GGGAGACTTGAGTG		TGGAATTCCATGTGTAGCGGT	600
Sbjct	595	TCATTGGAAACT	GGGAGACTTGAGTG	CAGAAGAGGAAAG	TGGAATTCCATGTGTAGCGGT	654
Query	601	GAAATGCGTAGA	GATATGGAGGAACAC	CAGTGGCGAAGG	CGACTTTCTGGTCTGTAACTG	660
Sbjct	655	GAAATGCGTAGA	GATATGGAGGAACAC	CAGTGGCGAAGG	CGACTTTCTGGTCTGTAACTG	714
Query	661	ACACTGAGGCGC	GAAAGCGTGGGGAG	CAAACAGGATTAG	ATACCCTGGTAGTCCACGCCG	720
Sbjct	715	ACACTGAGGCGC	GAAAGCGTGGGGAGC	CAAACAGGATTAG	ATACCCTGGTAGTCCACGCCG	774
Query	721	TAAACGATGAGT		GTTTCCGCCCTT	TAGTGCTGAAGTTAACGCATT	780
Sbjct	775	TAAACGATGAGT	GCTAAGTGTTAGAGO	GTTTCCGCCCTT	TAGTGCTGAAGTTAACGCATT	834
Query	781	AAGCACTCCGCC	TGGGGAGTACGGCCG	SCAAGGCTGAAAC	TCAAAGGAATTGACGGGGGGCC	840
Sbjct	835	AAGCACTCCGCC	TGGGGAGTACGGCCC	SCAAGGCTGAAAC	TCAAAGGAATTGACGGGGGGCC	894
Query	841	CGCACAAGCGGG	GGAGCATGTGGTTT	ATTCGAAGCAAC	GCAAAAAACCTTACCAGGTCT	900
Sbjct	895	CGCACAAGCGGT	GGAGCATGTGGTTT	ATTCGAAGCAAC	GCGAAAAACCTTACCAGGTCT	954
Query	901	TGACATCCTCCG		AGGGCTTCCCCTT	CCGGAACAAAATGACAGGGGG	960
Sbjct	955	TGACATCCTCTG	AAAACCCTAAAAAT	AGGGCTTCCCCTT	CGGGAACAAAATGACAGGTGG	1014

C5.	*kt/gaery_28035	# Bacillus cereas 165 ribosonial RNA gene, partial seasence
		# Bacillus sp. CH-KOV4 16S ribosonal RNA gene, partial sequence
	L	 Bacillus cereus strain MZ-01 16S ribosonal RNA gene, partial sequence
		• firmicutes 2 leaves
		 Bacillus tharingiensis strain DMB 165 robosonal RNA gene, partial sequence
		all femi-use [5] leaves
		 Baselins on XOWS 16S relevantial RNA cone, narrial sources
		Bacillus anthracis strain LAMA 1145 165 ribosonal RNA erne, natial sequence
		# Bacillus cereus strain B4 165 ribosonal RNA gene, partial sequence
		Uncultured bacterium clone 11_Am_27 16S ribosomal RNA gene, partial sequence
		 Uncultured bacterium clone 11 Am 52 165 ribosomal RNA gene, partial sequence
		 Uncultured bacterium clone 11 K 47 165 ribosomal RNA gene, partial sequence
		 Uncellared backenani cone 11 SI 27 Hos neosonal KNA gene, partai sequence Bestilar emissional district de characteri BNA emissione entral estatemente Anno 11 SI 2010 br/>SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI 2010 SI /li>
		 Bacillas en 1900. As 165 bioacend RNA pere particular experies
		 Bacillus coreus strain SN6 105 ribosomal RNA gene, partial sequence
		Bacillus thuringiensis strain PB 165 ribosomal RNA gene, partial sequence
		Bacillus cereus strain ZJB-12033 165 ribosomal RNA gene, partial sequence
		Pacillus anthracis strain A0157 sequence
		Bacilias cereus strain TS01 165 ribosomal RNA gene, partial sequence
		 Baculas coreas strain L2/2 fos mosenia RNA, gene, partial sequence
		 Boccillate antifactor strain Artics Arbona, compare genome Boccillate antifactoria strain AC acounterios
		 Recilias anthracis strain SK-102, complete pensone
		# Bacillas anthracis str. Sterne, complete genome
		Bacillus cereus G9241, complete genome
		#Bacillus cereus 03BB108, complete genome
		Bacillus cereus strain ZJU2 165 ribosomal RNA gene, partial sequence
		9 Bacillus coreus strain KES7 165 ribosomal RNA gene, partial sogaence
		 Crectulared backgrain (cone vib ios motornal RNA gene, partai sequence Backgrain (cone vib ios) motornal RNA gene, partai sequence
		 Bachars unimprents strain Ar 105 intervienta nuver gene, partai sequence Bachars nu VIII 165 cilocanal RNA sone martial security of Anno 1990 Intervienta R
		a Bacillus coreus strain NE5 105 ribosonal RNA cone, partial sourcece
		Bacilus sp. DB14780 16S ribosonal RNA gene, partial sequence
		⁴ Uncultured Bacillus sp. gene for 16S ribosomal RNA, partial sequence, isolate: LZX-37
		g Bacillus anthracis 16S ribosomal RNA gene, partial sequence
		 Uncultured Bacillus sp. clone RL-11 165 ribosomal RNA gene, partial sequence
		 Bacillas coreus A ECC 4342, compte ensorme Bacillas coreus A ECC 4342, compte ensorme
		 Exciting anomaly size view of the complete generation Reserves and the size of the Reserves of the size of
		 Bacinas antirações strain FAR, complete persone Bacinas atraine FAR, complete persone
		Bacillas corrus 03BB102, complete persone
		Bacillus anthracis 16S rRNA gene, isolate EM1
		^a Bacilus cereus strain SKSF55 16S ribosomal RNA gene, partial sequence
		Bacillus anthracis strain IHB B 7081 16S ribosomal RNA gene, partial sequence.
		 Bacillas sp. BAB-4397 165 ribosonal RNA gene, partial sequence
		 Bacillas coreus strain AUCEEATS TOS infosionnal KNA gene, partai sequence a Bacillas coreus partiel 165, e0 Na asse lacidat BBAIL
		 Declared exception and construction processing of the second processing and the second processing of the sec
		the same backing the case of the same and the same and the same the
		 Bachters p. 1996; etc. 1997 105 Februaria RCAS generative points requests etc. Bachters are chain HSTI 165 Februarial BNA area tortial secondore
		Uncultured bacterium close 11 St 14 16S reported BNA rene, natial sequence
		Uncultured bacterium clone 11 K 33 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone 11 Am 44 16S ribosomal RNA gene, nartial sequence
		Uncultured bacterium clone 11 Am 26 165 ribosomal RNA gene, partial sequence
		firmicutes 10 leaves
		# Bacillus sp. GN-12 partial 16S rRNA gene, strain GN-12
		^a Bacillus sp. DB14629 16S ribosomal RNA gene, partial sequence
		^a Uncultured Bacillus sp. gene for 16S ribosomal RNA, partial sequence, isolate: QX-17
		9 Bacillus sp. GIGAN2 165 nbosonal RNA gene, partial sequence
		9 Bacillus Duringiensis Strain 94-27, complete genome Bacillus Duringiensis strain 94-27, complete genome
		 Bacing interregions stars (10.51), complete periode
		 Bacillas antinacis strain BATOAS, complete genome Bacillas asthracis strain Partous consulta according
		Provide a manager statement of the Party of
		Bacillus anthracis strain Poline sequence
		Bacillus Burineiensis strain SR115-B716S ribosonal RNA cene, partial senarose
		 Bacillus cereus strain S6SB16 16S ribosonal RNA eene, natial secuence
		Bacillus cercus partial 16S rRNA gene, strain Cc-1
		Bacillus cereus strain Js16 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone K245_31 16S ribosomal RNA gene, partial sequence
		Bacillus cereus strain CASMBAUDALI 16S ribosomal RNA gene, partial sequence
		Bacillus cereus strain MGH118 16S ribosomal RNA gene, partial sequence
		Placifius cereus strain 165PP 165 ribosonal RNA gene, partial sequence
		Circultured bacterium clone 11 St_40 16S ribosonial RNA gene, partial sequence
		 Uncultured bacterium clone 11 St. 10 105 ribosomal RNA gene, partial sequence
		y Uncurated pacterian close 11 K, 20 10S mosterial RNA gene, partial sequence D language bacterian close 11 K and 13 10S means 10V cores and the second second br>second second s
1072		Uncountered protection charge 11 Ann. 25 155 fitostonial KNA gene, partial sequence
		S can share a bases
100 m		The process of the second

firmicutes | 5 leaves Bacillus thuringiensis strain MS8 165 ribosomal RNA gene, partial sequence

Bacillus subtilis DQ683077.1

Bacillus subtilis strain GB03 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb[DQ683077.1]</u> Length: 1124 Number of Matches: 1

Range 1:	Range 1: 26 to 1004 GenBank Graphics Vext Match A Previous Match								
Score 1739 bits	s(1928)	Expect 0.0	Identities 973/979(99	9%)	Gaps 0/979(0%)	Stra) Plus	nd /Plus		
Query	1	AGATGGGAGCTT	GCTCCCTG	ATGTTAGC	GCGGACG	GGTGAGTA	ACACGTGG	GTAACCTG	60
Sbjct	26	AGATGGGAGCTT	GCTCCCTG	ATGTTAGC	GCGGACG	GGTGAGTA	ACACGTGG	GTAACCTG	85
Query	61	CCTGTAAGACTG	GGATAACT			AATACCGG	ATGGTTGT	TGAACCG	120
Sbjct	86	CCTGTAAGACTG	GGATAACT	CCGGGAAA	CCGGGGGCT	AATACCGG	ATGGTTGT	TGAACCG	145
Query	121	CATGGTTCAGAC	ATAAAAGG	TGGCTTCG	GCTACCAC	TTACAGAT	GGACCCGCC	GCGCATT	180
Sbjct	146	CATGGTTCAGAC	ATAAAAGG	TGGCTTCG	GCTACCAC	TTACAGAT	GGACCCGCC	GCGCATT	205
Query	181	AGCTAGTTGGTG	AGGTAACG	GCTCACCA	AGGCGACG	ATGCGTAG	CCGACCTG	AGAGGGTG	240
Sbjct	206	AGCTAGTTGGTG	AGGTAACG	GCTCACCA	AGGCGACG	ATGCGTAG	CCGACCTG	AGAGGGTG	265
Query	241	ATCGGCCACACT	GGGACTGA	GACACGGC	CCAGACTC	CTACGGGA	GGCAGCAG	FAGGGAAT	300
Sbjct	266	ATCGGCCACACT	GGGACTGA	GACACGGC	CCAGACTC	CTACGGGA	GGCAGCAG	TAGGGAAT	32 <mark>5</mark>
Query	301	CTTCCGCAATGG	ACGAAAGT	CTGACGGA	GCAACGCC	GCGTGAGT	GATGAAGG	ITTTCGGA	360
Sbjct	326	CTTCCGCAATGG	ACGAAAGT	CTGACGGA	GCAACGCC	GCGTGAGT	GATGAAGG	ITTTCGGA	38 <mark>5</mark>
Query	361	TCGTAAAGCTCT	GTTGTTAG	GGAAGAACA	AAGTGCCG	TTCAAATA	GGGCGGCA	CCTTGACG	420
Sbjct	386	TCGTAAAGCTCT	GTTGTTAG	GGAAGAACA	AAGTGCCG	TTCAAATA	GGGCGGCA	CCTTGACG	445
Query	421	GTACCTAACCAG	AAAGCCAC	GCTAACTA	ACGTGCCA	GCAGCCGC	GGTAATAC	GTAGGTGG	480
Sbjct	446	GTACCTAACCAG	AAAGCCAC	GCTAACT	ACGTGCCA	GCAGCCGC	GGTAATAC	GTAGGTGG	<mark>505</mark>
Query	481	CAAGCGTTGTCC	GGAATTAT	IGGGCGTA	AAGGGCTC	GCAGGCGG	TTTCTTAA	GTCTGATG	540
Sbjct	506	CAAGCGTTGTCC	GGAATTAT	IGGGCGTA	AAGGGCTC	GCAGGCGG	TTTCTTAA	GTCTGATG	565
Query	541	TGAAAGCCCCCG	GCTCAACCO	GGGGAGGG	CATTGGA		AACTTGAGI	GCAGAAG	600
Sbjct	566	TGAAAGCCCCCG	GCTCAACCO	GGGGAGGG	CATTGGA	AACTGGGG	AACTTGAGI	GCAGAAG	625
Query	601	AGGAGAGTGGAA	TTCCACGT	GTAGCGGT	GAAATGCG1	FAGAGATG	rggaggaac	ACCAGTG	660
Sbjct	626	AGGAGAGTGGAA	TTCCACGT	GTAGCGGT	GAAATGCGI	TAGAGATG	IGGAGGAAC	ACCAGTG	685
Query	661								720
Ouerv	721	GCGAAGGCGACT	CTCTGGTC:		ACGUTGAG			GCGTTTC	745
Sbict	746	GGATTAGATACC	CTGGTAGT		TAAACGAT			 GGGTTTC	805
Query	781	CGCCCCTTAGTG	CTGCAGCT	AACGCATT	AGCACTCO	CGCCTGGG	GAGTACGGI	CGCAAGA	840
Sbjct	806	CGCCCCTTAGTG	IIIIIII CTGCAGCTA	AACGCATT	AGCACTCO	IIIIIII CGCCTGGGG		CGCAAGA	865
Query	841	CTGAAACTCAAA	GGAATTGA	ceeeecco	GCACAAG	CGGTGGAG	CATGTGGTI	TAATTCG	900
Sbjct	866	CTGAAACTCAAA	GGAATTGA	Ceeeecco	GCACAAG	CGGTGGAG	CATGTGGTT	TAATTCG	925
Query	901	AAGCAACGCGAA	AACCTTAC	CAGGTCTT	GACATCCT	CTGACATCO		AGGACGTC	960
Sbjct	926	AAGCAACGCGAG	AACCTTAC	CAGGTCTTC	SACATCCT	CTGACATCO	CTAGAGATA	AGGACGTC	985
Query	961	CCCTTCCGGGGC	AAAATGA	979					
Sbjct	986	CCCTTCGGGGGGC	AGAGTGA	1004					

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Bacillus sp. YG06290174 16S	ribosomal RNA gene, partial sequence
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	firmicutes 3 leaves
	Pacillus sp. PGPR 4 16S ribosomal RNA gene, partial sequence
	Bacillus sp. S22829 16S ribosomal RNA gene, partial sequence
	Bacillus sp. S22219 16S ribosomal RNA gene, partial sequence
	Bacillus sp. S122907 16S ribosomal RNA gene, partial sequence
	 Bacillus sp. RA-19 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain H34 16S ribosomal RNA gene, partial sequence
	Bacillus sp. XR 16S ribosomal RNA gene, partial sequence
	Bacillus SP, XJZ123 16S ribosomal RNA gene, partial sequence
	 Bacillus methylotrophicus strain SCSB 1312 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain SCDB 1365 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone L63 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain CB1 16S ribosomal RNA gene, partial sequence
	Bacillus sp. BDRC3 16S ribosomal RNA gene, partial sequence
	Bacillus amvlolique faciens strain FS6 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain KFRI-GS4-1 16S ribosomal RNA gene, partial sequence
	Bacillus sp. LS-508 16S ribosomal RNA gene, partial sequence
	 Bacillus amylolique faciens strain Ab-525 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain HT-22 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens gene for 16S ribosomal RNA, partial sequence, strain: RNB-113
	Bacillus sp. OD-4 16S ribosomal RNA gene, partial sequence
	 Bacillus methylotrophicus strain WZ-4 16S ribosomal RNA gene, partial sequence
	Bacillus sp. ZYJ-11 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain X-17 16S ribosomal RNA gene, partial sequence
	Bacillus amvlolique faciens strain CSY20 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain CSA26 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain CSL22 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain CSS13 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain CSM11 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain CSS7 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone AGGG 16S ribosomal RNA gene, partial sequence
	Bacillus sp. enrichment culture clone DSFB301 16S ribosomal RNA gene, partial sequence
	Bacillus sp. enrichment culture clone ZSTA204 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain STF NIO 3 16S ribosomal RNA gene, partial sequence
	Bacillus sp. H405 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain Z2 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain Z9 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain F29 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain MNNIT SR18 16S ribosomal RNA gene, partial sequence
	Bacillus sp. w-1(2014) 16S ribosomal RNA gene, partial sequence
0.0006	Bacillus amylolique faciens strain w-2 16S ribosomal RNA gene, partial sequence
	Bacterium DS-1 16S ribosomal RNA gene, partial sequence
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Bacillus cereus JQ659737.1

Bacillus cereus strain R5-339 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb[JQ659737.1]</u> Length: 1492 Number of Matches: 1

Range 1:	Range 1: 100 to 1025 GenBank Graphics V Next Match 🔺 Previous Match						
Score 1631 bit	s(1808)	Expect Ider 0.0 918	ntities /926(99%)	Gaps 1/926(0%)	Strand Plus/Plus		
Query	1	ACGGGTGAGTAACAC	STGGGTAACCTG	CCCATAAGACTO	GGATAACTCCGGG	AAACCGGG	60
Sbjct	100	ACGGGTGAGTAACAC	GTGGGTAACCTG	CCATAAGACTO	GGATAACTCCGGG	AAACCGGG	159
Query	61	GCTAATACCGGATAA	CATTTTGAACCG	CATGGTTCGAAA	TTGAAAGGCGGCT	TCGGCTGT	120
Sbjct	160	GCTAATACCGGATAA	CATTTTGAACCG	CATGGTTCGAAA	TTGAAAGGCGGCT	TCGGCTGT	219
Query	121	CACTTATGGATGGAC	CCGCGTCGCATT	AGCTAGTTGGTG	AGGTAACGGCTCA	CCAAGGCA	180
Sbjct	220	CACTTATGGATGGAC	CGCGTCGCATT	AGCTAGTTGGTG	AGGTAACGGCTCA	CCAAGGCA	279
Query	181	ACGATGCGTAGCCGA	CTGAGAGGGTG	ATCGGCCACACI	GGGACTGAGACAC	GGCCCAGA	240
Sbjct	280	ACGATGCGTAGCCGA	CTGAGAGGGTG	ATCGGCCACACI	GGGACTGAGACAC	GGCCCAGA	339
Query	241	CTCCTACGGGAGGCA	GCAGTAGGGAAT	CTTCCGCAATGG	ACGAAAGTCTGAC	GGAGCAAC	300
Sbjct	340	CTCCTACGGGAGGCA	GCAGTAGGGAAT	CTTCCGCAATGG	ACGAAAGTCTGAC	GGAGCAAC	399
Query	301	GCCGCGTGAGTGATG	AGGCTTTCGGG	CGTAAAACTCI	GTTGTTAGGGAAG	AACAAGTG	360
Sbjct	400	GCCGCGTGAGTGATG	AGGCTTTCGGG	ICGTAAAACTCI	GTTGTTAGGGAAG	AACAAGTG	459
Query	361	CTAGTTGAATAAGCT	GCACCTTGACG	STACCTAACCAG	AAAGCCACGGCTA	ACTACGTG	420
Sbjct	460	CTAGTTGAATAAGCT	GCACCTTGACG	STACCTAACCAG	AAAGCCACGGCTA	ACTACGTG	519
Query	421	CCAGCAGCCGCGGTA	ATACGTAGGTGG	CAAGCGTTATCO	GGAATTATTGGGC	GTAAAGCG	480
Sbjct	520	CCAGCAGCCGCGGTA	ATACGTAGGTGG	CAAGCGTTATCC	GGAATTATTGGGC	GTAAAGCG	579
Query	481	CGCGCAGGTGGTTTC	TAAGTCTGATG	GAAAGCCCACG	GCTCAACCGTGGA	GGGTCATT	540
Sbjct	580	CGCGCAGGTGGTTTC	TAAGTCTGATG	GAAAGCCCACG	GCTCAACCGTGGA	GGGTCATT	639
Query	541	GGAAACTGGGAGACT	FGAGTGCAGAAG	AGGAAAGTGGAA	ATTCCATGTGTAGC	GGTGAAAT	600
Sbjct	640	GGAAACTGGGAGACT	TGAGTGCAGAAG	AGGAAAGTGGAA	ATTCCATGTGTAGC	GGTGAAAT	699
Query	601	GCGTAGAGATATGGA	GGAACACCAGTG	GCGAAGGCGAC	TTCTGGTCTGTAA	CTGACACT	660
Sbjct	700	GCGTAGAGATATGGA	GGAACACCAGTG	GCGAAGGCGAC	TTCTGGTCTGTAA	CTGACACT	759
Query	661	GAGGCGCGAAAGCGT	GGGGAGCAAACA	GGATTAGATACO	CTGGTAGTCCACG	CCGTAAAC	720
Sbjct	760	GAGGCGCGAAAGCGT	GGGGAGCAAACA	GGATTAGATACO	CTGGTAGTCCACG	CCGTAAAC	819
Query	721	GATGAGTGCTAAGTG	TAGAGGGTTTC	CGCCCTTTAGT	SCTGAAGTTAACGC	ATTAAGCA	780
Sbjct	820	GATGAGTGCTAAGTG	TAGAGGGTTTC	CGCCCTTTAGTO	SCTGAAGTTAACGC	ATTAAGCA	879
Query	781	CTCCGCCCTGGGGAG	PACGGCCGCAAG	GCTGAAACTCCA	AGGAATTGACGGG	GGCCCGCA	840
Sbjct	880	CTCCGCCCTGGGGAG	TACGGCCGCAAG	GCTGAAACTCAA	AGGAATTGACGGG	GGCCCGCA	939
Query	841	CAAGCGGTGGAGCAT	GTGGTTTAATTC	GAAGC-ACCCGA	AAAACCTTACCAG	GTCTTGAC	899
Sbjct	940	CAAGCGGTGGAGCAT	GTGGTTTAATTC	GAAGCAACGCGZ	AGAACCTTACCAG	GTCTTGAC	999
Query	900	TTCCTCTGACAACCC	TAAAAATAGGG	925			
Sbjct	1000	ATCCTCTGAAAACCC	TAGAGATAGGG	1025			

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Interface and the second se	3		 Bacillus anthracis strain K3, complete genome
1000 100			Bacillus thuringiensis strain HD1011, complete genome
1006			 Bacillus sp. DB14780 16S ribosomal RNA gene, partial sequence
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 Ubcellured Bacillus en, gene for 165 rhosonal RNA, partal sequence, holdar CV, 17 Bacillus environ strain VIIIS (166 rhosonal RNA pene, partal sequence) Bacillus environ strain VIIIS (167 rhosonal RNA pene, partal sequence) Bacillus environ strain VIIIS (167 rhosonal RNA pene, partal sequence) Bacillus anthrasis (168 RNA pene, partal sequence) Bacillus anthrasis (167 RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus correas strain XIII (167 Rhosonal RNA pene, partal sequence) Bacillus coreas strain XIII (167 Rhosonal RNA pene, partal sequence)			Uncultured Bacillus sp. gene for 16S ribosomal RNA, partial sequence, isolate: LZX-37
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 Bacillas ceres train p50 166 ribosonal RNA gene, partial sequence Bacillas ceres train p50 166 ribosonal RNA gene, partial sequence Bacillas antinacia 166 ribosonal RNA gene, partial sequence Bacillas errors train 2022 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1272 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1272 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1272 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1272 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1272 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1272 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 1581 166 ribosonal RNA gene, partial sequence Bacillas ceres train 158 ribosonal RNA gene, partial sequence Bacillas ceres train 158 ribosonal RNA gene, partial sequence Bacillas ceres train 158 ribosonal RNA gene, partial sequence Bacillas ceres train 158 ribosonal RNA gene, partial sequence Bacillas ceres train 158 ribosonal RNA gene, partial sequence Bacillas ceres train 158 ribosonal RNA gene, partial sequence Bacillas ceres train 158 ribosonal RNA gene, partial sequence			Pacillus cereus strain VII52 16S ribosomal RNA gene, partial sequence
 Bacillas sp. PMO 166 ribosomal RNA gene, partial sequence Bacillas sp. PMO 166 ribosomal RNA gene, partial sequence Bacillas sp. PMO 166 ribosomal RNA gene, partial sequence Bacillas sp. PMO 166 ribosomal RNA gene, partial sequence Bacillas sp. PMO 166 ribosomal RNA gene, partial sequence Bacillas sp. Chen RL-111 RS ribosomal RNA gene, partial sequence Bacillas sp. Chen RL-111 RS ribosomal RNA gene, partial sequence Bacillas sp. Chen RL-111 RS ribosomal RNA gene, partial sequence Bacillas sp. Chen RL-111 RS ribosomal RNA gene, partial sequence Bacillas sp. Chen RL-111 RS ribosomal RNA gene, partial sequence Bacillas strain SR15 S15 166 ribosomal RNA gene, partial sequence Bacillas strain RD-166 ribosomal RNA gene, partial sequence Bacillas cercus train SR15 166 ribosomal RNA gene, partial sequence Bacillas cercus train SR15 166 ribosomal RNA gene, partial sequence Bacillas cercus train SR15 RS1 RS1 RS ribosomal RNA gene, partial sequence Bacillas cercus train SR15 166 ribosomal RNA gene, partial sequence Bacillas cercus train SR15 RS1 RS1 RS1 RS1 RS1 RS1 RS1 RS1 RS1 RS1		Ť	Bacillus cereus strain g50 16S ribosomal RNA gene, partial sequence
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1006 Bicilla se, GGAN2 HS rhoomal RNA gene, parial sequence Bicilla se, GGAN2 HS rhoomal RNA gene, parial sequence Cheather Bicilla versus train 212 165 rhoomal RNA gene, parial sequence Bicilla versus train SENIS-167 robord RNA gene, parial sequence Bicilla versus train SENIS-167 robord RNA gene, parial sequence Bicilla versus train SENIS-167 robord RNA gene, parial sequence Bicilla versus train SENIS-167 robord RNA gene, parial sequence Bicilla versus train SONIS-16 robord RNA gene, parial sequence Bicilla versus train SONIS-16 robord RNA gene, parial sequence Bicilla versus train SONIS-16 robord RNA gene, parial sequence Bicilla versus train SONIS-16 robord RNA gene, parial sequence Bicilla versus train SONIS-16 robord RNA gene, parial sequence Bicilla versus train SONIS-16 robord RNA gene, parial sequence Bicilla versus train SIS-16 robord RNA gene, parial sequence Bicilla versus train SIS-16 robord RNA gene, parial sequence Bicilla versus train SIS-16 robord RNA gene, parial sequence Bicilla versus train SIS-16 robord RNA gene, parial sequence Bicilla versus train SIS-16 robord RNA gene, parial sequence Bicilla versus train SIS-16 robord RNA gene, parial sequence Bicilla versus train SNA (sen. parial sequence Bicilla versus			Bacillus anthracis 16S ribosomal RNA gene, partial sequence
10005 11005 11005			Bacillus sp. GIGAN2 16S ribosomal RNA gene, partial sequence
10006			Bacillus cereus strain ZJU2 16S ribosomal RNA gene, partial sequence
10006 10006			Uncultured Bacillus sp. clone RL-11 I6S ribosomal RNA gene, partial sequence
10005 10005 10005 10005			Bacillus anthracis 16S rRNA gene, isolate EM1
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10005 10			Bacillus cereus strain Lr3/2 16S ribosomal RNA gene, partial sequence
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Dottom in the second			Bacillus cereus strain S6SB16 16S ribosomal RNA gene, partial sequence
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Staphylococcus haemolyticus KC139455.1

 $\label{eq:sequence} \begin{array}{l} Staphylococcus \ haemolyticus \ strain \ B-20 \ 16S \ ribosomal \ RNA \ gene, \ partial \ sequence \ sequence \ ID: \ \underline{gb|KC139455.1|} \ \ Length: \ 1428 \ \ Number \ of \ Matches: \ 1 \end{array}$

Range 1:	Range 1: 508 to 1366 GenBank Graphics Vext Match A Previous Match						
Score 1503 bits	s(1666)	Expect I 0.0 8	dentities 52/862(99%)	Gaps 3/862(0%)	Strand Plus/Minus		
Query	1	TGGTGTGACGGGC	GGTGTGTACAAGA	CCCGGGAACGT	ATTCACCGTAGCATG	CTGATCTA	60
Sbjct	1366	TGGTGTGACGGGC	GGTGTGTGTACAAGA	CCCGGGAACGT	ATTCACCGTAGCATG	CTGATCTA	1307
Query	61	CGATTACTAGCGA	TTCCAGCTTCATG	TAGTCGAGTTG	CAGACTACAATCCGA	ACTGAGAA	120
Sbjct	1306	CGATTACTAGCGA	TTCCAGCTTCATG	TAGTCGAGTTG	CAGACTACAATCCGA	ACTGAGAA	1247
Query	121		TTTGCTTGACCTC	GCGGTTTCGCT	ACCCTTTGTATTGTC	CATTGTAG	180
Sbjct	1246	CAACTTTATGGGA	ATTTGCTTGACCTC	GCGGTTTCGCT	ACCCTTTGTATTGTC	CATTGTAG	1187
Query	181	CACGTGTGTAGCO	CAAATCATAAGGG	GCATGATGATT	TGACGTCATCCCCAC	CTTCCTCC	240
Sbjct	1186	CACGTGTGTAGCO	CAAATCATAAGGG	GCATGATGATT	TGACGTCATCCCCAC	CTTCCTCC	1127
Query	241	GGTTTGTCACCGG	CAGTCAACTTAGA	GTGCCCAACTT	AATGATGGCAACTAA	GCTTAAGG	300
Sbjct	1126	GGTTTGTCACCGG	CAGTCAACTTAGA	GTGCCCAACTT	AATGATGGCAACTAA	.GCTTAAGG	1067
Query	301	GTTGCGCTCGTTG	CGGGACTTAACCC	AACATCTCACG	ACACGAGCTGACGAC	AACCATGC	360
Sbjct	1066	GTTGCGCTCGTTG	CGGGACTTAACCC	AACATCTCACG	ACACGAGCTGACGAC	AACCATGC	1007
Query	361	ACCACCTGTCACT	TTGTCCCCCGAAG	GGGAAAGCTCT	ATCTCTAGAGTTGTC	AAAGGATG	420
Sbjct	1006	ACCACCTGTCACI	TTGTCCCCCGAAG	GGGAAAGCTCT	ATCTCTAGAGTTGTC	AAAGGATG	947
Query	421	TCAAGATTTGGTA	AGGTTCTTCGCGT	TGCTTCGAATT	AAACCACATGCTCCA	CCGCTTGT	480
Sbjct	946	TCAAGATTTGGTA	AGGTTCTTCGCGT	TGCTTCGAATT	AAACCACATGCTCCA	CCGCTTGT	887
Query	481	GCGGGTCCCCGTC	AATTCCTTTGAGT	TTCANNCTTGC	GNTCGTACTCCCCAG	GCGGAGTG	540
Sbjct	886	GCGGGTCCCCGTC	AATTCCTTTGAGT	TTCAACCTTGC	GGTCGTACTCCCCAG	GCGGAGTG	827
Query	541	CTTAATGCGTTAG	CTGCAGCACTAAG	GGGCGGAAACC	CCCTAACACTTAGCA	CTCATCGN	600
Sbjct	826	CTTAATGCGTTAG	CTGCAGCACTAAG	GGGCGGAAACC	CCCTAACACTTAGCA	CTCATCGT	767
Query	601	TTACGGCGTGGAC	TACCAGGGTATCT	AATCCTGTTTG.	ATCCCCACNCTTTCG	CACATCAG	660
Sbjct	766	TTACGGCGTGGAC	TACCAGGGTATCT	AATCCTGTTTG.	ATCCCCACGCTTTCG	CACATCAG	707
Query	661	CGTCAGTTACAGA	CCAGAAAGTCGCC	TTCGCCACTGG	TGTTCCTCCATATCT	CTGCGCAT	720
Sbjct	706	CGTCAGTTACAGA	CCAGAAAGTCGCC	TTCGCCACTGG	TGTTCCTCCATATCT	CTGCGCAT	647
Query	721	TTCACCGCTACAC	ATGGAAATTCCAC	TTTCCTCTTCT	GCACTCAAGTTTTCC.	AGTTTCCA	780
Sbjct	646	TTCACCGCTACAC	ATGG-AATTCCAC	TTTCCTCTTCT	GCACTCAAGTTTTCC.	AGTTTCCA	588
Query	781	ATGACCCTCCACG	GTTGAGCCGTGGG	CTTTCCCATCC	GACTTAAAAAACCGC	CTACGCGC	840
Sbjct	587	ATGACCCTCCACG	GTTGAGCCGTGGG	CTTTCACATCA	GACTTAAAAAACCGC	CTACGCGC	528
Query	841	GCCTTTACGCCCA	ATAAATTCC 86	2			
Sbjct	527	G-CTTTACGCCCA	AT-AATTCC 50	8			

	*KiQuery 35409 *Human oral bacterium C20 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain Hb-6 16S ribosonal RNA gene, partial sequence Such laser haemolyticus strain B 17 16S ribosonal RNA gene, partial sequence
	Staphylococcus naemolyticus strain B-17 105 motional ROA gene, partial sequence 9 Staphylococcus petrasii subso, pragensis strain CCM 8529 16S ribosomal RNA gene, partial sequence
	Staphylococcus petrasii subsp. croceilyticus strain MCC11690 16S ribosomal RNA gene, partial sequence
	Bacterium NLAE-zI-P566 16S ribosomal RNA gene, partial sequence
	Staphylococcus periasi suoje, pragensis strain ENO 2528 ToS motionian ROV gene, partial sequence
	9 Staphylococcus haemolyticus strain B-20 16S ribosomal RNA gene, partial sequence
	Staphylococcus jettensis strain SEQ257 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 254/Chen 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain 383/Chen 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain Pn1016/Pant-Uttar/2007 16S ribosomal RNA gene, partial sequence Staphylococcus aureus subma aureus 11810-07 cornelate enorme.
	 Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000357 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000535 small subunit ribosomal RNA gene, partial sequence
	⁹ Uncultured organism clone EL00178-T478-S-NIPCRAMgANa 000494 small subunit ribosomal RNA gene, partial sequence Uncultured organism clone EL10034-T174-S-NIPCPAMgANa 000170 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL2003+T174-5-NPCRAMgANb (2003) 9 small subunit ribosonial ROA gene, partial sequence
	 Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000573 small subunit ribosomal RNA gene, partial sequence
	Staphylococcus aureus subsp. aureus stram XYL-Mo-1105 rhosomal RNA gene, partal sequence Staphylococcus aureus subsp. aureus stram XYL-Mo-1105 rhosomal RNA gene, partal sequence
	 Staphylococcus aureus strain 2221 165 ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain Z1092 16S ribosomal RNA gene, partial sequence
	 Staphylococcus autors strain 179 (RC10B) 165 tibsomal RNA gene, partial sequence Staphylococcus autors strain 179 (RC10B) 165 tibsomal RNA gene, partial sequence
	* Staphylococcus aureus subsp. aureus strain 92B (BSJ3) 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 129A (BC40Fs) 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain 140A (toc2) 105 ritosomat RNA genc, partial sequence Staphylococcus aureus strain 15 (RCID) 165 ritosomat RNA genc, partial sequence
	Staphylococcus aureus strain 29 (BR56) 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 74 (br12) 16S ribosomal RNA gene, partial sequence
	9 Staphylococcus aureus strain 94A (0r8) 105 ribosomal RNA gene, partial sequence 9 Staphylococcus sp. ChDC B590 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 2RMOY1 16S ribosomal RNA gene, partial sequence
	Saphylococcus aureus strain IS 1/2 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain KW-302 105 ribosoniai KNA gene, partiai sequence Staphylococcus aureus subsp. aureus DNA, complete genome, strain: TMUS2134
	Staphylococcus aureus strain OUC_Est2S 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus genome assembly NCTC13435, chromosome : 1
	 Staphylococcus aureus strain CICC 10384 165 ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain LH-Y1 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. SB-3 16S ribosomal RNA gene, partial sequence Stack-lococcus sp. SB-3 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain 89A (DR06) 10S ribosomal RNA gene, partial sequence Staphylococcus aureus strain 42 (C2TJ) 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 158B (BR61Fs) 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 147B (BR64Fs) 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain 130A (BR68) 16S ribosomal RNA gene, partial sequence Staphylococcus aureus strain 102 (BR65Er(A)) 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain 102 (BrCoTs(x)) Fos Hoosonial RNA gene, partial sequence Staphylococcus aureus strain 124B (BC38Fs) 16S ribosonial RNA gene, partial sequence
	Staphylococcus aureus strain O2 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain C123 16S ribosomal RNA gene, partial sequence
	 Saphylococcus aureus strain Z1201 105 molsomal RNA gene, partial sequence Sanbylococcus aureus strain KIBCE/ER24 165 molsomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain G18 16S ribosomal RNA gene, partial sequence
	Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000628 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL U0034-T174-S-NIPCRAM8ANb 000444 small subunit rbosomal RNA gene, partial sequence
	 Uncultured organism clone EL1003+1174-5-NFC KAMgANo 2001 / s mail subunit ribosoma RNA gene, partial sequence Uncultured organism clone EL1003+1714-5-NFC KAMgANo 2001 I small subunit ribosoma RNA sene, partial sequence
	Quncultured organism clone ELU0175-T455-S-NIPCRAMgANa_000843 small subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000359 small subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000082 small subunit ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain Sh29/312/L2, complete genome
	 Staphylococcus aureus subsp. aureus DNA, complete genome, strain: TMUS2126
	 Staphylococcus aureus subsp. aureus ST772-MRSA-V strain DAR4145, complete genome Staphylococcus aureus subsp. aureus GT720 A 165 (1)
	 Staphylococcus naemotyticus strain GZSF-3 105 ribosomal RNA gene, partial sequence Staphylococcus aureus strain AMRF2 16S ribosomal RNA gene, nartial sequence
	Staphylococcus sp. BAB-3242 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 89B (BR66) 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain 48 (br14) 16S ribosomal RNA gene, partial sequence Stabhylococcus aureus strain 166 (ARD/OF) 16S ribosomal RNA
	 Staphylococcus aureus strain 166A (BROOPS) 165 ribosonial RNA gene, partial sequence Staphylococcus aureus strain 148B (BC39Fs) 165 ribosonial RNA gene, partial sequence
	Staphylococcus aureus strain 139B (br27) 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain 118A (BR69Ms) 16S ribosomal RNA gene, partial sequence
	 Staphylococcus aureus strain 115B (BR67/is) 16S ribosomal RNA gene, partial sequence Staphylococcus aureus suber, aureus CNL complete aurous
	Staphylococcus aureus strain C126 16S ribosomal RNA eene, partial sequence
	Staphylococcus aureus strain Z1263 16S ribosomal RNA gene, partial sequence
	Staphylococcus aureus strain Z222 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. N2 partial 165 rRNA gene, strain N2 Uncultured organism clone FL DYRLT 172, SAIDCP AMAANS, 000662 cmall subunit electromal DNA come mariel commons.
	 Uncultured organism clone EL00034-T 174-S-NIPCRAMeAND 000500 small subunit ribosomal RNA gene, nartial sequence
	⁹ Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000310 small subunit ribosomal RNA gene, partial sequence
	⁹ Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000107 small subunit ribosomal RNA gene, partial sequence Allocational description of the Allocation of the Alloca
	 Uncultured organism clone ELU0177-1472-S-NPC KAMgANa_000516 small subunit ribosomal KNA gene, partial sequence Uncultured organism clone ELU0072-T477-S-NIPCRAMoANa_000382 small subunit ribosomal RNA gene, partial sequence
0.0009	Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa 000321 small suburit ribotomar RAVA gene, partial sequence
1 1	firmicutes 4 leaves
	Staphylococcus aureus strain RKI4, complete genome

3- The phylogenetic analysis of bacteria isolated from computer Keyboards and computer mice samples

Bacillus subtilis KJ746466.1

Bacillus subtilis strain ACHSOC779 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KJ746466.1</u>| Length: 1266 Number of Matches: 1

Range 1: 14 to 970 GenBank Graphics V Next Match 🔺 Previous Match							
Score 1701 bits	(1886)	Expect 0.0	Identities 952/957(99%)	Gaps 1/957(0%)	Strand Plus/Plus		
Query	1	CTGATGTTAGCG	GCGGACGGGTGAGTA	ACACGTGGGTAAC	CTGCCTGTAAGAC	TGGGATA	60
Sbjct	14	CTGATGTTAGCG	GGACGGGTGAGTA	ACACGTGGGTAAC	CTGCCTGTAAGAC	TGGGATA	73
Query	61	ACTCCGGGAAACC	GGGGCTAATACCGG	ATGGTTGTTTGAA	CCGCATGGTTCAA	ACATAAA	120
Sbjct	74	ACTCCGGGAAACC	GGGGCTAATACCGG	ATGGTTGTTTGAA	CCGCATGGTTCAA	ACATAAA	133
Query	121	AGGTGGCTTYGGC	TACCACTTACAGAI	GGACCCGCGGCGC	ATTAGCTAGTTGG	TGAGGTA	180
Sbjct	134	AGGTGGCTTCGGC	TACCACTTACAGAT	GGACCCGCGGCGC	ATTAGCTAGTTGG	TGAGGTA	193
Query	181	ACGGCTCACCAAG	GCAACGATGCGTAG	CCGACCTGAGAGG	GTGATCGGCCACA	CTGGGAC	240
Sbjct	194	ACGGCTCACCAAC	GCAACGATGCGTAG	CCGACCTGAGAGG	GTGATCGGCCACA	CTGGGAC	253
Query	241	TGAGACACGGCCC	AGACTCCTACGGGA		AATCTTCCGCAAT	GGACGAA	300
Sbjct	254	TGAGACACGGCCC	AGACTCCTACGGGA	GGCAGCAGTAGGG	AATCTTCCGCAAT	GGACGAA	313
Query	301	AGTCTGACGGAG	CAACGCCGCGTGAGI	GATGAAGGTTTTC	GGATCGTAAAGCT	CTGTTGT	360
Sbjct	314	AGTCTGACGGAG	CAACGCCGCGTGAGI	GATGAAGGTTTTC	GGATCGTAAAGCT	CTGTTGT	373
Query	361	TAGGGAAGAACAA	GTACCGTTCGAATA		ACGGTACCTAACC	AGAAAGC	420
Sbjct	374	TAGGGAAGAACAA	GTACCGTTCGAATA	GGGCGGTACCTTG	ACGGTACCTAACC	AGAAAGC	433
Query	421	CACGGCTAACTAC	GTGCCAGCAGCCGC	GGTAATACGTAGG	TGGCAAGCGTTGT	CCGGAAT	480
Sbjct	434	CACGGCTAACTAC	GTGCCAGCAGCCGC	GGTAATACGTAGG	TGGCAAGCGTTGT	CCGGAAT	493
Query	481	TATTGGGCGTAA		TTTCTTAAGTCTG	ATGTGAAAGCCCC	CGGCTCA	540
Sbjct	494	TATTGGGCGTAA	AGGGCTCGCAGGCGG	TTTCTTAAGTCTG	ATGTGAAAGCCCC	CGGCTCA	553
Query	541	ACCGGGGGAGGGT		BAACTTGAGTGCAG	AAGAGGAGAGTGG	AATTCCA	600
Sbjct	554	ACCGGGGGAGGGT	CATTGGAAACTGGGG	SAACTTGAGTGCAG	AAGAGGAGAGTGG	AATTCCA	613
Query	601	CGTGTAGCGGTG	AAATGCGTAGAGATG	TGGAGGAACACCA	GTGGCGAAGGCGA	CTCTCTG	660
Sbjct	614	CGTGTAGCGGTG	AAATGCGTAGAGATG	STGGAGGAACACCA	GTGGCGAAGGCGA	CTCTCTG	673
Query	661	GTCTGTAACTGA	CGCTGAGGAGCGAAA	AGCGTGGGGGAGCGA	ACAGGATTAGATA	CCCTGGT	720
Sbjct	674	GTCTGTAACTGA	CGCTGAGGAGCGAAA	AGCGTGGGGGAGCGA	ACAGGATTAGATA	CCCTGGT	733
Query	721	AGTCCACGCCGT	AAACGATGAGTGCT#	AGTGTTAGGGGGT	TTCCGCCCCTTAG	TGCTGCA	780
Sbjct	734	AGTCCACGCCGT	AAACGATGAGTGCTA	AGTGTTAGGGGGT	TTCCGCCCCTTAG	TGCTGCA	793
Query	781	GCTAACGCATTA		GAGTACGGTCGCA	AGACTGAAACTCA	AAGGAAT	840
Sbjct	794	GCTAACGCATTA	AGCACTCCGCCTGGG	GAGTACGGTCGCA	AGACTGAAACTCA	AAGGAAT	853
Query	841	TGAC-GGGGCCCC				AAGAACC	899
Sbjct	854	TGACGGGGGCCC	GCACAAGCGGTGGAG	JCATGTGGTTTAAT	TCGAAGCAACGCG	AAGAACC	913
Query	900		JACATCCTCTGACA7	CCTAAAAATAGGA		GGCA 95	6
Sbjet	914	TTACCAGGTCTT	JACATCCTCTGACAT	CCTAGAGATAGGA	CGTCCCCTTCGGG	GGCA 970	U

	⇒ lclQuery_18947
4	Bacillus sp. SSKSD11 I6S ribosomal RNA gene, partial sequence
	Bacillus sp. V4 I6S ribosonal RNA gene, partial sequence
	Bacillus methylotrophicus 165 ribosomal RNA gene, partial sequence
	Bacillus sublitis strain W2-3 165 ribosomal RNA gene, partial sequence
	Bacillus sublilis strain B11 16S ribosomal RNA gene, partial sequence
	Bacillus sublitis strain SAU-Ab-9 165 ribosomai RNA gene, partial sequence
	Bacillus sublitis strain L11 IbS infosomai RNA gene, partiai sequence
	Bacillus subilis strain Barrol ToS ribosomat RivA gene, partial sequence
	Bacillus sp. VIIN2 IoS ribosonal RNA gene, partial sequence
	Bacillus sublinis strain A 102 105 fitosomai RNA gene, partai sequence
	Protecting submits strain G10-99 105 motosima Riva gene, partial sequence
	Basilus advisiti attai CCO (CC) (S altra sequence
	Bacillus sublinis strain GG62 (165 Rhosenal RNA gene, partial sequence Deallus sublinis first and GCPR LAAL (65 Rhosenal RDA) areas and
	 Basilius gubilius subilitis strain SCKD1444 105 trobsomat KVA gene, partial sequence Basilius gubilitis strain SCKD1441165 tribatemal BNA gene activitie acquirece
	 Bacillus submis submis scale solution into solutional reverse gene, partial sequence Bacillus formitaria sona for 165 PDAA and its accuracy at the solution of the s
	Dealling requirements gene for 105 ININA, partial sequence, stanti 55-1
	Bacillus equificients gene no ros reivas, parta sequence, statili reiza
	Bacillus submis suam CH12 (05 Hospital KVA gene partial sequence
	Bacillus solutils strain Child IS ribasonal New Joint, para sequence
	Bacillus es or TVH.2 105 following black and the state of
	Bacillus or 21 YHz 165 ribosomal RVA gene partial sequence
	Bacillus aprinting statistics for the statistical statistics of the statistic sequence
	Bacillus subilis strain PARIS To Prosonal RAN gene participance
	Bacillus so HE08 I6S ribosomal RNA sene partial sequence
	Bacterium Y2 16S tribosonial RNA gene nartial sequence
	Bacterium YC-LK-LK144 165 ribosomal RNA sene, nartial sequence
	Bacillus sp. CEB332.1 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain SANN3 I6S ribosomal RNA gene, partial sequence
	Bacterium YC-LK-LKJ121 16S ribosomal RNA cene, partial sequence
	Bacterium YC-LK-LKJ43 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain DL47 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain PPL-SC9 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain Lys-1249 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain jk-45 16S ribosomal RNA gene, partial sequence
	Bacillus sp. ZLXH-3 I6S ribosomal RNA gene, partial sequence
	Bacillus subtilis subsp. inaquosorum strain IHB B 7075 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain IC3 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain CH13 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone SA2 16S ribosomal RNA gene, partial sequence
	📬 firmicutes 20 leaves
	Pacillus subtilis subsp. subtilis strain DSM 10 16S ribosomal RNA gene, partial sequence
.0.0006	Bacillus subtilis strain RAS-1 16S ribosomal RNA gene, partial sequence
0.000	Bacterium YC-LK-LKJ45 16S ribosomal RNA gene, partial sequence
1	Firmicutes 36 leaves
	Bacillus subtilis strain W6 16S ribosomal RNA gene, partial sequence

Bacillus amyloliquefaciens AB301004.1

Bacillus amyloliquefaciens gene for 16S rRNA, partial sequence, strain: GH3 Sequence ID: <u>dbj|AB301004.1</u>| Length: 1481 Number of Matches: 1

Range 1: 447 to 1408 GenBank Graphics V Next Match 🛦 Previous Match				
Score 1709 bit	s(1894)	ExpectIdentitiesGapsStrand0.0957/962(99%)1/962(0%)Plus/Minus	_	
Query	1	GTGTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAA	CGTATTCACC	60
Sbjct	1408	GTGTTACAAACTCTCGTGGTGTGGACGGGCGGTGTGTACAAGGCCCGGGAA	CGTATTCACC	1349
Query	61	GCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAG	TTGCAGACTG	120
Sbjct	1348	GCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAG	TTGCAGACTG	1289
Query	121	CGATCCGAACTGAGAACAGATTTGTGGGATTGGCTTAACCTCGCGGTTTC	GCTGCCCTTT	180
Sbjct	1288	CGATCCGAACTGAGAACAGATTTGTGGGATTGGCTTAACCTCGCGGTTTC	GCTGCCCTTT	1229
Query	181	GTTCTGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATG	ATTTGACGTC 2	240
Sbjct	1228	GTTCTGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATG	ATTTGACGTC	1169
Query	241	ATCCCCACCTTCCTCCGGTTTGTCACCGGCAGTCACCTTAGAGTGCCCAA	CTGAATGCTG	300
Sbjct	1168	ATCCCCACCTTCCTCCGGTTTGTCACCGGCAGTCACCTTAGAGTGCCCAA	CTGAATGCTG	1109
Query	301	GCAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTC	ACGACACGAG	360
Sbjct	1108	GCAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTC	ACGACACGAG	1049
Query	361	CTGACGACAACCATGCACCACCTGTCACTCTGCCCCCGAAGGGGACGTCC	TATCTCTAGG 4	420
Sbjct	1048	CTGACGACAACCATGCACCACCTGTCACTCTGCCCCCGAAGGGGACGTCC	TATCTCTAGG	989
Query	421	ATTGTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAAT	TAAACCACAT	480
Sbjct	988	ATTGTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAAT	TAAACCACAT	929
Query	481	GCTCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGTCTTG	CGACCGTACT 5	540
Sbjct	928	GCTCCACCGCTTGTGCGGGGCCCCCGTCAATTCCTTTGAGTTTCAGTCTTG	CGACCGTACT 8	869
Query	541	CCCCAGGCGGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAAC	CCCCTAACAC (600
Sbjct	868	CCCCAGGCGGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAAC	CCCCTAACAC	809
Query	601	TTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTC	GCTCCCCACG	660
Sbjct	808	TTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTC	GCTCCCCACG	749
Query	661	CTTTCGCTCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTG	GTGTTCCTCC	720
Sbjct	748	CTTTCGCTCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTG	GTGTTCCTCC	689
Query	721	ACATCTCTACGCATTTCACCGCTACACGTGGAATTCCACTCTCCTCTTCT	GCACTCAAGT	780
Sbjct	688	ACATCTCTACGCATTTCACCGCTACACGTGGAATTCCACTCTCCTCTTCT	GCACTCAAGT	629
Query	781	TCCCCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGGCTTTCACATCA		840
Sbjct	628	TCCCCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTCACATCA	GACTTAAGAA	569
Query	841	CCGCCTGCGAGCCCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCT	ACGTATTACC	900
Sbjct	568	CCGCCTGCGAGCCCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCT	ACGTATTACC	509
Query	901	GCGGCTGCTGGCACGTAGTTAGCCG-GGCTTTCTGGTTAGGTACCGTCAA	GGGGCCCCCC	959
Sbjct	508	GCGGCTGCTGGCACGTAGTTAGCCGTGGCTTTCTGGTTAGGTACCGTCA	GGTGCCGCCC	449
Query	960	TA 961		
Sbjct	448	 TA 447		

	firmicutes 2 leaves
	Bacillus amylolique faciens strain GR53 16S ribosomal RNA gene, partial sequence
	Bacillus sp. H405 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain T45 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain Z7 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain D12 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain D48 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain MNNIT SR18 16S ribosomal RNA gene, partial sequence
	Pacterium DS-1 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain SDS14 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain August M2 16S ribosomal RNA gene, partial sequence
	Bacillus sp. LG 16S ribosomal RNA gene, partial sequence
	Bacterium OKR 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain IHB B 7126 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain MSEB 18 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain HB24 16S ribosomal RNA gene, partial sequence
	Bacillus vallismortis strain JBS-10 16S ribosomal RNA gene, partial sequence
	Bacillus sp. APG-1 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain KAVK1 16S ribosomal RNA gene, partial sequence
	Bacillus sp. 825 I6S ribosomal RNA gene, partial sequence
	Bacterium YC-LKJ56 I6S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain WY-3 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain CY14 16S ribosomal RNA gene, partial sequence
	Bacillus sp. YP1(2015) 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique facients strain Ixnuwy-1 CCTCC M 2014639 16S ribosomal RNA gene partial sequence
	P Bacillus subtilis strain L18 16S riboomal RNA gene narial sequence
	Bacillus amylolique facients substra plantarium strain Pro-32 16S ribosomal RNA gene, partial sequence
	 Bacillus valiemoria strain NBIE/01116S ribosomal RNA oran postoniar recercipartial sequence
	Bacillus anylolicus facients strain XVI 16S ribosomal RNA gene, partial sequence
	Bacterium VC-LK10 165 ribosomal PNA gene partial sequence
	Genious 121 Laure
	Bacillus on VDS 165 sibasomal DNA oana partial samanca
	T bachnus sp. 175 tos nosoniai KVA gene, panar sequence
	Decilius and oliginal focience therein X S 2 16 sibosomal DNA care partial sequence
	P Datarius any longue laters strain AS-2 105 morsonial KIVA gene, partial sequence
	Spacerum (C-LKJO) tos noosonai Kiva gene, paniai sequence
	armicules 25 leaves
	Bacillus amytoiduetaciens subsp. piantarum strain 1-1-b-5 16S ribosomal KNA gene, partial sequence
	Bacilius sp. DB14353 IoS ribosomal RNA gene, partial sequence
	Bacillus amyloliquefacients strain WJ-2 IoS ribosomal RNA gene, partial sequence
	Bacillus sp. PPM3 165 ribosomal RNA gene, partial sequence
	Bacterium YC-LKJ2 165 ribosomal RNA gene, partial sequence
	Bacillus sp. HU-2012 I6S ribosomal RNA gene, partial sequence
	Bacillus amytolique faciens strain 1D-7 16S ribosomal RNA gene, partial sequence
	Bacillus amytolique faciens strain Lys-1436 IoS ribosomal RNA gene, partial sequence
	Bacillus subtilis strain NLX4 16S ribosomal KNA gene, partial sequence
	Bacillus methylotrophicus strain HB26 16S ribosomal RNA gene, partial sequence
	Bacillus methylotrophicus strain HB11 I6S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens subsp. plantarum strain IHB B 12506 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain IHB B 7050 16S ribosomal RNA gene, partial sequence
	Bacillus sp. BAB-650 16S ribosomal RNA gene, partial sequence
	Bacillus vallismortis strain CM1E3 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain NLB12 16S ribosomal RNA gene, partial sequence
0004	Bacillus subtilis strain YA-3 16S ribosomal RNA gene, partial sequence
7.0004	Bacterium NJ2 16S ribosomal RNA gene, partial sequence
1	Bacillus amyloliquefaciens strain 1 16S ribosomal RNA gene, partial sequence
	Bacillus amylolique faciens strain Jxnuwx-1 CCTCC M 2014638 16S ribosomal RNA gene, partial sequence

Brevibacillus brostelensis EU816699.1

Range 1:	93 to 105	3 GenBank Graphics		1 🔻	Next Match 🔺 Previous Match	
Score 1656 bit	s(1836)	Expect 0.0	Identities 946/962(98%)	Gaps 2/962(0%)	Strand Plus/Plus	
Query	1	GTAACACGTAG	GCAACCTGCCCGT	AGCTCGGGATAAC	ATGGGGAAACTCATGCTAATAC	60
Sbjct	93	GTAACACGTAG	GCAACCTGCCCGT	AGCTCGGGATAAC	ATGGGGAAACTCATGCTAATAC	152
Query	61	CGGATAGGGTC	TTCTCTCGCATGA	GAGGAGACGGAAAG	GTGGCGCAAGCTACCACTTACC	÷ 120
Sbjct	153	CGGATAGGGTC	TCTCTCGCATGA	GGGAGACGGAAAG	GTGGCGCAAGCTACCACTTAC	÷ 212
Query	121	GATGGGCCTGC	GCGCATTAGCTA	STTGGTGGGGTAAC	GGCCTACCAAGGCGACGATGC	€ 180
Sbjct	213	GATGGGCCTGC	GCGCATTAGCTA	GTTGGTGGGGTAAC	GGCCTACCAAGGCGACGATGC	3 272
Query	181	TAGCCGACCTG	AGAGGGTGACCGG	CCACACTGGGACTG	AGACACGGCCCAGACTCCTAC	€ 240
Sbjct	273	TAGCCGACCTG	AGAGGGTGACCGG	CCACACTGGGACTG	AGACACGGCCCAGACTCCTAC	332
Query	241	GGAGGCAGCAG	PAGGGAATTTTCC	ACAATGGACGAAAG	TCTGATGGAGCAACGCCGCGT	300
Sbjct	333	GGAGGCAGCAG	PAGGGAATTTTCC	ACAATGGACGAAAG	TCTGATGGAGCAACGCCGCGT	÷ 392
Query	301	AACGATGAAGG	ICTTCGGATTGTA	AGTTCTGTTGTCA	GAGACGAACAAGTACCGTTCGA	A 360
Sbjct	393	AACGATGAAGG	ICTTCGGATTGTA	AGTTCTGTTGTCA	GAGACGAACAAGTACCGTTCGA	4 52
Query	361	ACAGGGCGGTA	CCTTGACGGTACC	IGACGAGAAAGCCA	CGGCTAACTACGTGCCAGCAGC	420
Sbjct	453	ACAGGGCGGTAG	CCTTGACGGTACC	IGACGAGAAAGCCA	CGGCTAACTACGTGCCAGCAGC	512
Query	421	CGCGGTAATAC	STAGGTGGCAAGC	GTTGTCCGGAATTA	TTGGGCGTAAAGCGCGCGCAG	4 80
Sbjct	513	CGCGGTAATAC	GTAGGTGGCAAGC	GTTGTCCGGAATTA	TTGGGCGTAAGGCGCGCGCAG	572
Query	481	CGGCTATGTAA	STCTGGTGTTAAA	GCCCGGGGGCTCAAC	CCCGGTTCGCATCGGAAACTG	540
Sbjct	573	CGGCTATGTAA	GTCTGGTGTTAAA	GCCCGGGGGCTCAAC	CCCGGTTCGCATCGGAAACTG	632
Query	541	GTAGCTTGAGT	GCAGAAGAGGAAA	GCGGTATTCCACGT	GTAGCGGTGAAATGCGTAGAGA	a. 600 I
Sbjct	633	GTAGCTTGAGT	GCAGAAGAGGAAA	GCGGTATTCCACGT	GTAGCGGTGAAATGCGTAGAGA	A 692
Query	601	TGTGGAGGAAC	ACCAGTGGCGAAG	GCGGCTTTCTGGTC	TGTAACTGACGCTGAGGCGCG	4 660 I
Sbjct	693	TGTGGAGGAAC	ACCAGTGGCGAAG	GCGGCTTTCTGGTC	TGTAACTGACGCTGAGGCGCG	A 752
Query	661	AAGCGTGGGGA		GATACCCTGGTAGT	CCACGCCGTAAACGATGAGTG	C 720
Sbjct	753	AAGCGTGGGGA	GCAAACAGGATTA	GATACCCTGGTAGT	CCACGCCGTAAACGATGAGTG	812
Query	721	TAGGTGGTggg	ggggTTTCAATAC	CCTCAGTGCCGCAG	CTAACGCAATAAGCACTCCGC	C 780
Sbjct	813	TAGGTGGTTGG	GGGGTTTCAATAC	CCTCAGTGCCGCAG	CTAACGCAATAAGCACTCCGC	872
Query	781	TGGGGAGTACG	CTCGCAAGAGTGA	AACTCAAAGGAATT	GACGGGGGGCCCGCACAAGCGG	3 840
Sbjct	873	TGGGGAGTACG	CTCGCAAGAGTGA	AACTCAAAGGAATT	GACGGGGGCCCGCACAAGCGG	r 932
Query	841	GGAGCATGGGG	ITTAATTCGAAGC	AACGCGAAGAACCT	TACCA-GTCTTGACATCCCGC	r 899
Sbjct	933	GGAGCATGTGG	TTTAATTCGAAGC	AACGCGAAGAACCT	TACCAGGTCTTGACATCCCGC	r 992
Query	900	GACCGTCCTAA	AAATAGGGCTTCC	CTTCGGGGGCAGCGG	GGAAgggggggggggATGGTGG	r 959
Sbjct	993	GACCGTCCTAG	AGATAGGGCTTCC	CTTCGGGGCAGCGG	TG-ACAGGTGGTGCATGGTTG	r 1051
Query	960	CG 961				
Sbjct	1052	CG 1053				

Brevibacillus borstelensis clone US12 16S ribosomal RNA gene, partial sequence Sequence ID: gbjEU816699.11 Length: 1529 Number of Matches: 1

	Rewinscillus on DR14566-16S ribosomal PNA gene partial sequence
	Previbacillus sp. EB14.00 Nos noosanal RNA gene, partial sequence Previbacillus sp. EB14.00 Nos noosanal RNA gene, partial sequence
	Revibacillus sp. R-31301 nartial 16S rRNA cene, strain R-31301
L	Brevibacillus panacihumi strain KB55 I6S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone L 147 16S ribosomal RNA gene, partial sequence
9	Bacterium KB37 16S ribosomal RNA eene, partial sequence
	Brevibacillus panacihumi strain CI7 IoS ribosonal RNA eene, partial sequence
Brevibaci	lus borstelensis strain LAMA 1184 16S ribosomal RNA gene, partial sequence
• • Bas	terium enrichment culture clone JH-2 16S ribosomal RNA gene, partial sequence
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L	firmicutes 2 leaves
	Rhodococcus rhodochrous strain CC14 16S ribosomal RNA gene, partial sequence
	firmicutes 3 leaves
	Brevibacillus borstelensis strain KNUC221 16S ribosomal RNA gene, partial sequence
	Brevibacillus borstelensis strain KNUC224 16S ribosomal RNA gene, partial sequence
	Pervibacillus sp. sjf_26 16S ribosomal RNA gene, partial sequence
	Brevibacillus sp. Sl64 gene for 16S ribosomal RNA, partial sequence
	Brevibacillus borstelensis strain AR9 16S ribosomal RNA gene, partial sequence
	firmicutes 3 leaves
	Brevibacillus borstelensis strain PBI 16S ribosomal RNA gene, partial sequence
	Brevibacillus borstelensis strain UTMIII 16S ribosomal RNA gene, partial sequence
	Brevibacillus borstelensis strain ZYJ-25 16S ribosomal KNA gene, partial sequence
	Wirevibacillus sp. 21-YC6800 16S ribosomal KNA gene, partial sequence
	Drevidacinus sp. E 105 ribosomat KivA gene, partial sequence
	Bachales pacterium INU55 fos fibosomal KINA gene, partial sequence Pravibacillus bastalansis statis NL LES ribosomal DNA care actical sequence
	Drevidachius borsiciensis siran INLT 105 ribosomai KIVA gene, partiat sequence
	Think and a Francis And Andrew Provide the State of
	Brevibacillus borstelensis strain IPH801 16S ribosomal RNA gene complete sequence
	Previbacillus borstelensis strain NRC52 16S ribosomal RNA gene, nartial sequence
	Brevibacillus borstelensis strain 56 16S ribosomal RNA gene, partial sequence
	° ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ _ ↓ _ ↓ _ ↓ _ ↓
	Brevibacillus borstelensis strain cifa chp34 I6S ribosomal RNA gene, partial sequence
	firmicutes 2 leaves
	firmicutes 4 leaves
	□ Brevibacillus sp. AK-P2 16S ribosomal RNA gene, partial sequence
	Bacterium India-M' I6S ribosomal RNA gene, partial sequence
	Brevibacillus sp. D4 16S ribosomal RNA gene, partial sequence
	firmicutes 4 leaves
	Brevibacillus borstelensis strain RC 16S ribosomal RNA gene, partial sequence
	firmicutes 3 leaves
	Turmicules 5 leaves
0.003	y Brevidaciilus borstelensis strain ZYJ-51 165 ribosomal KNA gene, partial sequence
	y Acunobacterium Nino Tos ribosomai KNA gene, partial sequence
	Furnicules 4 leaves
	Infinicules 29 leaves

Staphylococcus epidermidis KF575163.1

 $\label{eq:staphylococcus} Staphylococcus epidermidis strain G0242 \ 16S \ ribosomal \ RNA \ gene, \ partial \ sequence \ Sequence \ ID: \ \underline{gb}[KF575163.1] \ \ Length: \ 1434 \ \ Number \ of \ Matches: \ 1$

Range 1:	44 to 101	6 GenBank Graphics		V Ne	ext Match 🔺 Previous Match	
Score 1718 bit	s(1904)	Expect 0.0	Identities 967/973(99%)	Gaps 3/973(0%)	Strand Plus/Plus	
Query	1	TTGACGTTAGCG	GCGGACGGGTGAG	TAACACGTGGGTAA	CCTACCTATAAGACTGGGAT	A 60
Sbjct	44	TTGACGTTAGCG	GCGGACGGGTGAG	TAACACGTGGGTAA	CCTACCTATAAGACTGGGAT	A 103
Query	61	ACTTCGGGAAAC	CGGAGCTAATACC	GGATAATATTTCGA	ACCGCATGGTTCGATAGTGA	A 120
Sbjct	104	ACTTCGGGAAAC	CGGAGCTAATACC	GGATAATATTTCGA	ACCGCATGGTTCGATAGTGA	A 163
Query	121	AGATGGTTTTGC	TATCACTTATAGA	TGGACCCGCGCCGT	ATTAGCTAGTTGGTAAGGTAA	A 180
Sbjct	164	AGATGGTTTTGC	IIIIIIIIIIIIII TATCACTTATAGA	TGGACCCGCGCCGT	ATTAGCTAGTTGGTAAGGTA	A 223
Query	181	CGGCTTACCAAG	GCGACGATACGTA	GCCGACCTGAGAGG	GTGATCGGCCACACTGGAACT	240
Sbjct	224	CGGCTTACCAAG	GCGACGATACGTA	GCCGACCTGAGAGG	GTGATCGGCCACACTGGAACT	c 283
Query	241	GAGACACGGTCC.	AGACTCCTACGGG	AGGCAGCAGTAGGG	AATCTTCCGCAATGGGCGAA	A 300
Sbjct	284	GAGACACGGTCC.	AGACTCCTACGGG	AGGCAGCAGTAGGG	AATCTTCCGCAATGGGCGAA	A 343
Query	301	GCCTGACGGAGC	AACGCCGCGTGAG	TGATGAAGGTCTTC	GGATCGTAAAACTCTGTTAT	360
Sbjct	344	GCCTGACGGAGC.	AACGCCGCGTGAG	TGATGAAGGTCTTC	GGATCGTAAAACTCTGTTAT	c 403
Query	361	AGGGAAGAACAT.	ACGTGTAAGTAAC	TATGCACGTCTTGA	CGGTACCTAATCAGAAAGCCA	A 420
Sbjct	404	AGGGAAGAACAT.	ACGTGTAAGTAAC	TATGCACGTCTTGA	CGGTACCTAATCAGAAAGCCA	A 463
Query	421	CGGCTAACTACG	TGCCAGCAGCCGC	GGTAATACGTAGGT	GGCAAGCGTTAT-CCGGAAT	r 479
Sbjct	464	CGGCTAACTACG	TGCCAGCAGCCGC	GGTAATACGTAGGT	GGCAAGCGTTATCCCGGAAT	523 c
Query	480	ATTGGGCGTAAA	GCGCGCGTAGGCG	GTTTTTTAAGTCTG	ATGTGAAAGCCCACGGCTCAA	A 539
Sbjct	524	ATTGGGCGTAAA	GCGCGCGTAGGCG	GTTTTTTAAGTCTG	ATGTGAAAGCCCACGGCTCAA	A 583
Query	540		ATTGGAAACTGGA	AAACTTGAGTGCAG	AAGAGGAAAGTGGAATTCCAI	599
Sbjct	584	CCGTGGAGGGTC	ATTGGAAACTGGA	AAACTTGAGTGCAG	AAGAGGAAAGTGGAATTCCAT	643
Query	600	GTGTAGCGGTGA	AATGCGCAGAGAT	ATGGAGGAACACCA	GTGGCGAAGGCGACTTTCTGG	659
Sbjct	644	GTGTAGCGGTGA	AATGCGCAGAGAT	ATGGAGGAACACCA	GTGGCGAAGGCGACTTTCTGG	; 703
Query	660	TCTGTAACTGAC	GCTGATGTGCGAA	AGCGTGGGGGATCAA	ACAGGATTAGATACCCTGGTA	719
Sbjet	704	TCTGTAACTGAC	GCTGATGTGCGAA	AGCGTGGGGGATCAA		763
Query	720					, 022
Ouerv	780	GTEEACGCEGTA	CACTCCCCCTCC	GAGTACGACCGCA	AGGTTGAAACTCAAAGGAATT	, 839
Sbict	824	CTAACGCATTAA	JCACTCCGCCTGG	GAGTACGACCGCA	AGGTTGAAACTCAAAGGAATT	883
Query	840	GACGGGGGACCCG	CACAAGCGGTGGA	GCATGTGGTTTAAT	TCGAAGC-ACGCGAAAAACCI	898
Sbjct	884	GACGGGGGACCCG	CACAAGCGGTGGA	 GCATGTGGTTTAAT	IIIIII IIIII IIII TCGAAGCAACGCGAAGAACCI	943
Query	899	TACCAAATCTTG	ACATCCTTTGACA	ACTCTAAAGATAGA	GCCTTCCCCTTTCGGGGGGA-A	957
Sbjct	944	IIIIIIIIIII TACCAAATCTTG	ACATCCTTTGACA	ACTCTAGAGATAGA		1003
Query	958	AAGTGACAGGGG	G 970			
Sbjct	1004	AAGTGACAGGTG	G 1016			

Cl/Query_218737	
	Uncultured bacterium clone nck172b01c1 16S ribosomal RNA gene, partial sequence Stanbulococcus enidermidis strain G0242 16S ribosomal RNA gene, partial sequence
Ī	Staphylococcus consernations strain G0242 105 ribosoniai River gene, partial sequence
	Staphylococcus sp. BQN2P-01d 16S ribosomal RNA gene, partial sequence
	^o Uncultured bacterium clone nbu277a09c1 16S ribosomal RNA gene, partial sequence al locultured bacterium clone nbu282a12a1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbu283c12c1 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw830e11c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw831h11c1 16S ribosomal RNA gene, partial sequence
	Y Uncutured bacterium clone nbw1189e10c1 16S ribosomal RNA gene, partial sequence 9 Uncultured bacterium clone nbw1095d07c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw1105h02c1 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw1106f08c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw1106h08c1 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone nbw1107b10c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone now 1108/04c1 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw1065c02c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw118900c1 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone nbw1191b10c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nbw1190f12c1 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain LCR51 16S ribosomal RNA gene, partial sequence Stanhylococcus haemolyticus strain EOU2 16S ribosomal RNA gene, partial sequence
	Staphylococcus naemolyticus strain EQ12 105 ribosomal KNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain FR1 68 16S ribosomal RNA gene, partial sequence Staphylococcus strain FR1 68 16S riboso
	Vincultured bacterium clone ncd558b09c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncd577e07c1 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone ncd57611c1 16S ribosomal RNA
	Uncultured bacterium clone ncd93001101 105 nbosomal RNA gene, partial sequence Uncultured bacterium clone ncd983f02c1 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. YSL09-4 gene for 16S rRNA, partial sequence
	Uncultured bacterium clone ncd924b12c1 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone ncd924b12c1 16S ribosomal RNA
	Staphylococcus sp. HL-13 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain CIFRI D-TSB2 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain CIFRI P-TSB4 16S ribosomal RNA gene, partial sequence Staphylococcus for CIFRI CH TSB23 16S ribosomal RNA gene partial sequence
	Staphylococcus sp. CIFRI H-TSB-15-ZMA 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. CIFRI H-TSB-20-ZMA 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. CIFRI H-TSB-6-HA 16S ribosomal RNA gene, partial sequence Staphylococcus baemolyticus strain HNMCTP1 16S ribosomal RNA gene partial sequence
	Staphylococcus haemolyticus strain HNOCTEN TOS HOSOniai NAA gene, partial sequence Staphylococcus haemolyticus strain BP3_2B 16S ribosonial RNA gene, partial sequence
	Staphylococcus haemolyticus strain LEH2_2A 16S ribosomal RNA gene, partial sequence
	Y Staphylococcus sp. EB338 16S ribosomal RNA gene, partial sequence 9 Uncultured Staphylococcus sp. clone TJ-3 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. ARB1 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain 20.1 16S ribosomal RNA gene, partial sequence Staphylococcus epidermidis strain 21.1.16S ribosomal RNA gene, partial sequence
	Acinetobacter radioresistens strain 21.1 105 ribosomai RNA gene, partial sequence
	Staphylococcus haemolyticus strain G35 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. RH-29 gene for 16S rRNA, partial sequence Staphylococcus sp. M17 16S ribosomal RNA gene partial sequence
	Staphylococcus haemolyticus JCSC1435 strain JCSC1435 16S ribosomal RNA, complete sequence
	Staphylococcus haemolyticus strain M2 16S ribosomal RNA gene, partial sequence Restarium Culamore 10E 16S ribosomal RNA gene partial sequence
	Staphylococcus haemolyticus strain S001N 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain S006N 16S ribosomal RNA gene, partial sequence
	Y Staphylococcus haemolyticus strain S009b 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain S011b 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain VBW023 16S ribosomal RNA gene, partial sequence
	Bacterium KR 1996/1_182 16S ribosomal RNA gene, partial sequence
	v staphylococcus naemolyticus strain L58 165 ribosomal KNA gene, partial sequence 9 Staphylococcus haemolyticus strain BP/SU2 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. H-179 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain SH6 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain SH8 16S ribosomal RNA gene, partial sequence
	v staphylococcus naemolyticus strain SH8 105 ribosomal KNA gene, partial sequence Staphylococcus haemolyticus strain NIOT-sb3 165 ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain C0181 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. G0271 16S ribosomal RNA gene, partial sequence
	A suphyboloccus naemolyticus strain PAI-5 105 fibosomal RNA gene, partial sequence Uncultured bacterium clone g3b23 16S fibosomal RNA gene, partial sequence
	Uncultured bacterium clone g3b26 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone g3b27 16S ribosomal RNA gene, partial sequence Locultured bacterium clone g3b33 16S ribosomal RNA gene, partial acquires
	Uncultured bacterium clone g3b39 165 ribosomal RNA gene, partial sequence Uncultured bacterium clone g3b39 165 ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain XB20 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain MJMG7.10 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain K11-5-97 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain KJ1599 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain KJ1599 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck176a12c1 16S ribosomal RNA gene, partial sequence
	⁹ Uncultured bacterium clone nck170f04c1 16S ribosomal RNA gene, partial sequence 9 Uncultured bacterium clone nck171d06c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncm50e05c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncm50g10c1 16S ribosomal RNA gene, partial sequence
	Oncultured bacterium clone nck169d07c1 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone nck171e08c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck175c09c1 165 ribosonal RNA gene, partial sequence
	Our ultured bacterium clone nck177d11c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck188a05c1 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone nck310c02c1 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain DG-4 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain CIFRI P-TSB-72 16S ribosomal RNA gene, partial sequence Staphylococcus haemolyticus strain U2 16S ribosomal RNA gene, partial sequence
	Staphylococcus naemolyticus strain J12 165 ribosomal KNA gene, partial sequence Staphylococcus sp. 09-A3 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. 09-M3 16S ribosomal RNA gene, partial sequence
	Staphylococcus haemolyticus strain DSO4G14 16S ribosomal RNA gene, partial sequence Staphylococcus m. Ao08 16S ribosomal RNA gene, partial sequence
	Bacterium 7RO1 16S ribosomal RNA gene, partial sequence
	firmicutes 4 leaves
	Staphylococcus haemolyticus strain DSO6G11 16S ribosomal RNA gene, partial sequence

Pantoea calida gene for 16S ribosomal RNA, partial sequence, strain: LMG 25383 Sequence ID: <u>dbj[AB907785.1]</u> Length: 1495 Number of Matches: 1

Range 1: 51 to 1032 GenBank Graphics V Next Match 🔺 Previous Match							
Score 1745 bit	s(1934)	Expect 0.0	Identities 977/982(99%)	Gaps 1/982(0%	Strand) Plus/Plus		
Query	1	AGCAGCTTGCT	CTTCGCTGAC	SAGTGGCGGACGGG	TGAGTAATGTCTGGGGATCTG	CCC 60	
Sbjct	51	AGCAGCTTGCTG	GCTTCGCTGAC	GAGTGGCGGACGGG	TGAGTAATGTCTGGGGATCTG	CCC 110	
Query	61	GATGGAGGGGG	TAACCACTGG	AACGGTGGCTAAT	ACCGCATAACGTCGCAAGACC	AAA 120	
Sbjct	111	GATGGAGGGGG	TAACCACTGG	AACGGTGGCTAAT	ACCGCATAACGTCGCAAGACC	AAA 170	
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Sbjct	171	GTGGGGGACCTT	CGGGCCTCAC	ACCATCGGATGAAC	CCAGATGGGATTAGCTAGTAG	GTG 230	
Query	181	GGGTAACGGCTC	ACCTAGGCGAG	CGATCCCTAGCTGG	TCTGAGAGGATGACCAGCCAC	ACT 240	
Sbjct	231	GGGTAACGGCTC	ACCTAGGCGAC	CGATCCCTAGCTGG	TCTGAGAGGATGACCAGCCAC	ACT 290	
Query	241	GGAACTGAGACA	CGGTCCAGACT		AGCAGTGGGGGAATATTGCACAA	TGG 300	
Sbjct	291	GGAACTGAGACA	ACGGTCCAGACI	TCCTACGGGAGGCA	GCAGTGGGGGAATATTGCACAA'	TGG 350	
Query	301	GCGCAAGCCTGA	TGCAGCCATG	CCGCGTGTATGAAG		ACT 360	
Sbjct	351	GCGCAAGCCTGA	TGCAGCCATGO	CCGCGTGTATGAAG	AAGGCCTTCGGGTTGTAAAGT	ACT 410	
Query	361	TTCAGCGGGGGAG	GAAGGGATGG	TGCTTAATACGCGC	CGTCATTGACGTTACCCGCAG	AAG 420	
Sbjct	411	TTCAGCGGGGGAG	GAAGGGATGG	TGCTTAATACGCGC	CGTCATTGACGTTACCCGCAG	AAG 470	
Query	421	AAGCACCGGCTA	ACTCCGTGCCZ	AGCAGCCGCGGTAA	TACGGAGGGTGCAAGCGTTAA'	TCG 480	
Sbjct	471	AAGCACCGGCTA	ACTCCGTGCCZ	AGCAGCCGCGGTAA	TACGGAGGGTGCAAGCGTTAA	TCG 530	
Query	481	GAATTACTGGGC	GTAAAGCGCAC	CGCAGGCGGTCTGT	TAAGTCAGATGTGAAATCCCC	GGG <mark>54</mark> 0	
Sbjct	531	GAATTACTGGGC	GTAAAGCGCAC	CGCAGGCGGTCTGT		GGG 590	
Query	541						
Sbjet	591				GAGTCTCGTAGAGGGGGGGGAG	AAT 650	
Query	601 651						
ouerr	661	CCECCAGGIGIAGO	GUIGAAAIGUG	CECCARAC	CCC2CC22CC222CC2CC2	CCC 710	
Query	711						
Suger	711		CIGACGCICAG			mcc 700	
Query	721						
Suger	701	GGIAGICCACG		GICGACIIGGAGG		1CC 830	
Query	/01 021					ATG 840	
Sbjet	0.01		TTAAGTCGACC			ATG 890	
Query	041					AGA 900	
Suger	0.01	AATTGACGGGGG		GGIGGAGCAIGIG	A A B B C C C C B C C C C C C C C C C C	AGA 950	
Spict	901					AAC 960	
ouerr	951	CCTCADADACC	CITGACATCCA	001	MOATGUULIAGTGUUTTUGGG	MAC IUIU	
guery	901 1011			901 1032			
ວມງບບ	TOTT	COLONGHCHOGI	OCTOCATOOC	1002			



Bacillus cereus DQ923480.1

Bacillus cereus strain D23 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|DQ923480.1|</u> Length: 1429 Number of Matches: 1

Range 1: 430 to 1390 GenBank Graphics 💎 Next Match 🔺 Previous Match							
Score 1725 bit	s(1912)	Expect 0.0	Identities 959/961(99%)	Gaps 0/961(0%)	Strand Plus/Minus		
Query	1	TGTTACAAACT	CTCGTGGTGTGACG	GGCGGTGTGTAC	AGGCCCGGGAACGTA	TTCACCG	60
Sbjct	1390	TGTTACAAACT	CTCGTGGTGTGACG	GGCGGTGTGTAC	AGGCCCGGGAACGTA	TTCACCG	1331
Query	61	CGGCATGCTGA	TCCGCGATTACTAG	CGATTCCAGCTTC	CATGTAGGCGAGTTGC.	AGCCTAC	120
Sbjct	1330	CGGCATGCTGA	TCCGCGATTACTAG	CGATTCCAGCTTC	CATGTAGGCGAGTTGC.	AGCCTAC	1271
Query	121	AATCCGAACTG	AGAACGGTTTTATG	AGATTAGCTCCA	CTCGCGGTCTTGCAG	CTCTTTG	180
Sbjct	1270	AATCCGAACTG	AGAACGGTTTTATG	AGATTAGCTCCAC	CTCGCGGTCTTGCAG	CTCTTTG	1211
Query	181	TACCGTCCATT	GTAGCACGTGTGTA	GCCCAGGTCATA	AGGGGCATGATGATTT	GACGTCA	240
Sbjct	1210	TACCGTCCATT	GTAGCACGTGTGTA	GCCCAGGTCATA	AGGGGCATGATGATT	GACGTCA	1151
Query	241	TCCCCACCTTC	CTCCGGTTTGTCAC	CGGCAGTCACCT	TAGAGTGCCCAACTTA	ATGATGG	300
Sbjct	1150	TCCCCACCTTC	CTCCGGTTTGTCAC	CGGCAGTCACCT	TAGAGTGCCCAACTTA	ATGATGG	1091
Query	301	CAACTAAGATC	AAGGGTTGCGCTCG	TTGCGGGACTTA	ACCCAACATCTCACGA	CACGAGC	360
Sbjct	1090	CAACTAAGATC	AAGGGTTGCGCTCG	TTGCGGGACTTA	ACCCAACATCTCACGA	CACGAGC	1031
Query	361	TGACGACAACC	ATGCACCACCTGTC	ACTCTGCTCCCG	AGGAGAAGCCCTATC	TCTAGGG	420
Sbjct	1030	TGACGACAACC	ATGCACCACCTGTC	ACTCTGCTCCCG	AGGAGAAGCCCTATC	TCTAGGG	971
Query	421	TTTTCAGAGGA	TGTCAAGACCTGGT	AAGGTTCTTCGCC	GTTGCTTCGAATTAAA	CCACATG	480
Sbjct	970	TTTTCAGAGGA	TGTCAAGACCTGGT	AAGGTTCTTCGCC	TTGCTTCGAATTAAA	CCACATG	911
Query	481	CTCCACCGCTT	GTGCGGGGCCCCCGT	CAATTCCTTTGAC	GTTTCAGCCTTGCGGC	CGTACTC	540
Sbjct	910	CTCCACCGCTT	GTGCGGGGCCCCCGT	CAATTCCTTTGAC	TTTCAGCCTTGCGGC	CGTACTC	851
Query	541	CCCAGGCGGAG	IGCTTAATGCGTTA	ACTTCAGCACTA		TAACACT	600
Sbjct	850	CCCAGGCGGAG	IGCTTAATGCGTTA	ACTTCAGCACTA	AAGGGCGGAAACCCTC	TAACACT	791
Query	601	TAGCACTCATC	GTTTACGGCGTGGA	CTACCAGGGTAT		CCCACGC	<mark>66</mark> 0
Sbjct	790	TAGCACTCATC	GTTTACGGCGTGGA	CTACCAGGGTAT	CTAATCCTGTTTGCTC	CCCACGC	731
Query	661	TTTCGCGCCTCA	AGTGTCAGTTACAG		CCTTCGCCACTGGTGI	TCCTCCA	720
Sbjct	730	TTTCGCGCCTC	AGTGTCAGTTACAG	ACCAGAAAGTCG	CCTTCGCCACTGGTGI	TCCTCCA	671
Query	721		ATTTCACCGCTACA	CATGGAATTCCA		TCAAGTC	780
Sbjct	670	TATCTCTACGC	ATTTCACCGCTACA	CATGGAATTCCA	CTTTCCTCTTCTGCAC	TCAAGTC	<mark>6</mark> 11
Query	781	TCCCAGTTTCCA	AATGACCCTCCACG	GTTGAGCCGTGG	GCTTTCACATCAAACI	TAAGAAA	840
Sbjct	610	TCCCAGTTTCCA	AATGACCCTCCACG	GTTGAGCCGTGG	GCTTTCACATCAGACI	TAAGAAA	551
Query	841	CCACCTGCGCG	CGCTTTACGCCCAA	TAATTTCCGGAT		GTATTAC	900
Sbjct	550	CCACCTGCGCG	CGCTTTACGCCCAA	TAATTTCCGGAT	AACGCTTGCCACCTAC	GTATTAC	491
Query	901	CGCGGCTGCTG	GCACGTAGTTAGCC	GTGGCTTTCTGG		GGCCAGC	960
Sbjct	490	CGCGGCTGCTG	GCACGTAGTTAGCC	GTGGCTTTCTGG	TTAGGTACCGTCAAGG	TGCCAGC	431
Query	961	т 961 I					
Sbjct	430	т 430					

Icl Query_36213	
•	Uncultured bacterium clone 5_Ne_23 16S ribosomal RNA gene, partial sequence
¢	Bacillus cereus strain D23 16S ribosomal RNA gene, partial sequence
	Bacterium 8-gw1-9 16S ribosomal RNA gene, partial sequence
	firmicutes 10 leaves
	firmicutes 7 leaves
	Bacillus cereus strain g54 16S ribosomal RNA gene, partial sequence
	Bacillus toyonensis strain R18 16S ribosomal RNA gene, partial sequence
	Bacillus thuringiensis strain PWI-A4 16S ribosomal RNA gene, partial sequence
	Bacterium NR14 16S ribosomal RNA gene, partial sequence
	Bacterium NF8 16S ribosomal RNA gene, partial sequence
	Bacterium NF6 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain IBS2 16S ribosomal RNA gene, partial sequence
	Bacillus thuringiensis strain GG31 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain C-4 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain DER8 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain sm-sr14 16S ribosomal RNA gene, partial sequence
	Bacillus sp. LLH-Irr-28 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain MAL_3A 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain MAL_5A 16S ribosomal RNA gene, partial sequence
	Bacillus thuringiensis strain M4 16S ribosomal RNA gene, partial sequence
	Bacillus sp. INBio_4039AD 16S ribosomal RNA gene, partial sequence
	Bacillus sp. INBio_4042AS I6S ribosomal RNA gene, partial sequence
	Bacillus sp. INBio_4052G 16S ribosomal RNA gene, partial sequence
	Bacillus sp. INBio_4062F 16S ribosomal RNA gene, partial sequence
	Bacillus sp. INBio_4312S 16S ribosomal RNA gene, partial sequence
	Bacillus sp. INBio_4516D 16S ribosomal RNA gene, partial sequence
	Bacillus thuringiensis strain LAMA 1099 16S ribosomal RNA gene, partial sequence
	Bacillus sp. 1-4 I6S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5_Am_24 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5_Am_34 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 Am_39 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 Am 45 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5_Am_60 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5_Am_75 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 Am 81 165 ribosomal RNA gene, partial sequence
	Unclitured bacterium clone 5 K 55 ToS hostomat RNA gene, partial sequence
	Uncultured bacterium clone 5 Ne 28 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 Ne 31 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 No 54 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 No 50 166 biosomal RNA gene, partial sequence
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	Uncultured bacterium clone 5 St 15 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 St 15 105 filosomal RNA gene, partial sequence
	Uncultured bacterium clone 5 St 24 16S ribosomal RNA gene, partial sequence
0.0003	Uncultured bacterium clone 5 St 41 16S ribosomal RNA gene, partial sequence
	Tene anarea bacterium clone 5 51 41 105 nosonal INA gene, partai sequence
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	Tunne dies 50 reaves

Pseudomonas luteola KC429633.1

$\label{eq:sequence} \begin{array}{l} \mbox{Pseudomonas luteola strain XFB-BV 16S ribosomal RNA gene, partial sequence sequence ID: $\underline{gb}[KC429633.1]$ Length: 1388 Number of Matches: 1 \\ \end{array}$

Range 1:	42 to 99	1 <u>GenBank</u>	<u>Graphics</u>				V Next N	Match 🔺 P	revious Ma	atch	
Score 1669 bit	s(1850)		Expect 0.0	Identities 940/950(99%)	Gaps 0/950(0	%)	Strand Plus/Plu	6		
Query	1	GGCGGZ	ACGGGTGA	GTAATGCCTA	GGAATC	GCCTGG	TAGTGG	GGGACA	ACGTTI	CGAAAG	60
Sbjct	42	GGCGGZ	ACGGGTGA	IIIIIIIII GTAATGCCTA	 GGAATCI	GCCTGG	TAGTGG	GGGACA	ACGTTI	 CGAAAG	101
Query	61	GAACGO	CTAATACC	GCATACGTCC	TACGGG	GAAAGI	GGGGGA	TCTTCG	GACCTO	ACGCTA	120
Sbjct	102	GAACGO	CTAATACC	GCATACGTCC	TACGGG	IIIIII	GGGGGGA	TCTTCG	GACCTO	ACGCTA	161
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Sbjct	162	TCAGA	IGAGCCTA	GGTCGGATTA	GCTAGT	GGTGGG	GTAACG	GCTCAC	CAAGGO	GACGAT	221
Query	181	CCGTA	ACTGGTCT	GAGAGGATGA			AACTGA	GACACG	GTCCAG	ACTCCT	240
Sbjct	222	CCGTA	ACTGGTCT	GAGAGGATGA	TCAGTC	ACACTGO	AACTGA	GACACG	GTCCAG	ACTCCT	281
Query	241	ACGGG	AGGCAGCA	GTGGGGAATA	TTGGAC	ATGGGC	GAAAGC	CTGATC		TGCCGC	300
Sbjct	282	ACGGG	AGGCAGCA	GTGGGGAATA	TTGGAC	ATGGGC	GAAAGC	CTGATC	CAGCCA	TGCCGC	341
Query	301	GTGTGT	rgaagaag	GCCCTCGGGT	CGTAAA	GACTTI	AAGCTG	GGAGGA	AGGGTI	GTAACC	360
Sbjct	342	GTGTG	rgaagaag	GCCCTCGGGT	CGTAAA	CACTTI	AAGCTG	GGAGGA	AGGGTI	GTAACC	401
Query	361		CGTTGCAG	CTTTGACGTT		GAATAA	GCACCG	GCTAAC	TCTGTG	CCAGCA	420
Sbjct	402	TAATAC	CGTTGCAG	CTTTGACGTT	ACCAGC	GAATAA	GCACCG	GCTAAC	TCTGTG	CCAGCA	461
Query	421	GCCGCC	GTAATAC	AGAGGGTGCA		ATCGGA	ATTACT	GGGCGT	AAAGCG	CGCGTA	480
Sbjct	462	GCCGCC	GTAATAC	AGAGGGTGCA	AGCGTT	ATCGGA	ATTACT	GGGCGT	AAAGCO	CGCGTA	521
Query	481	GGTGGC	CTTGGTAA	GTTGAATGTG			CAACCT	GGGAAC	TGCATC	CAAAAC	540
Sbjct	522	GGTGGC	CTTGGTAA	GTTGAATGTG	AAATCCO	CGGGCI	CAACCT	GGGAAC	TGCATC	CAAAAC	581
Query	541	TGCCTO	GCTAGAG	TACGGTAGAG	GGTGGT(GAATTI	CCTGTG	TAGCGG	TGAAA1	GCGTAG	600
Sbjct	582	TGCCTO	GCTAGAG	TACGGTAGAG	GGTGGT	GAATTI	CCTGTG	TAGCGG	TGAAAI	GCGTAG	641
Query	601		GAAGGAA	CACCAGTGGC	GAAGGC	SACCACC	TGGACT	GATACT	GACACI	GAGGTG	660
Sbjct	642	ATATAC	GAAGGAA	CACCAGTGGC	GAAGGC	SACCACO	TGGACT	GATACT	GACACI	GAGGTG	701
Query	661		GCGTGGGG	AGCAAACAGG	ATTAGA:		GTAGTC	CACGCC	GTAAAC	GATGTC	720
Sbjct	702	CGAAAG	CGTGGGG	AGCAAACAGG	ATTAGA	TACCCTO	GTAGTC	CACGCC	GTAAAO	GATGTC	761
Query	721	AACTAG	GCCGTTGG	GGTCCTTGAG		GTGGCGC	AGCTAA	CGCAAT	AAGTTO	ACCGCC	780
Sbjct	762	AACTAC	SCCGTTGG	GGTCCTTGAG	ACTTTA	STGGCGC	AGCTAA	CGCAAT	AAGTTO	ACCGCC	821
Query	781	TGGGGZ	AGTACGGC	CGCAAGGTTA			TTGACG	GGGGCC		AGCGGT	840
Sbjct	822	TGGGGZ	AGTACGGC	CGCAAGGTTA	AAACTC	AATGAA	TTGACG	GGGGCC	CGCACA	AGCGGT	881
Query	841	GGAGCZ	ATGTGGTT	TATTTCGAAG			CTTACC	AGGCCT	TGACAI	GCAAAA	900
Sbjct	882	GGAGCZ	ATGTGGTT	TAATTCGAAG	CAACGC	GAAGAAC	CTTACC	AGGCCT	TGACAI	GCAGAG	941
Query	901	AACTTT		AGGATTGGTG		GAACTO	TGAACC		TG 95	50	
Sbjct	942	AACTTI	CCAGAGA	TGGATTGGTG	CCTTCG	GAACTO	TGACAC	AGGTGC	TG 99	91	

bacteria | 2 leaves

4- The phylogenetic analysis of bacteria isolated from mobile phones samples

Staphylococcus epidermidis KJ398217.1

Staphylococcus epidermidis strain LH-Y4 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|KJ398217.1</u>| Length: 1413 Number of Matches: 1

Range 1: 590 to 1369 GenBank Graphics					🔻 Next Match 🔺 Previous Match				
Score	(1550)	Expect Identities		Gaps	Strand				
1398 bit	ts(1550)	0.0	778/780(99%)	0/780(0%)	Plus/Minus				
Query	3	CTCTCGTGGT	GTGACGGGCGGTGTG	TACAAGACCCGGG	AACGTATTCACCGTAGCATGCI	62			
Sbjct	1369	CTCTCGTGGT	GTGACGGGCGGTGTG	TACAAGACCCGGG	AACGTATTCACCGTAGCATGCT	1310			
Query	63	GATCTACGAT		CTTCATATAGTCG	AGTTGCAGACTACAATCCGAAC	: 122			
Sbjct	1309	GATCTACGAT	TACTAGCGATTCCAG	CTTCATATAGTCG	AGTTGCAGACTACAATCCGAAC	: 1250			
Query	123	TGAGAACAAC	TTTATGGGATTTGCT	TGACCTCGCGGTT	TCGCTACCCTTTGTATTGTCCA	182			
Sbjct	1249	TGAGAACAAC	TTTATGGGATTTGCT	TGACCTCGCGGTT	TCGCTGCCCTTTGTATTGTCCA	1190			
Query	183	TTGTAGCACG	TGTGTAGCCCAAATC	ATAAGGGGCATGA	TGATTTGACGTCATCCCCACCI	242			
Sbjct	1189	TTGTAGCACG	TGTGTAGCCCAAATC	ATAAGGGGCATGA	TGATTTGACGTCATCCCCACCI	1130			
Query	243	TCCTCCGGTT	TGTCACCGGCAGTCA	ACTTAGAGTGCCC	AACTTAATGATGGCAACTAAGC	302			
Sbjct	1129	TCCTCCGGTT	TGTCACCGGCAGTCA	ACTTAGAGTGCCC	AACTTAATGATGGCAACTAAGC	: 1070			
Query	303	TTAAGGGTTG	CGCTCGTTGCGGGAC	TTAACCCAACATC	TCACGACACGAGCTGACGACAA	362			
Sbjct	1069	TTAAGGGTTG	CGCTCGTTGCGGGAC	TTAACCCAACATC	TCACGACACGAGCTGACGACAA	1010			
Query	363	CCATGCACCA	CCTGTCACTCTGTCC	CCCGAAGGGGAAA	ACTCTATCTCTAGAGGGGTCAG	, 422			
Sbjct	1009	CCATGCACCA	CCTGTCACTCTGTCC	CCCGAAGGGGAAA	ACTCTATCTCTAGAGGGGGTCAG	950			
Query	423	AGGATGTCAA	GATTTGGTAAGGTTC	TTCGCGTTGCTTC	GAATTAAACCACATGCTCCACC	482			
Sbjct	949	AGGATGTCAA	GATTTGGTAAGGTTC	TTCGCGTTGCTTC	GAATTAAACCACATGCTCCACC	890			
Query	483	GCTTGTGCGG	GTCCCCGTCAATTCC	TTTGAGTTTCAAC	CTTGCGGTCGTACTCCCCAGGC	542			
Sbjct	889	GCTTGTGCGG	GTCCCCGTCAATTCC	TTTGAGTTTCAAC	CTTGCGGTCGTACTCCCCAGGC	830			
Query	543	GGAGTGCTTA	ATGCGTTAGCTGCAG	CACTAAGGGGCGG	AAACCCCCTAACACTTAGCACT	602			
Sbjct	829	GGAGTGCTTA	ATGCGTTAGCTGCAG	CACTAAGGGGCGG	AAACCCCCTAACACTTAGCACT	770			
Query	603	CATCGTTTAC	GGCGTGGACTACCAG	GGTATCTAATCCT	GTTTGATCCCCACGCTTTCGCA	662			
Sbjct	769	CATCGTTTAC	GGCGTGGACTACCAG	GGTATCTAATCCT	GTTTGATCCCCACGCTTTCGCA	710			
Query	663	CATCAGCGTC	AGTTACAGACCAGAA	AGTCGCCTTCGCC	ACTGGTGTTCCTCCATATCTCT	722			
Sbjct	709	CATCAGCGTC	AGTTACAGACCAGAA	AGTCGCCTTCGCC	ACTGGTGTTCCTCCATATCTCT	650			
Query	723	GCGCATTTCA	CCGCTACACATGGAA	ATTCCACTTTCGT	CTTCTGCACTCAAGTTTTCCAG	; 782			
Sbjct	649	GCGCATTTCA	CCGCTACACATGGAA	ATTCCACTTTCCT	CTTCTGCACTCAAGTTTTCCAG	; 590			

	21d[Query 53347
	Bacterium N47 16S ribosomal RNA gene, nartial sequence
2	Bacterium N47 165 ribosonal RNA gene, partial sequence
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	Staphylococcus epidermidis strain Y5 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain JDM2 4A 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain LEH7 1A 16S ribosomal RNA gene, partial sequence
	 Staphylococcus epidermidis strain ECNU-UE2 16S ribosomal RNA gene, partial sequence
	Staphylococcus endermidi strain ECNU-UII 16S ribosomal RNA gene, partial sequence
	Uncultured oreans plant in the FL 10023-T100-S-NIPCR AMeANb 000153 small subunit thosomal RNA gene partial sequence
	Uncultured organism clone EL 10072-T477-SNIPCRAM eAva 000053 small subunit ribosomal RNA eene nartial sequence
	Uncultured organism clone EL 10072-T477-S-NIPCRAMeANa 000167 small subunit ribosomal RNA eene, partial sequence
	Uncultured organism clone EL 10072-1477.5 NI PCP AM ANa 000384 small submit ribosomal RNA gene partial sequence
	Uncultured organism clone EL10072-1477-5441 CRABGANA Na 0000045 small subunit fitosomal RAA gane, partial sequence
	Uncultured organism clone EE 0012-1477-3-544 CRAINGALM 000009 small subunit ribosonal RAM gene partial sequence
	Uncultured organism clone LE U0157-1387-5 VIII CRAMBANK 000108 cmall subunit ribosomal RVA gane, partial sequence
	Uncultured organism clone ELECOTO / 150/-5/WFCRAMgANG_000/05 mill subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone ELCO107-1415-5-WFCRAMgRAa 000502 small subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone ELCO105-1415-5-WFCRAMgrAnd 200517 small subunit ribosomat RVA gene, partial sequence
	Uncounted organism clone EL 20107/14155/NIP CRAWGA1a 000054 small solunit hosonial RVA gene, partial sequence Uncounted organism clone EL 20107/1478 CMDC AMAANa 000054 small solunit hosonial RVA gene, partial sequence
	Uncultured organism clone ELCO176-1478-5-NIPCRAMIGATA 000004 small suburnt ribosomat RNA gene, partai sequence
	Checultured organism clone EL 00026-1115-S-NIPC RAMBANA 000049 small subunit ribosomal RNA gene, partial sequence
	 Unculniced organism clone EL 000/20-1115-5-NIFC RAMGANA 000025 small subunit inDosomal RNA gene, partial sequence Unculniced organism clone EL 1000/2 TLIS S NIFC RAMA AND 000085 small subunit in base and DNA second participation of the second participation
	T Unculnured organism clone EL UW26-1113-3-WIFCRAMGANA UW283 Small subunit inDosomal KNA gene, partial sequence
	 Uncohunced organism clone ELCOW20+1115-5-NIFCKAMIGANA_UOV101 small subunit inDosomal KNA gene, partial sequence Uncohunced organism clone ELV002(TLLS C) NIFCKAMIGANA_UOV101 small subunit inDosomal KNA gene, partial sequence
	7 Uncultured organism clone EL 00026-1115-S-NIPCRAMgANa 00037/8 small subunit ribosomal RNA gene, partial sequence Uncultured organism clone EL 00026-1115-6 NIPCRAMGANA 00037/8 small subunit ribosomal RNA gene, partial sequence
	y Uncultured organism cione EL U0026-1115-S-NIPCRAMGANa 000491 small subunit ribosomal RNA gene, partial sequence Uncultured organism cione EL U0026-1125-S-NIPCRAMGANA 0006923
	y Uncultured organism cione EL UM28-11/2-S-NIPCRAMgANa UM2053 small subunit ribosomal KNA gene, partial sequence Uncultured organism cione EL UM27 ELSO STATES AND
	 Uncultured organism clone ELU0024-1150-S-NIPC KAMgAND 000297 small subunit ribosomal RNA gene, partial sequence Uncultured organism clone EL 00024 TEGE S NIPC RAMGANE 000470 and the basic cloned by the second secon
	Cheffer and State and Stat
	⁴ Uncultured organism clone EL 00052-T156-S-NIPCRAMgANb 000548 small subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone EL 00032-T156-S-NIPCRAMgANb 000554 small subunit ribosomal RNA gene, partial sequence
	Checultured organism clone EL 00032-1156-5-NIPCRAMgAND 000565 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL 00052-1156-5-NIPCRAMgANb 000564 small subunit ribosomal RNA gene, partial sequence
	⁴ Uncultured organism clone EL 00038-T189-S-NPCRAMgANb 000076 small subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone EL 00038-T189-S-NPCRAMgANb 000155 small subunit ribosomal RNA gene, partial sequence
	⁹ Uncultured organism clone ELU0038-T189-S-NPCRAMgANb (000193 small subunit ribosomal RNA gene, partial sequence Uncultured organism clone ELU0038-T189-S-NPCRAMgANb (000193 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL 00038-T189-S-NPCRAMgANb 00021/ small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL 00038-1189-5-NPCRAMgANb 000397 small subunit ribosomal RNA gene, partial sequence
	² Uncultured organism clone ELU0038-T189-S-NPCRAMgANb (000425 small subunit ribosomal RNA gene, partial sequence Uncultured organism clone ELU0038-T189-S-NPCRAMgANb (000425 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL 00038-T189-S-NPCRAMgANb_000467 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL 00038-T189-S-NPCRAMgANb 000482 small subunit ribosomal RNA gene, partial sequence
	Checultured organism clone EL 00038-1189-5-NPCRAMgAND 000549 small subunit ribosomal RNA gene, partial sequence Unclutured organism clone EL 00038-1189-5-NPCRAMgAND 000549 small subunit ribosomal RNA gene, partial sequence
	Chelinared organism cione EL 00041-1385-5-NIPCRAMgAND 000188 small subunit ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis gene for ISS ribosomal RNA, partial sequence, isolate: B0619
	Staphylococcus epidermidis strain S9-627 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain M-S-MRS 6 165 ribosmal RNA gene, partial sequence
	Bacterium NLAE-zI-G4/0 16S ribosomai RNA gene, partial sequence
	Staphylococcus epidermidis strain XJFH-J-1 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis partial 16S rRNA gene, isolate 0609ALT19Q2-GH
	Starburyhococcus epiderminis strain HKG 165 165 ribosomai RNA gene, partial sequence
	Stanbulgeocome anidermidie stanie IMC 182 165 sibosomal RNA gene, partial sequence Stanbulgeocome anidermidie stanie IMC 182 165 sibosomal RNA come natial esquence
	y staphyrococcus epidermikits strain FIKU 162 105 mossoniai KINA gene, partial sequence 9 staphyrococcus epidermikits strain HKU 183 165 ribosonial PNA gane pagial sequence
	Bactering L-20 (BS ribosonal RNA one narrial source
	Uncultured Staphylococcus sp, clone C139100025 165 ribosomal RNA eene, partial sequence
	Staphylococcus sp. W2.10-181 16S ribosomal RNA (16S rRNA) gene, complete sequence
	Staphylococcus epidermidis strain 72 (br3) 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain ATHA24 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. DF7n-C small subunit ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis partial 16S rRNA gene, isolate OCAT7
	Staphylococcus epidermidis partial 16S rRNA gene, isolate OCAT8
	Starbuy lococcus sp. OCA 110 partial 105 rKiNA gene, isolate OCA 110
	- Staphylococcus spateminus partial 105 fRVA gene, isolate OCAT31
	Staphylococcus epidermidis partial 105 rRNA gene isolate OCOR0
	Staphylococcus epidermidis strain Au22 16S ribosomal RNA cene nartial semence
	Staphylococcus epidermidis strain B7 3CO2 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. QD53 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis partial 16S rRNA gene, strain JPR-05
	Uncultured bacterium clone Ap.ba-F-DM-HN-1-27 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain W 3/4 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain W+5/13 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. PJRI24 IoS ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis PM221 complete genome
	y staphytococcus epidermidis strain ASI IOS fibosomal KNA gene, partial sequence
	Staphytococcus epideminicis strain C10 105 noosoniai Kivo gene, partial sequence
	Stanbulgeorens in R17 165 ribosonal RNA one narial sequence
	Staphylococcus and JPR 165 ribosonal RNA gene, partial sequence
	Staphylococcus epidermidis strain P 1/1 16S ribosomal RNA gene, partial sequence
	Bacterium EM-2014-133 genomic DNA containing 16S-23S intergenic spacer region, isolate 133
	Uncultured bacterium gene for 16S rRNA, partial sequence, clone: smkt_Fir_002_002
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Staphylococcus warneri KP771665.1

Staphylococcus warneri strain SuMS_N03 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KP771665.11</u> Length: 1463 Number of Matches: 1

Range 1:	112 to	563 GenBank Grap	V N	ext Match 🔺 Previous Mato	:h	
Score		Expect	Identities	Gaps	Strand	
807 bits	(894)	0.0	450/452(99%)	0/452(0%)	Plus/Plus	
Query	5	ACACGTGGAT			GAAACCGGAGCTAAT	ACCGG 64
Sbjct	112	ACACGTGGAT	ACCTACCTATAAGAC	TGGGATAACTTCGG	GAAACCGGAGCTAAT	ACCGG 171
Query	65	ATAACCCATTO	SAACCGCATGGTTCAA	TAGTGAAAGGCGGG	CTTTGCTGTCACTTAT	AGATG 124
Sbjct	172	ATAACATATT	SAACCGCATGGTTCAA	TAGTGAAAGGCGGC	TTTGCTGTCACTTAT	AGATG 231
Query	125	GATCCGCGCCC	GTATTAGCTAGTTGGI			GTAGC 184
Sbjct	232	GATCCGCGCCC	TATTAGCTAGTTGGI	AAGGTAACGGCTTA	CCAAGGCAACGATAC	GTAGC 291
Query	185	CGACCTGAGAG	GGTGATCGGCCACAC	TGGAACTGAGACAC	CGGTCCAGACTCCTAC	GGGAG 244
Sbjct	292	CGACCTGAGAG	GGTGATCGGCCACAC	TGGAACTGAGACAC	CGGTCCAGACTCCTAC	GGGAG 351
Query	245	GCAGCAGTAG	GAATCTTCCGCAATG	GGCGAAAGCCTGAC	CGGAGCAACGCCGCGT	GAGTG 304
Sbjct	352	GCAGCAGTAG	GAATCTTCCGCAATG	GGCGAAAGCCTGAC	CGGAGCAACGCCGCGT	GAGTG 411
Query	305	ATGAAGGTCT	CGGATCGTAAAACTC	TGTTATCAGGGAAG	SAACAAATGTGTAAGT	AACTG 364
Sbjct	412	ATGAAGGTCT	CGGATCGTAAAACTC	TGTTATCAGGGAAG	GAACAAATGTGTAAGT	AACTG 471
Query	365	TGCACATCTT	GACGGTACCTGATCAG	AAAGCCACGGCTA	ACTACGTGCCAGCAGC	CGCGG 424
Sbjct	472	TGCACATCTT	GACGGTACCTGATCAG	BAAAGCCACGGCTA	ACTACGTGCCAGCAGC	CGCGG 531
Query	425	TAATACGTAG	GTGGCAAGCGTTATCC	GGAATT 456		
Sbjct	532	TAATACGTAG	GTGGCAAGCGTTATCC	GGAATT 563		

VelQuery, 6441
 Staphyloccccus sp. Int 23 165 ribosomal RNA gene, partial sequence
 Staphyloccccus sp. partial 165 rRNA gene, joidat 20, 141
 Staphyloccccus sp. partial 165 rRNA gene, joidat 20, 141
 Staphylocccus swameri SGI attrains SGI 165 ribosomal RNA gene, partial sequence
 Staphylocccus swameri SGI attrains SGI 165 ribosomal RNA gene, partial sequence
 Staphylocccus swameri Starin 11BP 165 ribosomal RNA gene, partial sequence
 Staphylocccus swameri Starin 2000 H23 165 ribosomal RNA gene, partial sequence
 Staphylocccus swameri Starin 2000 H24 165 ribosomal RNA gene, partial sequence
 Staphylocccus swameri strina 7.0602 165 ribosomal RNA gene, partial sequence
 Staphylocccus wameri strina 7.06120 165 ribosomal RNA gene, partial sequence
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 Staphylocccus wameri strina 7.06120 165 ribosomal RNA gene, partial sequence
 Staphylocccus wameri strina 7.06120 165 ribosomal RNA gene, partial sequence
 Staphylocccus wameri strina 7.0516 ribosomal RNA gene, partial sequence
 Staphylocccus wameri strina 7.0516 ribosomal RNA gene, partial sequence
 Staphylocccus wameri strina 7.0516 ribosomal RNA gene, partial sequence
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 Staphylocccus wameri strina 7.072 166 ribosomal RNA gene, partial sequence
 Staphylocccus wameri strina 7.116 (RB11-51) fib ribosomal RNA gene, partial sequence
 Staphylocccus wameri strina 7.116 (RB11-51) fib ribosomal RNA gene, partial sequence
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 Staphylocccus wameri strina 7.1116 (RB11-51) fib ribosomal RNA ge

Bacillus subtilis HG764646.1

Bacillus subtilis partial 16S rRNA gene, isolate C737 Sequence ID: <u>emb|HG764646.11</u> Length: 995 Number of Matches: 1

Range 1: 1	3 to 95	1 GenBank Graphics		V Ne	ext Match 🔺 Previous Match	
Score 1656 bits	(1836)	Expect 0.0	Identities 931/939(99%)	Gaps 1/939(0%)	Strand Plus/Minus	
Query	1	CTTCGGGTGTTA	CAAGCTCTCGTGGT	STGACGGGCGGTGT	GTACAAGGCCCGGGAACGTA	60
Sbjct	951	CTTCGGGTGTTA	CAAACTCTCGTGGT	GTGACGGGCGGTGT	GTACAAGGCCCGGGAACGTA	892
Query	61	TTCACCGCGGCA	TGCTGATCCGCGAT	TACTAGCGATTCCA	GCTTCACGCAGTCGAGTTGC	120
Sbjct	891	TTCACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGC			832	
Query	121	AGACTGCGATCC	GAACTGAGAACAGA	TTGTGGGATTGGC	TTAACCTCGCGGTTTCGCTG	180
Sbjct	831	AGACTGCGATCC	GAACTGAGAACAGAT	TTGTGGGATTGGC	TTAACCTCGCGGTTTCGCTG	772
Query	181	CCCTTTGTTCTG	TCCATTGTAGCACG	IGTGTAGCCCAGGT	CATAAGGGGCATGATGATTT	240
Sbjct	771	CCCTTTGTTCTG	TCCATTGTAGCACG	GTGTAGCCCAGGT	CATAAGGGGCATGATGATTT	712
Query	241	GACGTCATCCCC	ACCTTCCTCCGGTT	IGTCACCGGCAGTC	ACCTTAGAGTGCCCAACTGA	300
Sbjct	711	GACGTCATCCCC.	ACCTTCCTCCGGTT	GTCACCGGCAGTC	ACCTTAGAGTGCCCAACTGA	652
Query	301	ATGCTGGCAACT.	AAGATCAAGGGTTG	CGCTCGTTGCGGGA	CTTAACCCAACATCTCACGA	360
Sbjct	651	ATGCTGGCAACT.	AAGATCAAGGGTTG	CGCTCGTTGCGGGA	CTTAACCCAACATCTCACGA	592
Query	361	CACGAGCTGACG	ACAACCATGCACCAC	CTGTCACTCTGCC	CCCGAAGGGGACGTCCTATC	420
Sbjct	591	CACGAGCTGACGACAACCATGCACCACCTGTCACTCTGCCCCCGAAGGGGACGTCCTATC	532			
Query	421	TCTAGGATTGTC	AGAGGATGTCAAGAG	CTGGTAAGGTTCT	TCGCGTTGCTTCGAATTAAA	480
Sbjct	531	TCTAGGATTGTC.	AGAGGATGTCAAGAG	CTGGTAAGGTTCT	TCGCGTTGCTTCGAATTAAA	472
Query	481	CCACATGCTCCA	CCGCTTGTGCGGGCC	CCCGTCAATTCCT	TTGAGTTTCAGTCTTGCGAC	540
Sbjct	471	CCACATGCTCCA	CCGCTTGTGCGGGCC	CCCGTCAATTCCT	TTGAGTTTCAGTCTTGCGAC	412
Query	541	CGTACTCCCCAN	GCGGAGTGCTTAAT	GCGTTAGCTGCAG	CACTAAGGGGGGGGAAACCCCC	600
Sbjct	411	CGTACTCCCCAG	GCGGAGTGCTTAAT	GCGTTAGCTGCAG	CACTAAGGGGCGGAAACCCCC	352
Query	601	TAACACTTAGCA	CACTCATCGTTTACGGCGTGGACTA	CGTGGACTACCAG	GGTATCTAATCCTGTTCGCTC	660
Sbjct	351	TAACACTTAGCA	CTCATCGTTTACGG	CGTGGACTACCAG	GTATCTAATCCTGTTCGCTC	292
Query	661	CCCACGCTTTCG	CTCCTCAGCGTCAG	TTACAGANCAGAGA	AGTCGCCTTCGCCACTGGTGT	720
Sbjct	291	CCCACGCTTTCG	CTCCTCAGCGTCAG	TTACAGACCAGAGA	AGTCGCCTTCGCCACTGGTGT	232
Query	721	TCCTCCACATCT	CTACGCATTTCACC	GCTACACGTGGAAJ	TTCCACTCTCCTCTTCTGCAC	780
Sbjct	231	TCCTCCACATCT	CTACGCATTTCACC	GCTACACGTGGAAI	TTCCACTCTCCTCTTCTGCAC	172
Query	781	TCAAGTTCCCCA	GTTTCCAATGACCC	TCCCCGGTTGAGCO	CGGGGGCTTTCACATCAAACT	840
Sbjct	171	TCAAGTTCCCCA	GTTTCCAATGACCC	TCCCCGGTTGAGCO	CGGGGGCTTTCACATCAGACT	112
Query	841	TAAAAAACCGCC	TGCGAGCCCTTTAC	GCCCAATAATTCCC	CGGA-AACGCTTGCCCCCTAC	899
Sbjct	111	TAAGAAACCGCC	TGCGAGCCCTTTAC	GCCCAATAATTCCC	CGGACAACGCTTGCCACCTAC	52
Query	900	GTATTACCGCGG	CTGCTGGCACGTAG	TTAGCCGGGGGCTT	938	
Sbjct	51	GTATTACCGCGG	CTGCTGGCACGTAG	TTAGCCGTGGCTT	13	



Bacillus cereus KJ612539.1

Bacillus cereus strain EM13 16S ribosomal RNA gene, partial sequence sequence ID: $\underline{gb[KJ612539.1]}$ Length: 1397 Number of Matches: 1

Range 1: 431 to 1378 GenBank Graphics Vext Match A Previous Match							
Score 1660 bits	(1840)	Expect 0.0	Identities 941/951(99%)	Gaps 3/951(0%)	Strand Plus/Minus		
Query	1	GTTACAAACTC	CGTGGTGTGACGGGC	GGTGTGTACAAG	GCCCGGGAACGTA	TCACCGC	60
Sbjct	1378	GTTACAAACTC	CGTGGTGTGACGGGC	 GGTGTGTACAAG	GCCCGGGAACGTA	 TTCACCGC	1319
Query	61	GGCATGCTGAT	CCGCGATTACTAGCGA	TTCCAGCTTCAT	GTAGGCGAGTTGC	AGCCTACA	120
Sbjct	1318	GGCATGCTGAT	CGCGATTACTAGCGA	TTCCAGCTTCAT	 GTAGGCGAGTTGCA	AGCCTACA	1259
Query	121	ATCCGAACTGA	GAACGGTTTTATGAGA	TTAGCTCCACCT	CGCGGTCTTGCAG	CTCTTTGT	180
Sbjct	1258	ATCCGAACTGA	GAACGGTTTTATGAGA	TTAGCTCCACCT	CGCGGTCTTGCAG	CTCTTTGT	1199
Query	181	ACCGTCCATTG	TAGCACGTGTGTAGCC	CAGGTCATAAGG	GGCATGATGATTT	GACGTCAT	240
Sbjct	1198	ACCGTCCATTG	PAGCACGTGTGTAGCC	CAGGTCATAAGG	GGCATGATGATTT	GACGTCAT	1139
Query	241	CCCCACCTTCC	ICCGGTTTGTCACCGG	CAGTCACCTTAG	AGTGCCCAACTTA	ATGATGGC	300
Sbjct	1138	CCCCACCTTCC	ICCGGTTTGTCACCGG	CAGTCACCTTAG	AGTGCCCAACTTA	ATGATGGC	1079
Query	301	AACTAAGATCA	AGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACG	CAACATCTCACGA	CACGAGCT	360	
Sbjct	1078	AACTAAGATCA	AGGGTTGCGCTCGTTG	CGGGACTTAACC	CAACATCTCACGA	CACGAGCT	1019
Query	361	GACGACAACCA	IGCACCACCTGTCACT	CTGCTCCCGAAG	GAGAAGCCCTATC	ICTAGGGT	420
Sbjct	1018	GACGACAACCA	IGCACCACCTGTCACT	CTGCTCCCGAAG	GAGAAGCCCTATC	TCTAGGGT	959
Query	421	TTTCAGAGGAT	STCAAGACCTGGTAAG	GTTCTTCGCGTT	GCTTCGAATTAAA	CCACATGC	480
Sbjct	958	TTTCAGAGGAT	GTCAAGACCTGGTAAG	GTTCTTCGCGTT	GCTTCGAATTAAA	CCACATGC	899
Query	481	TCCACCGCTTG	IGCGGGCCCCCGTCAA	TTCCTTTGAGTT	TCAGCCTTGCGGC	CGTACTCC	540
Sbjct	898	TCCACCGCTTG	IGCGGGCCCCCGTCAA	TTCCTTTGAGTT	TCAGCCTTGCGGC	CGTACTCC	839
Query	541	CCAGGCGGAGT	GCTTAATGCGTTAACT	TCAGCACTAAAG	GGCGGAAACCCTC	FAACACTT	600
Sbjct	838	CCAGGCGGAGT	GCTTAATGCGTTAACT	TCAGCACTAAAG	GGCGGAAACCCTC	FAACACTT	779
Query	601	AGCACTCATCG	ITTACGGCGTGGACTA	CCAGGGTATCTA	ATCCTGTTTGCTC	CCCACGCT	660
Sbjct	778	AGCACTCATCG	ITTACGGCGTGGACTA	CCAGGGTATCTA	ATCCTGTTTGCTC	CCCACGCT	719
Query	661	TTCGCGCCTCA	GTGTCAGTTACAGACC	AGAAAGTCGCCT	TCGCCACTGGTGT	ICCTCCAT	720
Sbjct	718	TTCGCGCCTCA	GTGTCAGTTACAGACC	AGAAAGTCGCCT	TCGCCACTGGTGT	ICCTCCAT	659
Query	721	ATCTCTACGCA	ITTCACCGCTACACAT	GGAATTCCACTT	TCCTCTTCTGCAC:	FCAAGTCT	780
Sbjct	658	ATCTCTACGCA	ITTCACCGCTACACAT	GGAATTCCACTT	TCCTCTTCTGCAC	ICAAGTCT	599
Query	781	CCCAGTTTCCA	ATGACCCTCCACGGGT	TGAGCCGTGGGC	TTTCACATCAAAG	ГТАААААА	840
Sbjct	598	CCCAGTTTCCA	ATGACCCTCCAC-GGT	TGAGCCGTGGGC	TTTCACATCAGAC	IIII III FTAAGAAA	540
Query	841	CCACCTGCGCGG	CGCTTTACGCCCAATA	ATTCCCGGAAAA	CGCTTGCCCCCTA	CGTATTAC	900
Sbjct	539	CCACCTGCGCG	CGCTTTACGCCCAATA	ATTCCCGGATAA	CGCTTGCCACCTA	CGTATTAC	480
Query	901	CGCGGCTGCTG	GCACGTAATTAGCCGG	GGCTTTCCTGGT	TAAGGTACCGTC	951	
Sbjct	479	CGCGGCTGCTG	GCACGTAGTTAGCCGT	GGCTTT-CTGGT	T-AGGTACCGTC	431	

	Icl Query	47323		
	• • • •	Uncultured	bacterium partial 16S rRNA gene, isolate Be20	
		Bacillue a	Uncultured Bacillus sp. gene for 16S ribosomal RNA, partial sequence, isolate: XZX-159 athracis strain 1-Si-5-2-3-M 16S ribosomal PNA gene partial sequence	/
		Bacillus a	3. ROIS-2 16S ribosomal RNA cene, nartial sequence	
		Bacillus s	p. BBAR-01d 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. QY1 16S ribosomal RNA gene, partial sequence	
		Bacillus o	ereus strain EM13 16S ribosomal RNA gene, partial sequence	
		Bacillus o	ereus strain JC80 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. XM1 16S ribosomal RNA gene, partial sequence	
		Bacillus (ereus partial 165 rKNA gene, isolate BCsi	
		Dacinus s	Sinos gene for 105 rRivA, partial sequence A gene nartial sequence	
		5	Uncultured bacterium partial 16S rRNA gene, isolate BF0001D030	
			Bacillus sp. ES2-45 16S ribosomal RNA gene, partial sequ	uence
		1	firmicutes 2 leaves	
		Bacillus t	nuringiensis strain RB90 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. hb39(2014) 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. VOC11 16S ribosomal RNA gene, partial sequence	
			Bacilius sp. nb55 165 nbosomal KNA gene, partial sequence	- Bactarium C0528 165 ribosomal PNA gana partial seguence
		Bacillus s	p. I4X 16S ribosomal RNA gene, partial sequence	· bacterian coszo ros noosonar revergene, partar sequence
		Bacillus o	ereus strain NM2E11 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. FGG1 16S ribosomal RNA gene, partial sequence	
		Unculture	d bacterium clone PT_19 16S ribosomal RNA gene, partial sequence	
		Unculture	d bacterium clone PT_32 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. DAB-4041 105 noosonal KNA gene, partial sequence	
		Bacillus t	uringiensis strain IHB B 7117 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. N5/665 partial 16S rRNA gene, strain N5/665	
		Onculture	d bacterium partial 16S rRNA gene, isolate Br2	
	1	Unculture	d bacterium partial 16S rRNA gene, isolate Be21	
		Bacillus s	p. EM7 16S rRNA gene, isolate EM7	
		Bacillus a	nthracis 168 rKNA gene, isolate EM36	
		Bacillus s	ubtilis strain PVR-YHB-1-1 16S ribosomal RNA gene, partial sequence	
		Bacillus a	nthracis strain BHR1P2B2 16S ribosomal RNA gene, partial sequence	
		Bacillus s	p. BS24 16S ribosomal RNA gene, partial sequence	
		Bacillus t	huringiensis strain FJM2 16S ribosomal RNA gene, partial sequence	
		Unculture	d Bacillus sp. gene for 16S ribosomal RNA, partial sequence, isolate: QX-54	
		Bacillus s	p. DB14544 16S ribosomal RNA gene, partial sequence	
		Linculture	d Bacillus sp. gene for 16S ribosomal RNA, partial sequence, isolate: OX-85	
		Bacillus t	uringiensis strain FJM3 16S ribosomal RNA gene, partial sequence	
		Bacillus o	ereus strain 5A 16S ribosomal RNA gene, partial sequence	
	1	Bacillus o	ereus strain BS1 16S ribosomal RNA gene, partial sequence	
		Baciflus ce Bacillus th	eus strain JMG-03 16S ribosomal KNA gene, partial sequence iringiansis strain GZDE1 16S ribosomal RNA gana, partial sequence	
		Bacillus sp	WM64 16S rRNA gene, isolate WM64	
		Bacillus an	thracis 16S rRNA gene, isolate EM11	
		Bacillus sp	SF2-1 16S ribosomal RNA gene, partial sequence	
		Bacillus ce	reus strain LP10 S10 16S ribosomal RNA gene, partial sequence	
		Bacillus th	ringiensis strain IHB B 7196 16S ribosomal RNA gene, partial sequence	
		Bacillus an	thracis strain IHB B 7021 16S ribosomal RNA gene, partial sequence	
		Uncultured	bacterium clone PT 38 16S ribosomal RNA gene, partial sequence	
		Uncultured	bacterium clone PT_22 16S ribosomal RNA gene, partial sequence	
		Bacillus th	iringiensis serovar galleriae strain HD-29, complete genome	
		Bacillus sp Bacillus an	B1(2015) 16S ribosomal KNA gene, partial sequence thracis strain IHR B 18033 16S ribosomal RNA gene partial sequence	
		Uncultured	Bacillus sp. clone RHDTWG188 16S ribosomal RNA gene, partial sequence	
		Uncultured	Bacillus sp. clone RHDTWG185 16S ribosomal RNA gene, partial sequence	
		Uncultured	Bacillus sp. clone KHD1WG181 16S ribosomal KNA gene, partial sequence Bacillus sp. clone C6A08 16S ribosomal PNA gene, partial sequence	
		Uncultured	Bacillus sp. clone C6A22 16S ribosomal RNA gene, partial sequence	
		Uncultured	Bacillus sp. clone C5B09-1 16S ribosomal RNA gene, partial sequence	
		Bacillus so	INBio 4516D 16S ribosomal RNA gene, partial sequence	
		Bacillus sp	INBio_4312S 16S ribosomal RNA gene, partial sequence	
		Bacillus co	reus strain DC2 16S ribosomal RNA gene, partial sequence	
		firmicute	i bacierium parual 165 fKNA gene, isolate B134	
		Uncultured	Bacillus sp. gene for 16S ribosomal RNA, partial sequence, isolate: LZX-54	
		Bacillus ce	reus strain FJM4 16S ribosomal RNA gene, partial sequence	
		Bacillus ce Bacillus ce	reus strain 11B 16S ribosomal RNA gene, partial sequence	
		Bacillus ce	reus strain MHS 16S ribosomal RNA gene, partial sequence	
		Bacillus ce	reus strain NXUGDS005 16S ribosomal RNA gene, partial sequence	
		Bacillus ce	reus strain K2-1 16S ribosomal RNA gene, partial sequence	
		Bacillus sp	JBS-28 16S ribosomal RNA gene, partial sequence	
		Uncultur	ed bacterium partial 16S rRNA gene, isolate Be6	
		Bacillus	cereus strain KG5 16S ribosomal RNA gene, partial sequence	
		Bacillus	huringiensis strain IHB B 7206 16S ribosomal RNA gene, partial sequence	
		Bacillus	nuringiensis strain IHB B 7070 16S ribosomal RNA gene, partial sequence	
		Bacillus	nuringrensis strain HD807 105 ribosomal RNA gene, partial sequence	
		Bacilluc	zu bacterium cione F1_40-105 ribosomai KNA gene, partial sequence	
		Bacillus	prigrate ros noosonial revergence, partial sequence	
		Bacillus	sp. CMAP11 16S ribosomal RNA gene, partial sequence	
		firmicut	s 3 leaves	
		Unidenti	ied bacterium clone MEB002 16S ribosomal RNA gene, partial sequence	
.0.001		Bacillus	sp. SMF5 partial 16S rRNA gene	
			 Uncultured prokaryote clone seq_M-L8_16SR 16S ribosomal RNA ger 	ne, partial sequence
		Bacillus	p. INBio_4109M 16S ribosomal RNA gene, partial sequence	



5- The phylogenetic analysis of bacteria isolated from lift buttons samples

Staphylococcus warneri BCL-34

Staphylococcus warneri strain MBS022 16S ribosomal RNA gene, partial sequence Sequence ID: <u>KT582294.1</u> Length: 1486 Number of Matches: 1

Range 1:	Range 1: 820 to 1424 GenBank Graphics 💎 Next Match 🔺 Previous Match						
Score 1077 bit	ts(583)	Expect 0.0	Identities 599/606(99%)	Gaps 3/606(0%)	Strand Plus/Minus		
Query	19	TAAATGGTI	ACTCCACCGGCTTCG	GGTGTTACAAACT	CTCGTGGTGTGACGGGCGGT	GTG 78	
Sbjct	1424	TAAATGGT	ACTCCACCGGCTTCG	GGTGTTACAAACT		 GTG 1365	
Query	79	TACAAGACO	CGGGAACGTATTCAC	CGTAGCATGCTGA	ICTACGATTACTAGCGATTC	CAG 138	
Sbjct	1364	TACAAGACO	CGGGAACGTATTCAC	CGTAGCATGCTGA	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	CAG 1305	
Query	139	CTTCATGTZ	GTCGAGTTGCAGACT	ACAATCCGAACTG	AGAACAACTTTATGGGATTT	GCT 198	
Sbjct	1304	CTTCATGT	AGTCGAGTTGCAGACT.	ACAATCCGAACTG	AGAACAACTTTATGGGATTT	III GCT 1245	
Query	199	TGACCTCGO	GGTTTAGCTGCCCTT	TGTATTGTCCATT	GTAGCACGTGTGTAGCCCAA	ATC 258	
Sbjct	1244	TGACCTCG	GGTTTAGCTGCCCTT	TGTATTGTCCATT	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII		
Query	259	ATAAGGGGG	ATGATGATTTGACGT	CATCCCCACCTTC	CTCCGGTTTGTCACCGGCAG	TCA 318	
Sbjct	1184	ATAAGGGGG	ATGATGATTTGACGT	CATCCCCACCTTC	CTCCGGTTTGTCACCGGCAG	 TCA 1125	
Query	319	ACTTAGAGI	GCCCAACTTAATGAT	GGCAACTAAGCTT	AAGGGTTGCGCTCGTTGCGG	GAC 378	
Sbjct	1124	ACTTAGAG	GCCCAACTTAATGAT	GGCAACTAAGCTT	AAGGGTTGCGCTCGTTGCGG	 GAC 1065	
Query	379	TTAACCCAR	CATCTCACGACACGA	GCTGACGACAACC	ATGCACCACCTGTCACTTTG	TCC 438	
Sbjct	1064	TTAACCCAZ	CATCTCACGACACGA	GCTGACGACAACC	ATGCACCACCTGTCACTTTG	 FCC 1005	
Query	439	CCCGAAGGG	GAAGACTCTATCTCT	TAGAGCGGTCAAAGGATGTCAAGATTTGGTAAGG	TTC 498		
Sbjct	1004	CCCGAAGGG	GAAGACTCTATCTCT	AGAGCGGTCAAAG	GATGTCAAGATTTGGTAAGG	 FTC 945	
Query	499	TTCGCGTTG	CTTCAAATTAAACCA	CATGCTCCACCGT	IGGTGCGGCCCCGTCAAT	TCT 556	
Sbjct	944	TTCGCGTTG	CTTCAAATTAAACCA	CATGCTCCACCGC	 TTGTGCGGGGTCCCCGTCAAT	 !TCC 885	
Query	557	TTTGATTT	CAACCTTGCGGTCGT	ACTCCCCAGGCGG	AGTGCTTAATGCGTTTAGCT	GCA 616	
Sbjct	884	TTTGAGTT	CAACCTTGCGGTCGT	ACTCCCCAGGCGG	AGTGCTTAATGCGTT-AGCT	III GCA 826	
Query	617	GCACTA (522				
Sbjct	825	GCACTA 8	20				

	firmicutes 2 leaves	
ф (Staphylococcus warneri strain MBS022 16S ribosomal RNA gene, part	tial sequence
		Staphylococcus pasteuri strain MBS011 16S ribosomal RNA gene, partial sequence
	•	Staphylococcus warneri strain CPI 2 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. ChDC B392 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. ChDC B393 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. PSB-18 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. PSB-21 I6S ribosomal RNA gene, partial sequence
		Psychrobacter pulmonis strain SL-9 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. DF6n-C small subunit ribosomal RNA gene, partial sequence
		Staphylococcus pasteuri partial 16S rRNA gene, isolate OCATI
		Staphylococcus warneri partial 16S rRNA gene, isolate OCAT2
		Staphylococcus warneri partial 16S rRNA gene, isolate OCAT9
		Staphylococcus sp. OCAT11 partial 16S rRNA gene, isolate OCAT11
		Staphylococcus warneri partial 16S rRNA gene, isolate OCAT13
		Staphylococcus sp. OCAT14 partial 16S rRNA gene, isolate OCAT14
		Staphylococcus warneri partial 16S rRNA gene, isolate OCAT15
		Staphylococcus warneri partial I6S rRNA gene, isolate OCAT20
		Staphylococcus warneri partial 16S rRNA gene, isolate OCOB3
		Staphylococcus warneri partial 16S rRNA gene, isolate OCOB8
		Uncultured Staphylococcus sp. clone JU-EB2 16S ribosomal RNA gene, partial sequence
		Staphylococcus warneri strain JU-SB2 16S ribosomal RNA gene, partial sequence
		Staphylococcus warneri strain MRS-2 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. DVRSG-2 16S ribosomal RNA gene, partial sequence
		Staphylococcus warneri strain LH-T2 16S ribosomal RNA gene, partial sequence
		Staphylococcus warneri strain YZ-T1 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. NBM-83 16S ribosomal RNA gene, partial sequence
		Staphylococcus warneri strain 81b 16S ribosomal RNA gene, partial sequence
		Staphylococcus pasteuri strain MJMG7.13 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. P34 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. Bts 21b partial 16S rRNA gene, isolate Bts 21b
		Staphylococcus warneri strain STP02 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. enrichment culture clone zhuowei 16S ribosomal RNA gene, partial sequence
		Staphylococcus warneri strain RJ24 16S ribosomal RNA gene, partial sequence
		Staphylococcus pasteuri strain CS.B10 16S ribosomal RNA gene, partial sequence
		Staphylococcus pasteuri strain MAL_IA I6S ribosomal RNA gene, partial sequence
		Staphylococcus warneri strain MOH119 16S ribosomai RNA gene, partial sequence
		Staphylococcus sp. CE3E4 16S nbosomal RNA gene, partial sequence
		Staphylococcus sp. CE3E5 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. CE3E1 16S ribosomal RNA gene, partial sequence
		Staphylococcus pasteuri strain KG-63 168 ribosomal RNA gene, partial sequence
		Staphylococcus warneri gene for 165 ribosomal RNA, partial sequence, strain: SH13
		Staphylococcus warneri strain 1058 165 ribosomal RNA gene, partial sequence
		Stapnylococcus sp. \$7151a 105 mbosomat KNA gene, partial sequence
8,0004		Uncultured bacterium clone M21P1F10 16S ribosomal RNA gene, partial sequence
		Girmicutes 55 Jeaves
		- Interesting 22 million

Micrococcus luteus MBS022

Micrococcus luteus strain BCL-34 16S ribosomal RNA gene, partial sequence Sequence ID: <u>KM378607.1</u> Length: 1394 Number of Matches: 1

Range 1: 1 to 1393 GenBank Graphics Vext Match 🛦 Previous Match						
Score 2560 bits	s(1386)	Expect 0.0	Identities 1391/1393(99%)	Gaps 1/1393(0%)	Strand Plus/Plus	
Query	1	TGC-AGTCGAA	CGATGAAGCCCAGCTT	GCTGGGTGGATTA	GTGGCGAACGGGTGAGTAAC	59
Sbjct	1	TGCAAGTCGAA	CGATGAAGCCCAGCTT	GCTGGGTGGATTA		60
Query	60	ACGTGAGTAAC	CTGCCCTTAACTCTGG	GATAAGCCTGGGA	AACTGGGTCTAATACCGGAT	119
Sbjct	61	ACGTGAGTAAC	CTGCCCTTAACTCTGG	GATAAGCCTGGGA	AACTGGGTCTAATACCGGAT	120
Query	120	AGGAGCGCCGA	CCGCATGGTGGGTGTT	GGAAAGATTTATCO	GGTTTTGGATGGACTCGCGG	179
Sbjct	121	AGGAGCGCCTA	CCGCATGGTGGGTGTT	GGAAAGATTTATCO		180
Query	180	CCTATCAGCTT	GTTGGTGAGGTAATGG	CTCACCAAGGCGA	CGACGGGTAGCCGGCCTGAG	239
Sbjct	181	CCTATCAGCTT	GTTGGTGAGGTAATGG	CTCACCAAGGCGA	CGACGGGTAGCCGGCCTGAG	240
Query	240	AGGGTGACCGG	CCACACTGGGACTGAG	ACACGGCCCAGAC	ICCTACGGGAGGCAGCAGTG	299
Sbjct	241	AGGGTGACCGG	CCACACTGGGACTGAG	ACACGGCCCAGAC		300
Query	300	GGGAATATTGC	ACAATGGGCGCAAGCC	TGATGCAGCGACG	CCGCGTGAGGGATGACGGCC	359
Sbjct	301	GGGAATATTGC	 ACAATGGGCGCAAGCC	 TGATGCAGCGACG(CCGCGTGAGGGATGACGGCC	360
Query	360	TTCGGGTTGTA	AACCTCTTTCAGTAGG	GAAGAAGCGAAAG'	IGACGGTACCTGCAGAAGAA	419
Sbjct	361	TTCGGGTTGTA	AACCTCTTTCAGTAGG	GAAGAAGCGAAAG'		420
Query	420	GCACCGGCTAA	CTACGTGCCAGCAGCC	GCGGTAATACGTA	GGGTGCGAGCGTTATCCGGA	479
Sbjct	421	GCACCGGCTAA	CTACGTGCCAGCAGCC	 GCGGTAATACGTA		480
Query	480	ATTATTGGGCG	TAAAGAGCTCGTAGGC	GGTTTGTCGCGTC	IGTCGTGAAAGTCCGGGGCT	539
Sbjct	481	ATTATTGGGCG	TAAAGAGCTCGTAGGC	GGTTTGTCGCGTC		540
Query	540	TAACCCCGGAT	CTGCGGTGGGTACGGG	CAGACTAGAGTGC	AGTAGGGGAGACTGGAATTC	599
Sbjct	541	TAACCCCGGAT	CTGCGGTGGGTACGGG	CAGACTAGAGTGC		600
Query	600	CTGGTGTAGCG	GTGGAATGCGCAGATA	TCAGGAGGAACAC	CGATGGCGAAGGCAGGTCTC	659
Sbjct	601	CTGGTGTAGCG	GTGGAATGCGCAGATA	TCAGGAGGAACAC	CGATGGCGAAGGCAGGTCTC	660
Query	660	TGGGCTGTAAC	TGACGCTGAGGAGCGA	AAGCATGGGGAGCO	GAACAGGATTAGATACCCTG	719
Sbjct	661 720	TGGGCTGTAAC	TGACGCTGAGGAGCGA	AAGCATGGGGAGC	SAACAGGATTAGATACCCTG	720
Sbjct	721	GTAGTCCATGO	CGTAAACGTTGGGCAC	TAGGTGTGGGGAC	CATTCCACGGTTTCCGCGCC	780
Query	780	GCAGCTAACGC	ATTAAGTGCCCCGCCT	GGGGAGTACGGCC	GCAAGGCTAAAACTCAAAGG	839
Sbjct	781	GCAGCTAACGC	ATTAAGTGCCCCGCCT	GGGGAGTACGGCCC	GCAAGGCTAAAACTCAAAGG	840
Query	840	AATTGACGGGG		GAGCATGCGGATTZ	AATTCGATGCAACGCGAAGA	899
Query	900	ACCTTACCAAG	GCTTGACATGTTCTCG	ATCGCCGTAGAGAS	TACGGTTTCCCCTTTGGGGC	959
Sbjct	901	ACCTTACCAAG	GCTTGACATGTTCTCG	ATCGCCGTAGAGAS	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	960
Query	960	GGGTTCACAGG	TGGTGCATGGTTGTCG	TCAGCTCGTGTCG	IGAGATGTTGGGTTAAGTCC	1019
Sbjet	961	GGGTTCACAGG	TGGTGCATGGTTGTCG	TCAGCTCGTGTCG	rgagatgttggggttaagtcc	1020
Query	1020	CGCAACGAGCG	CAACCCTCGTTCCATG	TTGCCAGCACGTA TTGCCAGCACGTA	ATGGTGGGGGACTCATGGGAG	1079
Query	1080	ACTGCCGGGGT	CAACTCGGAGGAAGGT	GAGGACGACGTCA	AATCATCATGCCCCTTATGT	1139
Sbjet	1081	ACTGCCGGGGT	CAACTCGGAGGAAGGT	GAGGACGACGTCA	AATCATCATGCCCCTTATGT	1140
Query	1140	CTTGGGCTTCA	CGCATGCTACAATGGC	CGGTACAATGGGT	IGCGATACTGTGAGGTGGAG	1199
Sbjct	1141	CTTGGGCTTCA	CGCATGCTACAATGGC	CGGTACAATGGGT	IGCGATACTGTGAGGTGGÅG	1200
Sbjct	1200	CTAATCCCAAA	AGCCGGTCTCAGTTC AAGCCGGTCTCAGTTC	GGATTGGGGTCTG	CAACTCGACCCCATGAAGTC	1260
Query	1260	GGAGTCGCTAG	TAATCGCAGATCAGCA	ACGCTGCGGTGAA	FACGTTCCCGGGCCTTGTAC	1319
Sbjet	1261	GGAGTCGCTAG	TAATCGCAGATCAGCA	ACGCTGCGGTGAA	PACGTTCCCGGGCCTTGTAC	1320
Query	1320	ACACCGCCCGT	CAAGTCACGAAAGTCG	GTAACACCCGAAG	CCGGTGGCCTAACCCTTGTG	1379
Sbjct	1321 1380	GGGGAGCCGT	CAAGTCACGAAAGTCG	GTAACACCCGAAG	CCGGTGGCCTAACCCTTGTG	1380
Sbjct	1381	Geggaecceto	 GA 1393			



Staphylococcus epidermidis

Staphylococcus epidermidis partial 16S rRNA gene, strain BGHMC5 Sequence ID: <u>FR797804.1</u> Length: 815 Number of Matches: 1

Range 1: 1 to 815 GenBank Graphics Vext Match 🛦 Previous Match							
Score	(045)	Expect	Identities	Gaps	Strand		
1506 bit	s(815)	0.0	815/815(100%)	0/815(0%)	Plus/Plus		
Query	1	AGAAAGTGGAA	TCCCATGTGTAGCGG	IGAAATGCGCAGA	GATATGGAGGAACACCAGTG	3-60 ∣	
Sbjct	1	1 AGAAAGTGGAATCCCATGTGTAGCGGTGAAATGCGCAGAGATATGGAGGAACACCAGTGG 60					
Query	61	CGAAGGCGACT	TTCTGGTCTGTAACT	GACGCTGATGTGC	GAAAGCGTGGGGATCAAACA	3 120	
Sbjct	61	CGAAGGCGACT	TTCTGGTCTGTAACT	SACGCTGATGTGC	GAAAGCGTGGGGATCAAACA	5 120	
Query	121	GATTAGATACC	CTGGTAGTCCACGCC	GTAAACGATGAGI	GCTAAGTGTTAGGGGGTTTC	c 180	
Sbjct	121	GATTAGATACC	CTGGTAGTCCACGCC	TAAACGATGAGI	GCTAAGTGTTAGGGGGTTTC	2 180	
Query	181	GCCCCTTAGTG	CTGCAGCTAACGCAT	TAAGCACTCCGCC	TGGGGAGTACGACCGCAAGG	r 240	
Sbjct	181	GCCCCTTAGTG	CTGCAGCTAACGCAT	TAAGCACTCCGCC	TGGGGAGTACGACCGCAAGG	r 240	
Query	241	TGAAACTCAAA	GGAATTGACGGGGGAC	CCGCACAAGCGGI	GGAGCATGTGGTTTAATTCG	A 300	
Sbjct	241	TGAAACTCAAA	GGAATTGACGGGGGAC	CCGCACAAGCGGI	GGAGCATGTGGTTTAATTCG	A 300	
Query	301	AGCAACGCGAA	GAACCTTACCAAATC	TTGACATCCTCTG	ACCCCTCTAGAGATAGAGTT	r 360	
Sbjct	301	AGCAACGCGAA	GAACCTTACCAAATC	TTGACATCCTCTC	ACCCCTCTAGAGATAGAGTT	r 360	
Query	361	TCCCCTTCGGG	GGACAGAGTGACAGG	IGGTGCATGGTTG	TCGTCAGCTCGTGTCGTGAG	A 420	
Sbjct	361	TCCCCTTCGGG	GGACAGAGTGACAGG	IGGTGCATGGTTG	TCGTCAGCTCGTGTCGTGAG	A 420	
Query	421	TGTTGGGTTAA	GTCCCGCAACGAGCG	CAACCCTTAAGCI	TAGTTGCCATCATTAAGTTG	3 480	
Sbjct	421	TGTTGGGTTAA	GTCCCGCAACGAGCG	CAACCCTTAAGCI	TAGTTGCCATCATTAAGTTG	480	
Query	481	GCACTCTAAGT	TGACTGCCGGTGACA		TGGGGATGACGTCAAATCAT	C 540	
Sbjct	481	GCACTCTAAGT	TGACTGCCGGTGACA	AACCGGAGGAAGG	TGGGGATGACGTCAAATCAT	540	
Query	541	ATGCCCCTTAT	GATTTGGGCTACACAC	CGTGCTACAATGG	ACAATACAAAGGGCAGCGAA	A 600	
Sbjct	541	ATGCCCCTTAT	GATTTGGGCTACACAC	CGTGCTACAATGG	ACAATACAAAGGGCAGCGAA	A 600	
Query	601	CCGCGAGGTCA	AGCAAATCCCATAAAG	GTTGTTCTCAGTI	CGGATTGTAGTCTGCAACTC	3 660	
Sbjct	601	CCGCGAGGTCA	AGCAAATCCCATAAAG	GTTGTTCTCAGTI	CGGATTGTAGTCTGCAACTC	- 5 660	
Query	661	ACTATATGAAG	CTGGAATCGCTAGTA	ATCGTAGATCAGO	ATGCTACGGTGAATACGTTC	c 720	
Sbjct	661	ACTATATGAAG	CTGGAATCGCTAGTA	ATCGTAGATCAGO	ATGCTACGGTGAATACGTTC	720	
Query	721	CGGGTCTTGTA	CACACCGCCCGTCAC	ACCACGAGAGTTI	GTAACACCCGAAGCCGGTGG	A 780	
Sbjct	721	CGGGTCTTGTA	CACACCGCCCGTCAC	ACCACGAGAGTTI	GTAACACCCGAAGCCGGTGG	A 780	
Query	781	GTAACCATTGG	AGCTAGCCGTTGAAG	GGGGACAAA 81	.5		
Sbjct	781	GTAACCATTGG	AGCTAGCCGTTGAAG	GGGACAAA 81	.5		

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9	 Staphylococcus epidermidis strain SEI, complete 	te genome
	Staphylococcus e	epidermidis strain 14.1.R1, complete genome
		Staphylococcus epidermidis strain PBR-19 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain GKY 8/1A 16S ribosomal RNA gene, partial sequence
		Oncultured Staphylococcus sp. clone 12L_53 16S holosomai KivA gene, partial sequence
		Stanbulococcus enidermidis strain TWSL 19 16S ribosomal RNA gene nartial sequence
		Staphylococcus epidermidis strain Fe-3a 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis partial 16S rRNA gene, isolate M0201
		Staphylococcus epidermidis strain SE4.6 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain SE4.8 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain LLP-16 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain BC7 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain 10A 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain S7 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain MB 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. YSL09-5 gene for 16S rRNA, partial sequence
		Staphylococcus sp. DGM Man1a 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain HNMCTR2 16S ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0096-T30-S-NI_000100 small subunit ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0169-T415-S-NIPCRAMgANa_000133 small subunit ribosomal RNA gene, partial sequence
		Uncultured organism clone EL U0169-1415-S-NIPCRAMgANa_000559 small subunit ribosomal RNA gene, partial sequence
		Staphylococcus epidemidis strain IHB B 12019 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidemiulus strain MJMUS.1 Ibs ribosomai RNA gene, partial sequence
		Uncultured organism clone EL 0002-5-117-5-5-NIPCRAMIGANa_000450 sman subunit ribosomia RVA gene, partial sequence
		Concurance organism clone ELU0109-1413-5-INFC/CAMBANA_000145 small subunit ribosomal KNA gene, partial sequence
		Organisat organism clone ELCOTOP (2005) or (Covergence_000020 small subunit fibosomal RIVA gene, palfial sequence
		Timmedies 8 leaves
		Staphylococcus epidermidis strain 23 16S ribosomal RNA gene, partial sequence
		Signify supervised and the second se Second second sec
		Uncuntred organism clone ELU0169-1415-5-NIPCKAMgANa_000262 small subunit ribosomal RNA gene, partial sequence
		Chountined organismic cone ELUUTO9-1413-5-INECRAMIGANA_000005 small subunit ribosomal KNA gene, partial sequence Stanbulacoccus enidermidis gene for 16S rBNA, partial sequence, etmin NEDC 12002
		Stanbulacoccus epiderinius gene for 105 refor, partial sequence, strain: NDRC 12995
		Stanhylococcus sp. Dom milita 103 flowoniai rovy gene, partial sequence
		Uncultured Staphylococcus sp. clone F3Baue.1 16S ribosomal RNA gene, partial sequence
		Stanhylococcus epidermidis strain F149 16S ribosomal RNA gene, nartial sequence
		Stanhylococcus epidermidis strain 80B 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain 104.1.2 16S ribosomal RNA gene, partial sequence
		Staphylococcus sp. ZW-3 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain IARI-CDK 18 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain SE4.7 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain SE2.9 16S ribosomal RNA gene, partial sequence
		Staphylococcus capitis subsp. capitis strain MER_TA_69.2 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain TWSL_17 16S ribosomal RNA gene, partial sequence
		Staphylococcus epidermidis strain KS3H17 16S ribosomal RNA gene, partial sequence
Dealthread bacterium ages for 16S rBNA partia	segmence clone enkt Fir 007 001	
	addressed ensure sum [1 a _ and _ and	
Tirmicutes 8 leaves		
Staphylococcus epidermidis strain O1 16S ribosc	mal RNA gene, partial sequence	
Uncultured organism clone ELU0169-T415-S-N	PCRAMgANa_000539 small subunit riboson	mal RNA gene, partial sequence
Uncultured organism clone ELU0169-T415-S-N	PCRAMgANa 000084 small subunit ribosor	mal RNA cene, partial sequence
Uncultured organism clone FI U0002-T15-S-NI	000452 small subunit ribosomal RNA gene	mantial samance
Unsultand Starbadesson an alore 200 160 mil	out to a state of the second state of the seco	han an addressed
Cheunared staphytocoecus sp. clone 268 165 ht	osomai KIVA gene, paruai sequence	
Staphylococcus epidermidis strain NM160-2 168	ribosomal KNA gene, partial sequence	
Staphylococcus epidermidis culture-collection IN	IAU:80829 16S ribosomal RNA gene, partial	I sequence
Uncultured bacterium partial 16S rRNA gene, clo	ne MB05G10	
Uncultured bacterium partial 16S rRNA gene, clo	me MB04C05	
Uncultured bacterium partial 16S rRNA gene, ele	me MB02F05	
Ouncultured bacterium partial 16S rRNA eene, ele	ne MB01H05	
Ouncultured bacterium partial 16S rPNA gene ele	me MB01G03	
Uncultured bactorium partial 165 rPNA sono al	ine SD05E11	
Unaultured basterium partial 165 rRNA gene, cit		
Uncultured bacterium partial 165 rRNA gene, ele	0007000	
Uncultured bacterium partial 165 rRNA gene, clo	ne 510/5106	
Uncultured bacterium partial 16S rRNA gene, clo	one SD04D06	
Uncultured bacterium partial 16S rRNA gene, clo	one SD02H09	
Uncultured bacterium partial 16S rRNA gene, ele	nc SD02F04	
Uncultured bacterium partial 16S rRNA gene, clo	one SD01F01	
Uncultured bacterium partial 16S rRNA sene. cla	nc SB04H03	
Uncultured bacterium partial 16S rPNA area al	one SB04G12	
Allocations of the stations and all (10 all 11	SPO1COR	
Concurrent bacterium partial 165 rRNA gene, cle	0004008	
Uncultured bacterium partial 16S rRNA gene, clo	ine SB04G01	
Uncultured bacterium partial 16S rRNA gene, ck	me SB04D06	
Uncultured bacterium partial 16S rRNA gene, cle	me SB04C12	
T		Staphylococcus epidermidis partial 16S rRNA gene, isolate H6-16S-LEGIO-BO
QUncultured bacterium partial 16S rRNA gene, ck	me SB04D08	 An ensurement of the approximation of the approximation of the state o
Uncultured bacterium nartial 16S rRNA sens, ck	ine SB03G05	
9		Stanbylococcus sp. JCM 10537 gene for 16S ribosonial RNA, partial sequence
Of Insultrand homenian marial 122 aP\$14	SB04D11	- oupury resources appressive researing sine for the interesting restrict, partial acqueince
The street the street of the s	0002003	
Uncultured bacterium partial 16S rRNA gene, ck	ine 51903G03	
4	Treese tore	Staphylococcus epidermidis strain NBRC 100911 16S ribosomal RNA gene, partial sequence
Uncultured bacterium partial 16S rRNA gene, ck	ne SB04E04	
Uncultured bacterium partial 16S rRNA gene, clo	me SB03F12	
-		firmicutes 2 leaves

Staphylococcus epidermidis strain 23.1 16S ribosomal RNA gene, partial sequence
6 - The phylogenetic analysis of bacteria isolated from vacuum cleaner dust samples

Bacillus thuringiensis

FJ174596.1

Bacillus thuringiensis strain 104XG46 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb[FJ174596.1]</u> Length: 1049 Number of Matches: 1

Range 1: 59 to 1018 GenBank Graphics Vext Match 🛦 Previous Match							
Score 1651 bit	ts(1830)	Expect 0.0	Identities 944/960(98%)	Gaps 3/960(0%)	Strand Plus/Plus		
Query	1	AGCGGCGGACG	GTGAGTAACACGT	GGTAACCTGCCCA	TAAGACTGGGATAACTCCGGG	60	
Sbjct	59	AGCGGCGGACG	GTGAGTAACACGT	GGTAACCTGCCCA	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	118	
Query	61	AAACCGGGGGCTA	AATACCGGATAATA	TTTGAACTGCATG	GTTCGAAATTGAAAGGCGGCT	120	
Sbjct	119	AAACCGGGGGCTA	AATACCGGATAACAT	TTTGAACTGCATG	GTTCGAAATTGAAAGGCGGCT	178	
Query	121	TCGGCTGTCAC	TTATGGATGGACCCC	GCGTCGCATTAGCT	AGTTGGTGAGGTAACGGCTCA	180	
Sbjct	179	TCGGCTGTCAC	TTATGGATGGACCCO	GTCGCATTAGCT	AGTTGGTGAGGTAACGGCTCA	238	
Query	181	CCAAGGCAACG	ATGCGTAGCCGACCI	GAGAGGGTGATCG	GCCACACTGGGACTGAGACAC	240	
Sbjct	239	CCAAGGCAACG	ATGCGTAGCCGACCI	GAGAGGGGTGATCG	GCCACACTGGGACTGAGACAC	298	
Query	241	GGCCCAGACTC	CTACGGGAGGCAGC	GTAGGGAATCTTC	CGCAATGGACGAAAGTCTGAC	300	
Sbjct	299	GGCCCAGACTC	CTACGGGAGGCAGC	AGTAGGGAATCTTC	CGCAATGGACGAAAGTCTGAC	358	
Query	301	GGAGCAACGCC	GCGTGAGTGATGAAG	GCTTTCGGGTCGT	AAAACTCTGTTGTTAGGGAAG	360	
Sbjct	359	GGAGCAACGCC	GCGTGAGTGATGAAG	GCTTTCGGGTCGT	AAAACTCTGTTGTTAGGGAAG	418	
Query	361	AACAAGTGCTA	GTTGAATAAGCTGGC	ACCTTGACGGTAC	CTAACCAGAAAGCCACGGCTA	420	
Sbjct	419	AACAAGTGCTA	GTTGAATAAGCTGGC	CACCTTGACGGTAC	CTAACCAGAAAGCCACGGCTA	478	
Query	421	ACTACGTGCCA	GCAGCCGCGGTAATZ	CGTAGGTGGCAAG	CGTTATCCGGAATTATTGGGC	480	
Sbjct	479	ACTACGTGCCA	GCAGCCGCGGTAAT	ACGTAGGTGGCAAG	GCGTTATCCGGAATTATTGGGC		
Query	481	GTAAAGCGCGCG	GCAGGTGGTTTCTT	AGTCTGATGTGAA	AGCCCACGGCTCAACCGTGGA	540	
Sbjct	539	GTAAAGCGCGCC	GCAGGTGGTTTCTT	AGTCTGATGTGAA	AGCCCACGGCTCAACCGTGGA	598	
Query	541	GGGTCATTGGA		AGTGCAGAAGAGGA	AAGTGGAATTCCATGTGTAGC	600	
Sbjct	599	GGGTCATTGGA	AACTGGGAGACTTG	AGTGCAGAAGAGGA	AAGTGGAATTCCATGTGTAGC	658	
Query	601	GGTGAAATGCG	TAGAGATATGGAGG	ACACCAGTGGCGA	AGGCGACTTTCTGGTCTGTAA	660	
Sbjct	659	GGTGAAATGCG	TAGAGATATGGAGG	ACACCAGTGGCGA	AGGCGACTTTCTGGTCTGTAA	718	
Query	661	CTGACACTGAG	GCGCGAAAGCGTGG	GAGCAAACAGGAT	TAGATACCCTGGTAGTCCACG	720	
Sbjct	719	CTGACACTGAG	GCGCGAAAGCGTGG	GAGCAAACAGGAT	TAGATACCCTGGTAGTCCACG	778	
Query	721	CCGTAAACGAT	GAGTGCTAAGTGTT	AGAGGGTTTCCGCC	CTTTAGTGCTGAAGTTAACGC	780	
Sbjct	779	CCGTAAACGAT	GAGTGCTAAGTGTT	AGAGGGTTTCCGCC	CTTTAGTGCTGAAGTTAACGC	838	
Query	781	ATTAAGCACTC	CGCCTGGGGGAGTAC	GCCGCAAGGCTGA	AACTCAAGGAATTGACGGGGG	840	
Sbjct	839	ATTAAGCACTC	CGCCTGGGGGAGTAC	GCCGCAAGGCTGA	AACTCAAGGAATTGACGGGGG	898	
Query	841	CCCGCACAANC	GG-GGAGCATGGGG	TTTAATTCAAAGC-	ACGCGAAAACCTTACCAGGTC	898	
Sbjct	899	CCCGCACAAGC	GGTGGAGCATGTGG	TTAATTCGAAGCA	ACGCGAGAACCTTACCAGGTC	958	
Query	899	TTGACTTCCTC	TGAAACCCTANAAAT	TAGGGCTTC-CCTT	CGGGAACAAAAGGACAggggg	957	
Sbjct	959	TTGACATCCTC	TGAAACCCTAGAGA	TAGGGCTTCTCCTT	CGGGAGCAGAGTGACAGGTGG	1018	

	Bacillus thuringiensis clone C05 16S ribosomal RNA gene, partial sequence
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	Bacillas so. ST21 I6S ribosonal RAVA gene, parta sequence addita sector for the sector of t
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	⁴ Bacillus toyonensis strain AEMREG6 16S ribosomal RNA gene, partial sequence
	firmicutes 2 leaves
	Bacillus thuringiensis strain HS18-1, complete genome
	farmicules [2 leaves
	 Bactilus sp. 1434 105 mbosimal RNA gene, partial sequence
	 Baterium Nr14 IOS relosimati RNA gene, partial sequence
	 Destruit NF 8 105 ROSONIA RAVA gene partai sequence Destruit NF 8 105 ROSONIA RAVA gene partai sequence
	Bacilla or 108 in 0500 (16 channel in 1994) Bacilla or 108 in 0500 (16 channel in 1994)
	Bacillus si Island Schwarz (Starberg 1997)
	Lincultured bacterium clone 5 Am 27 16S ribosomal RNA cene, rartial scauence
	Uncultured bacterium clone 5 Am 34 16S ribosomal RNA certe, partial sequence
	Uncultured bacterium clone 5 Am 41 16S ribosonul RNA gene, partial sequence
	Uncultured bacterium clone 5 Am 69 16S ribosonul RNA gene, partial sequence
	Uncultured bacterium clone 5 Am 75 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5_Ne_54 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5_S1_25 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5_51_31165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 5, S1 43 165 ribosomal RNA gene, partial sequence
	9 Bacillas cercus strain LS-8 16S ribosomal RNA gene, partial sequence
	 Bachlos antracis strain IIIS B 15/54 105 rubosinal KNA gene, partial sequence Bachlos e COCU Législand BNA com anciel a serie and a series of the series of
	 Dealing Sp. 100-1105 (Decomain KAV) gene, particular dependence Dealing the functional sector of the function of the
	Bachuss Hamilgeens Main Record State Re
	Bucillus thurineirosis strain K206 16S ribosomal RNA eene, rarial source
	Bacillus sp. NSH-2 165 ribosomal RNA gene, partial sequence
	Bacillus sp. CC6G 16S ribosomal RNA gene, partial sequence
	Bacillus toyonensis strain 265.2 16S ribosomal RNA gene, partial sequence
	Bacillus toyonensis strain 265.1 16S ribosomal RNA gene, partial sequence
	9 Bacillus toyonensis strain 26K.2 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain NCIM 2155 16S ribosomal RNA gene, partial sequence
	Bacillus sp. 31HI-8 ISS ribosomal RNA gene, partial sequence
	 Bactinum Y1M08/16 from China 165 ribosomal KNA gene, partial sequence
	 Bacillus cereirs strain ITB B 10262 ToS rhosomal KNA gene, partial sequence Dealthy divergence IDB (and the second DN) and the second strain second sec
	A backus submignensis tos mosomai KAA gene, partiai sequence A Backus ar TEDTDE 1601 165 abosomai 140 acosomai
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	Bacillus thurinojensis strain 2022 16S ribosomal BNA cone partial source
	Bacillus so. POUC 165 ribosomal RNA cene, narial sequence
	Bacillus cereus strain R12 I6S nitosomal RNA gene, partial source
	Bacillus sp. R12(2014) 16S ribosomal RNA gene, partial sequence
	 Bacillus cereus strain YB18 16S ribosomal RNA gene, partial sequence
	Bacillus sp. enrichment culture clone PRGSd-MS-2 16S ribosonal RNA gene, partial sequence
	Bacillus sp. DB14692 16S nbosonal RNA gene, partial sequence
	Bacillus cereus strain FJM4 16S ribosonal RNA gene, partial sequence
	Bacillus thuringiensis strain FJM3 16S ribosonal RNA gene, partial sequence
	Bacillus thuringiensis strain FJM2 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain JMG-03 16S ribosomal RNA gene, partial sequence
	Bacillus sp. B42(2014) 16S ribosomal RNA gene, partial sequence
	Bacillus sp. B37(2014) 16S ribosomal RNA gene, partial sequence
	Bacillus sp. TD8#11 16S ribosomal RNA gene, partial sequence
	Bacillus mycoides strain YR7 16S ribosomal RNA gene, partial sequence
	Bacillus cereus gene for 16S ribosomal RNA, partial sequence, strain: NRKT
	³ Bacillus cereus strain NXUGD8005 16S ribosomal RNA gene, partial sequence
	 Bacillus cereus strain 6N 165 ribosomal RNA gene, partial sequence
	9 Bacillus thuringiensis strain DL10 165 ribosomal RNA gene, partial sequence
	Cincultured bacterium partial T65 rRNA gene, isolate Be6
	9 Bacillas șp. N5665 partial 165 (RNA gene, strain N5665
	Pacillas thuringiensis strain IHB B 7070 IoS ribosomal RNA gene, partial sequence
	Bacilus cereus strain P1 165 ribosonal RNA gene, partial sequence
	♥ Bacilus thurngiensis strain HU867 165 ribotomal RNA gene, partial sequence Attach to be used and the second
	V Uncultured backrum clone P1 38 165 mbosional KNA gene, partial sequence
1000	g Uncultured backtrum cone (*1_19) tos mosomal KNA gene, partial sequence
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Bacillus mycoides KR088435.1

Bacillus mycoides strain St02 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|KR088435.1</u>] Length: 1398 Number of Matches: 1

Range 1: 396 to 1367 GenBank Graphics Vext Match 🛦 Previous Match						
Score 1703 bits	s(1888)	Expect 0.0	Identities 964/973(99%)	Gaps 3/973(0%)	Strand Plus/Minus	
Query	1	TCTCGTGGTGT	GACGGGCGGTGTGTAC	AAGGCCCGGG	AACGTATTCACCGCGGCATGCTG	60
Sbjct	1367	TCTCGTGGTGT	GACGGGCGGTGTGTAC	CAAGGCCCGGG.	AACGTATTCACCGCGGCATGCTG	1308
Query	61	ATCCGCGATTA	CTAGCGATTCCAGCTI	CATGTAGGCG.	AGTTGCAGCCTACAATCCGAACT	120
Sbjct	1307	ATCCGCGATTA	CTAGCGATTCCAGCT	CATGTAGGCG.	AGTTGCAGCCTACAATCCGAACT	1248
Query	121	GAGAACGGTTT	TATGAGATTAGCTCCA	ACCTCGCGGTC	TTGCAGCTCTTTGTACCGTCCAT	180
Sbjct	1247	GAGAACGGTTT	TATGAGATTAGCTCC	ACCTCGCGGTC	TTGCAGCTCTTTGTACCGTCCAT	1188
Query	181	TGTAGCACGTG	TGTAGCCCAGGTCAT	AGGGGCATGA	TGATTTGACGTCATCCCCACCTT	240
Sbjct	1187	TGTAGCACGTG	TGTAGCCCAGGTCAT	AGGGGCATGA	TGATTTGACGTCATCCCCACCTT	1128
Query	241	CCTCCGGTTTG	TCACCGGCAGTCACCI	TAGAGTGCCC	AACTTAATGATGGCAACTAAGAT	300
Sbjct	1127	CCTCCGGTTTG	TCACCGGCAGTCACC	TAGAGTGCCC	AACTTAATGATGGCAACTAAGAT	1068
Query	301	CAAGGGTTGCG	CTCGTTGCGGGACTT	ACCCAACATC	TCACGACACGAGCTGACGACAAC	360
Sbjct	1067	CAAGGGTTGCG	CTCGTTGCGGGACTT	ACCCAACATC	TCACGACACGAGCTGACGACAAC	1008
Query	361	CATGCACCACC	TGTCACTCTGCCCCC	GAAGGGGAAGC	CCTATCTCTAGGGTTGTCAGAGG	420
Sbjct	1007	CATGCACCACC	TGTCACTCTGCCCCCG	GAAGGGGAAGC	CCTATCTCTAGGGTTGTCAGAGG	948
Query	421	ATGTCAAGACC	TGGTAAGGTTCTTCGC	GTTGCTTCGA	ATTAAACCACATGCTCCACCGCT	480
Sbjct	947	ATGTCAAGACC	TGGTAAGGTTCTTCGC	CGTTGCTTCGA	ATTAAACCACATGCTCCACCGCT	888
Query	481	TGTGCGGGCCC	CCGTCAATTCCTTTG	GTTTCAGCCT	TGCGGCCGTACTCCCCAGGCGGA	540
Sbjct	887	TGTGCGGGCCC	CCGTCAATTCCTTTG	AGTTTCAGCCT	TGCGGCCGTACTCCCCAGGCGGA	828
Query	541	GTGCTTAATGC	GTTAACTTCAGCACTA		ACCCTCTAACACTTAGCACTCAT	600
Sbjct	827	GTGCTTAATGC	GTTAACTTCAGCACTA	AAGGGCGGAA	ACCCTCTAACACTTAGCACTCAT	768
Query	601	CGTTTACGGCG	IGGACTACCAGGGTAI	CTAATCCTGT	ITGCTCCCCACGCTTTCGCGCCT	660
Sbjct	767	CGTTTACGGCG	IGGACTACCAGGGTAT	CTAATCCTGT	TTGCTCCCCACGCTTTCGCGCCT	708
Query	661	CAGTGTCAGTT	ACAGACCAGAAAGTCG	CCTTCGCCAC	IGGTGTTCCTCCATATCTCTACG	720
Sbjct	707	CAGTGTCAGTT	ACAGACCAGAAAGTCG	CCTTCGCCAC	IGGTGTTCCTCCATATCTCTACG	648
Query	721	CATTTCACCGC	TACACATGGAATTCCA	CTTTCCTCTT	CTGCACTCAAGTCTCCCAGTTTC	780
Sbjct	647	CATTTCACCGC	TACACATGGAATTCCA	CTTTCCTCTT	CTGCACTCAAGTCTCCCAGTTTC	588
Query	781	CAATGACCCTC	CACGGTTGAGCCGTGG	GCTTTCACAT	CAAACTTAAAAAACCACCTGCGC	840
Sbjct	587	CAATGACCCTC	CACGGTTGAGCCGTGG	GCTTTCACAT	CAGACTTAAGAAACCACCTGCGC	528
Query	841	GCGCTTTACGC	CCAATAATTCCGGA-A	ACGCTTGCCA	CCTACGTATTACCGCGGCTGCTG	899
Sbjct	527	GCGCTTTACGC	CCAATAATTCCGGATA		CCTACGTATTACCGCGGCTGCTG	468
Query	900	GCACGTAATTA	GCCG-GGCTTTCTGGG	TTAGGTACCG	ICAAGGGGCCAGCTTATTCAACT	958
Sbjct	467	GCACGTAGTTA	GCCGTGGCTTTCT-GG	TTAGGTACCG	ICAAGGTGCCAGCTTATTCAACT	409
Query	959	AACCCTTGTTC	TT 971			
Sbjct	408	AGCACTTGTTC	II TT 396			

alcl Query_11	1989
	Bacillus thuringiensis strain DD287 16S ribosomal RNA gene, partial sequence
	▲ bacteria 2 leaves
v	Bacillus mycoides strain 219298, complete genome
	Bacillus sp. eg1 16S ribosomal RNA gene, partial sequence
	Bacillus mycoides strain 6462 16S ribosomal RNA gene, partial sequence
	Bacillus sp. CSS-7 16S ribosomal RNA gene, partial sequence
	Bacillus samanii strain CI 16S ribosomal RNA gene, partial sequence
	Bacillus pseudomycoides strain R1543 16S ribosomal RNA gene, partial sequence
	Bacillus sp. YXA3-49 16S ribosomal RNA gene, partial sequence
	Bacillus sp. 4828 16S ribosomal RNA gene, partial sequence
	Bacillus pseudomycoides strain XZPGS4 16S ribosomal RNA gene, partial sequence
	firmicutes 5 leaves
	Bacillus sp. SE8 16S ribosomal RNA gene, partial sequence
	Bacillus pseudomycoides strain A1-R33 16S ribosomal RNA gene, partial sequence
	Bacillus pseudomycoides strain JN72 16S ribosomal RNA gene, partial sequence
	Bacillus gaemokensis strain ASR-14 16S ribosomal RNA gene, partial sequence
	Bacillus sp. ZJZF05 16S ribosomal RNA gene, partial sequence
	Bacillus mycoides strain IHB B 6544 16S ribosomal RNA gene, partial sequence
	Bacillus pseudomycoides gene for 16S rRNA, partial sequence, strain: GMA158
	Bacterium NLAE-zI-P444 16S ribosomal RNA gene, partial sequence
	Bacterium FJAT-13831 16S ribosomal RNA gene, partial sequence
	Bacterium MCF45(2011) 16S ribosomal RNA gene, partial sequence
	firmicutes 2 leaves
	firmicutes 71 leaves
8.8896	

Bacillus licheniformis DQ071560.1

Bacillus licheniformis strain MKU 1 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|DQ071560.1</u> Length: 1430 Number of Matches: 1

Range 1:	425 to 13	93 GenBank Graphics		🔻 Next Match 🔺 Previous Match	
Score 1694 bit	s(1878)	Expect Identities 0.0 957/969(Gaps 99%) 0/969(0%)	Strand Plus/Minus	
Query	1	TTACAAACTCTCGTGGTG	IGACGGGCGGTGTGTACA	AGGCCCGGGAACGTATTCACCGCG	60
Sbjct	1393	TTACAAACTCTCGTGGTG		AGGCCCGGGAACGTATTCACCGCG	1334
Query	61	GCATGCTGATCCGCGATT	ACTAGCGATTCCAGCTTC.	ACGCAGTCGAGTTGCAGACTGCGA	120
Sbjct	1333	GCATGCTGATCCGCGATT	ACTAGCGATTCCAGCTTC.	ACGCAGTCGAGTTGCAGACTGCGA	1274
Query	121	TCCGAACTGAGAACAGAT	TTGTGGGATTGGCTTAGC	CTCGCGGCTTCGCTGCCCTTTGTT	180
Sbjct	1273	TCCGAACTGAGAACAGAT	TTGTGGGATTGGCTTAGC	CTCGCGGCTTCGCTGCCCTTTGTT	1214
Query	181	CTGCCCATTGTAGCACGT	GTGTAGCCCAGGTCATAA	GGGGCATGATGATTTGACGTCATC	240
Sbjct	1213	CTGCCCATTGTAGCACGT	GTGTAGCCCAGGTCATAA	GGGGCATGATGATTTGACGTCATC	1154
Query	241	CCCACCTTCCTCCGGTTT	GTCACCGGCAGTCACCTT.	AGAGTGCCCAACTGAATGCTGGCA	300
Sbjct	1153	CCCACCTTCCTCCGGTTT	GTCACCGGCAGTCACCTT.	AGAGTGCCCAACTGAATGCTGGCA	1094
Query	301	ACTAAGATCAAGGGTTGC	GCTCGTTGCGGGACTTAA	CCCAACATCTCACGACACGAGCTG	360
Sbjct	1093	ACTAAGATCAAGGGTTGC	GCTCGTTGCGGGACTTAA	CCCAACATCTCACGACACGAGCTG	1034
Query	361	ACGACAACCATGCACCAC	CTGTCACTCTGCCCCCGA	AGGGGAAGCCCTATCTCTAGGGTT	420
Sbjct	1033	ACGACAACCATGCACCAC	CTGTCACTCTGCCCCCGA	AGGGGAAGCCCTATCTCTAGGGTT	974
Query	421	GTCAGAGGATGTCAAGAC	CTGGTAAGGTTCTTCGCG	TTGCTTCGAATTAAACCACATGCT	480
Sbjct	973	GTCAGAGGATGTCAAGAC	CTGGTAAGGTTCTTCGCG	TTGCTTCGAATTAAACCACATGCT	914
Query	481	CCACCGCTTGTGCGGGCC	CCCGTCAATTCCTTTGAG	TTTCAGTCTTGCGACCGTACTCCC	540
Sbjct	913	CCACCGCTTGTGCGGGCC	CCCGTCAATTCCTTTGAG	TTTCAGTCTTGCGACCGTACTCCC	854
Query	541	CAGGCGGAGTGCTTAATG	CGTTTGCTGCAGCACTAA	AGGGCGGAAACCCTCTAACACTTA	600
Sbjct	853	CAGGCGGAGTGCTTAATG	CGTTTGCTGCAGCACTAA	AGGGCGGAAACCCTCTAACACTTA	794
Query	601	GCACTCATCGTTTACGGC	GTGGACTACCAGGGTATC'	TAATCCTGTTCGCTCCCCACGCTT	660
Sbjct	793	GCACTCATCGTTTACGGC	GTGGACTACCAGGGTATC	TAATCCTGTTCGCTCCCCACGCTT	734
Query	661	TCGCGCCTCAGCGTCAGT	FACAGACCAGAGAGTCGC	CTTCGCCACTGGTGTTCCTCCACA	720
Sbjct	733	TCGCGCCTCAGCGTCAGT	FACAGACCAGAGAGTCGC	CTTCGCCACTGGTGTTCCTCCACA	674
Query	721	TCTCTACGCATTTCACCG	CTACACGTGGAATTCCAC'	TCTCCTCTTCTGCACTCAAGTTCC	780
Sbjct	673	TCTCTACGCATTTCACCG	CTACACGTGGAATTCCAC	TCTCCTCTTCTGCACTCAAGTTCC	614
Query	781	CCAGTTTCCAATGACCCT	CCCCGGTTGAGCCGGGGG	CTTTCACATCAAACTTAAAAAACC	840
Sbjct	613	CCAGTTTCCAATGACCCT	CCCCGGTTGAGCCGGGGGG	CTTTCACATCAAACTTAAAAAACC	554
Query	841	GCCTGCGCGCGCGCTTTACCO	CCCAATAATTCCCGGAAA	CCCTTGCCACCTACGTATTACCGC	900
Sbjct	553	GCCTGCGCGCGCTTTACG	CCCAATAATTCCGGACAA	CGCTTGCCACCTACGTATTACCGC	494
Query	901	GGCTGCTGGGACGTAATT		AGGTACCGTCAAGGAACCCCCCTA	960
Sbjct	493	GGCTGCTGGCACGTAGTT	AGCCGTGGCTTTCTGGTT	AGGTACCGTCAAGGTACCGCCCTA	434
Query	961	TTCAAAAGG 969			
Sbjct	433	TTCGAACGG 425			

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			Bacillus licheniformis strain C1-5-8 16S ribosomal RNA gene, partial sequence

Bacillus subtilis KF220577.1

Bacillus subtilis strain YLB-P1 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KF220577.1</u>| Length: 1050 Number of Matches: 1

Range 1: 51 to 1025 GenBank Graphics Vext Match 🛦 Previous Match					Match		
Score 1653 bits	s(1832)	Expect 0.0	Identities 955/975(98%)	Gaps 5/975(0%)	Strand Plus/Plus		
Query	1	CTGATGTTAGCG	GCGGACGGGTG	AGTAACACGTGGG	TAACCTGCCTGTAAGA	ACTGGGATA	60
Sbjct	51	CTGATGTTAGCG	GCGGACGGGTG	AGTAACACGTGGG		ACTGGGATA	110
Query	61	ACTCCGGGAAAC	CGGGGGCTAATA	CCGGATGGTTGTT	GAACCGCATGGTTC	AACATAAA	120
Sbjct	111	ACTCCGGGAAAC	CGGGGGCTAATA	CCGGATGGTTGTT	GAACCGCATGGTTC	AACATAAA	170
Query	121	AGGTGGCTTYGG	CTACCACTTAC	AGATGGACCCGCGC	CGCATTAGCTAGTT	GTGAGGTA	180
Sbjct	171	AGGTGGCTTCGG	GTACCACTTAC	AGATGGACCCGCGC	GCGCATTAGCTAGTT	GTGAGGTA	230
Query	181	ACGGCTCACCAA	GGCAACGATGC	GTAGCCGACCTGAC	AGGGTGATCGGCCAC	CACTGGGAC	240
Sbjct	231	ACGGCTCACCAA	GGCAACGATGC	GTAGCCGACCTGAC	GAGGGTGATCGGCCAC	ACTGGGAC	290
Query	241	TGAGACACGGCC	CAGACTCCTAC	GGGAGGCAGCAGTA	AGGGAATCTTCCGCAZ	TGGACGAA	300
Sbjct	291	TGAGACACGGCC	CAGACTCCTAC	GGGAGGCAGCAGTZ	AGGGAATCTTCCGCA	ATGGACGAA	350
Query	301	AGTCTGACGGAG	CAACGCCGCGT	GAGTGATGAAGGT	TTCGGATCGTAAAGO	TCTGTTGT	360
Sbjct	351	AGTCTGACGGAG	CAACGCCGCGT	GAGTGATGAAGGT	TTCGGATCGTAAAGO	TCTGTTGT	410
Query	361	TAGGGAAGAACA	AGTACCGTTCG	AATAGGGCGGTAC	TTGACGGTACCTAAC	CAGAAAGC	420
Sbjct	411	TAGGGAAGAACA	AGTACCGTTCG	AATAGGGCGGTACO	CTTGACGGTACCTAAC	CAGAAAGC	470
Query	421	CACGGCTAACTA	CGTGCCAGCAG	CCGCGGTAATACG	AGGTGGCAAGCGTT	STCCGGAAT	480
Sbjct	471	CACGGCTAACTA	CGTGCCAGCAG	CCGCGGTAATACG	TAGGTGGCAAGCGTTG	STCCGGAAT	530
Query	481	TATTGGGCGTAA			CTGATGTGAAAGCCC	CCGGCTCA	540
Sbjct	531	TATTGGGCGTAA	AGGGCTCGCAG	GCGGTTTCTTAAG	CTGATGTGAAAGCCC	CCGGCTCA	590
Query	541	ACCGGGGGAGGG	CATTGGAAACT	GGGGAACTTGAGT	GCAGAAGAGGAGAGT	GAATTCCA	600
Sbjct	591	ACCGGGGAGGGI	CATTGGAAACT	GGGGAACTTGAGT	GCAGAAGAGGAGAGT	GAATTCCA	650
Query	601	CGTGTAGCGGT	SAAATGCGTAGA	GATGTGGAGGAAC	ACCAGTGGCGAAGGC	ACTCTCTG	660
Sbjct	651	CGTGTAGCGGTG	GAAATGCGTAGA	GATGTGGAGGAAC	ACCAGTGGCGAAGGCC	ACTCTCTG	710
Query	661	GTCTGTAACTGA	ACGCTGAGGAGC	GAAAGCGTGGGGA	GCGAACAGGATTAGAT	TACCCTGGT	720
Sbjct	711	GTCTGTAACTGA	ACGCTGAGGAGC	GAAAGCGTGGGGA	GCGAACAGGATTAGAT	TACCCTGGT	770
Query	721	AGTCCACGCCGI	TAAACGATGAGT	GCTAAGTGTTAGG	GGTTTCCGCCCCTTZ	AGTGCTGCA	780
Sbjct	771	AGTCCACGCCGI	TAAACGATGAGT	GCTAAGTGTTAGG	GGTTTCCGCCCCTT	AGTGCTGCA	830
Query	781	GCTAACGCATTA	AGCACTCCGCC	TGGGGAGTACGGT	CGCAAGACTGAAACTC	CAAAGGAAT	840
Sbjct	831	GCTAACGCATTA	AGCACTCCGCC	TGGGGAGTACGGT	CGCAAGACTGAAACTC	CAAAGGAAT	890
Query	841	TCACGGGGGGCCC	GCACAAGCGGG	GGAGCATGTGATT	-ATTCGAAGC-ACGO	GAAAACCT	898
Sbjct	891	TGACGGGGGGCCC	GCACAAGCGGT	GGAGCATGTGGTT	TAATTCGAAGCAACGO	CGAAGACCT	950
Query	899	TACCA-GTCTTC	GACTTCCTCTGA	GATCCTaaaaaaa	GACGTCCCCTTC-G	GCAAAAGG	956
Sbjct	951	TACCAGGTCTTG	GACATCCTCTGA	CATCCTAGAGATA	GACGTCCCCTTCGG	GCCAAGTG	1010
Query	957	ACAG-GGGGGGAI	rggt 970				
Sbjct	1011	ACAGTGGTGCAI	r <mark>GGT 1025</mark>				

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	Uncultured Bacillus sp. clone PU 12 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain jk-45 16S ribosomal RNA gene, partial sequence
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	Bacillus sp. LS-036 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain CH16 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain IC3 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain Bp-1 16S ribosonal RNA, partial sequence
	Bacillus sp. ZLXH-2 16S ribosomal RNA gene, partial sequence
	Bacillus sp. <i>LL</i> AI-to IOS ribosonal KNA gene, partial sequence
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	Bacillus n. CEB332. 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain W6 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis subsp. inaquosorum strain B373 16S ribosomal RNA gene, partial sequence
	🖗 Bacillus subtilis subsp. inaquosorum strain MRS3 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain DBT13 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain F-24 16S ribosomal RNA gene, partial sequence
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	Bacillus subtilis strain SRCF17 165 ribosomal RNA gene, partial sequence
	Bacillus sp. KSRH34 partial 16S rRNA gene, strain KSRH34
	Bacillus vallismortis strain DD007 16S ribosomal RNA gene, partial sequence
	BAB-4886 16S ribosomal RNA gene, partial sequence
	Bacillus sp. BAB-5142 16S ribosomal RNA gene, partial sequence
	Bacillus subilis strain F-36 165 ribosomal RNA gene, partial sequence
	Bacillus surdins strain F-20 105 ribosomai KNA gene, partai sequence
	Bacillus spit bits ninacuscommistrain R111165 tiposonal RNA cene, partial sequence Bacillus subtilis subson inacuscommistrain R111165 tiposonal RNA cene, partial sequence
	Bacillus sp. JES1 16S ribosomal RNA gene, partial sequence
0.001	Bacillus subtilis subsp. subtilis strain DSM 10 16S ribosomal RNA gene, partial sequence
	Bacterium YC-LK-LKJ121 16S ribosomal RNA gene, partial sequence
	Bacterium YC-LK-LKJ43 16S ribosomal RNA gene, partial sequence
	firmicutes 6 leaves
	Bacillus subtilis strain D1 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain DD156 16S ribosomal RNA gene, partial sequence
	Bacillus sp. CanL-1 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain WMA-LM15 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain F-27 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain BKDS1 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis subsp. inaquosorum strain MRS11 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis subsp. inaquosorum strain B381 16S ribosomal RNA gene, partial sequence
	Bacillus sp. VRT4 16S ribosomal RNA gene, partial sequence
	Bacillus subtilis strain SANN3 16S ribosomal RNA gene, partial sequence
	Bacterium YC-LK-LKJ44 16S ribosomal RNA gene, partial sequence
	Bacterium Y2 16S ribosomal RNA gene, partial sequence
	firmicutes 6 leaves
	Bacillus subtilis strain S6CF30 16S ribosomal RNA gene, partial sequence

7-The phylogenetic analysis of bacteria isolated from sole of shoes samples

Echireshia coli LN558643.1

Escherichia coli partial 16S rRNA gene, isolate RS1 Sequence ID: <u>emb[LN558643.1]</u> Length: 1484 Number of Matches: 1

Range 1: 87 to 1069 GenBank Graphics V Next Match 🔺 Previous Match						
Score 1710 bits	s(1896)	Expect 0.0	Identities 972/983(99%)	Gaps 5/983(0%)	Strand Plus/Plus	
Query	1	AACCCACTCCCA	TGGTGTGACGGGC	GGTGTGTACAAG	GCCCGGGAACGTATTCACCGTGG	60
Sbjct	87	AACCCACTCCCA	TGGTGTGACGGGC	 GGTGTGTACAAG		146
Query	61	CATTCTGATCCA	CGATTACTAGCGA	TTCCGACTTCAT	GGAGTCGAGTTGCAGACTCCAAT	120
Sbjct	147	CATTCTGATCCA	CGATTACTAGCGA	TTCCGACTTCAT	GGAGTCGAGTTGCAGACTCCAAT	206
Query	121	CCGGACTACGAC	GCACTTTATGAGG	TCCGCTTGCTCT	CGCGAGGTCGCTTCTCTTTGTAT	180
Sbjct	207	CCGGACTACGAC	GCACTTTATGAGG	TCCGCTTGCTCT	CGCGAGGTCGCTTCTCTTTGTAT	266
Query	181	GCGCCATTGTAG	CACGTGTGTAGCC	CTGGTCGTAAGG	GCCATGATGACTTGACGTCATCC	240
Sbjct	267	GCGCCATTGTAG	CACGTGTGTAGCC	CTGGTCGTAAGG	GCCATGATGACTTGACGTCATCC	326
Query	241	CCACCTTCCTCC	AGTTTATCACTGG	CAGTCTCCTTTG	AGTTCCCGGCCGGACCGCTGGCA	300
Sbjct	327	CCACCTTCCTCC	AGTTTATCACTGG	CAGTCTCCTTTG	AGTTCCCGGCCGGACCGCTGGCA	386
Query	301	ACAAAGGATAAG	GGTTGCGCTCGTT	GCGGGACTTAAC	CCAACATTTCACAACACGAGCTG	360
Sbjct	387	ACAAAGGATAAG	GGTTGCGCTCGTT	GCGGGACTTAAC	CCAACATTTCACAACACGAGCTG	446
Query	361	ACGACAGCCATG	CAGCACCTGTCTC	ACAGTTCCCGAA	GGCACCAATCCATCTCTGGAAAG	420
Sbjct	447	ACGACAGCCATG	CAGCACCTGTCTC	ACAGTTCCCGAA	GGCACCAATCCATCTCTGGAAAG	506
Query	421	TTCTGTGGATGT	CAAGACCAGGTAA	GGTTCTTCGCGT	TGCATCGAATTAAACCACATGCT	480
Sbjct	507	TTCTGTGGATGT	CAAGACCAGGTAA	GGTTCTTCGCGT	TGCATCGAATTAAACCACATGCT	566
Query	481	CCACCGCTTGTG	CGGGCCCCCGTCA	ATTCATTTGAGT	TTTAACCTTGCGGCCGTACTCCC	540
Sbjct	567	CCACCGCTTGTG	CGGGCCCCCGTCA	ATTCATTTGAGT	TTTAACCTTGCGGCCGTACTCCC	626
Query	541	CAGGCGGTCGAC	TTAACGCGTTAGC	ICCGGAAGCCAC	GCCTCAAGGGCACAACCTCCAAG	600
Sbjct	627	CAGGCGGTCGAC	TTAACGCGTTAGC	ICCGGAAGCCAC	GCCTCAAGGGCACAACCTCCAAG	686
Query	601	TCGACATCGTTT	ACGGCGTGGACTA	CCAGGGTATCTA	ATCCTGTTTGCTCCCCACGCTTT	660
Sbjct	687	TCGACATCGTTT	ACGGCGTGGACTA	CCAGGGTATCTA	ATCCTGTTTGCTCCCCACGCTTT	746
Query	661	CGCACCTGAGCG	TCAGTCTTCGTCC	AGGGGGCCGCCT	TCGCCACCGGTATTCCTCCAGAT	720
Sbjct	747	CGCACCTGAGCG	TCAGTCTTCGTCC	AGGGGGGCCGCCT	TCGCCACCGGTATTCCTCCAGAT	806
Query	721	CTCTACGCATTT	CACCGCTACACCT	GGAATTCTACCC	CCCTCTACGAGACTCAAGCTTGC	780
Sbjct	807	CTCTACGCATTT	CACCGCTACACCT	GGAATTCTACCC	CCCTCTACGAGACTCAAGCTTGC	866
Query	781	CAGTATCAGATG	CAGTTCCCAGGTT	GAGCCCGGGGAT	TTCACATCTGACTTAACAAACCG	840
Sbjct	867	CAGTATCAGATG	CAGTTCCCAGGTT	GAGCCCGGGGAT	TTCACATCTGACTTAACAAACCG	926
Query	841	CCTGCGTGCGCT	TTACGCCCAGTAA	TTCCGATTAACG	CTTGC-CCCTCCGTATTACCGCG	899
Sbjct	927	CCTGCGTGCGCT	TTACGCCCAGTAA	TTCCGATTAACG	CTTGCACCCTCCGTATTACCGCG	986
Query	900	GCTGCTGGCACG	GAATTAGCCGGGG	CTTC-TCTGGGG	GTAACGTCAATGAAC-AAGGTAT	957
Sbjct	987	GCTGCTGGCACG	GAGTTAGCCGGTG	CTTCTTCTGCGG	GTAACGTCAATGAACAAAGGTAT	1046
Query	958	TACTTACTCC	CTTccccccc	978		
Sbjct	1047	TAACTTTACTCC	CTTCCTTCCCC	1069		

	Productive and second
	² Escherichia coli strain MAC 5 165 ribosomal KNA gene, partial sequence
	Escherichia coli strain XJALT-127-1YF3 16S ribosomal RNA gene, partial sequence
	Escherichia coli strain KLU01 16S ribosomal RNA gene, partial sequence
	enterohacteria 2 leaves
	Escherichia coli APEC 078, complete genome
	La de la contra contra e constructiva de la contra de la contr
	Escherichia coli O157:H7 str. Sakai strain Sakai 16S ribosomal RNA, complete sequence
	Shigella sonnei Ss046 strain Ss046 16S ribosomal RNA, complete sequence
	Escherichia sp. 26N 16S ribosomal RNA gene, partial sequence
	antaroha ctaria 11 Januar
	Escherichia coli strain BAB-538 168 ribosomal KNA gene, partial sequence
	Escherichia sp. XJ134-1212-NF2 16S ribosomal RNA gene, partial sequence
	Escherichia coli Q157:H7 str SS17, complete genome
	Chicalle davassi structure (ASDS) (65 sibussing IDNA sana partis) esquance
	singena nexten suan russons tos nosonar Riva gene, partar sequence
	Escherichia coli O157:H7 str. EDL953, complete genome
	Shigella sonnei strain PS 69 16S ribosomal RNA gene, partial sequence
	Escherichia sp. UIWRE1299 16S ribosomal RNA gene partial sequence
	Eschericking 3D. Office 1227 (05 flows) and Kirk gene, partial sequence
	Escherichia sp. UTWRF1140-16S ribosomai RNA gene, partial sequence
	Escherichia sp. UIWRF1138 16S ribosomal RNA gene, partial sequence
	Escherichia sp. UIWRF0963 16S ribosomal RNA gene, partial sequence
	Echarichi soli PCN/061 complete consume (gene partie) sequence
	Tscherkna con Fervori, comprete genome
	Escherichia coli PUN053, complete genome
	Escherichia coli strain CFSAN029787, complete genome
	Escherichia coli strain CI5, complete genome
	Use the set of a state of the set
	Uncultured Escherichia sp. clone RL-9 105 ribosonial RNA gene, partial sequence
	Escherichia coli strain BL21 (TaKaRa), complete genome
	Escherichia coli strain C43(DE3), complete genome
	a Uncultured organism clone FLU0043-T225-S-NIPCRAMeANa (00008 small subunit ribosomal RNA eene partial sequence
	Cheminer of gammer of the theory of the second of the seco
	Escherichia coli partial 16S rRNA gene, isolate RS1
	Escherichia sp. UIWRF0339 16S ribosomal RNA gene, partial sequence
	Echerichia sp. UIWRE0398 16S ribosomal RNA gene, partial sequence
	Lastrikina sp. of two loss to strosoma kata gan, panar septence
	Escherichia sp. UTWRF0413 ToS ribosomal RNA gene, partial sequence
	Escherichia sp. UIWRF0464 16S ribosomal RNA gene, partial sequence
	Escherichia sp. UIWRF0682 16S ribosomal RNA gene, partial sequence
	Echarichia sp. UIWPEIS22 ISS shoomal DNA ana pastial carumana
	Tscherkna sp. Ofw.RF0.22 (05 h0050ma RVA gene, partar sequence
	Escherichia sp. UIWRF0606 16S ribosomal RNA gene, partial sequence
	Escherichia sp. UIWRF0837 16S ribosomal RNA gene, partial sequence
	Excherichia sp. UIWRE0021 16S ribosomal RNA gene partial sequence
	Eschericking JJ, Office Office Statement BNA series partial sequence
	Esciencia sp. 01w RF0920-105 noosomai RVA gene, partial sequence
	Escherichia sp. UIWRF0927 16S ribosomal RNA gene, partial sequence
	Escherichia sp. UIWRF0934 16S ribosomal RNA gene, partial sequence
	Excherichia sp. UIWRE0045.16S ribosomal RNA gene, partial sequence
	Escherchina Sp. Of WKI 0745 TOS HOUSSMAN KVA gene, partian sequence
0.0008	Escherichia sp. UTWRF0946-165 ribosomai RNA gene, partial sequence
	Escherichia sp. UIWRF0399 16S ribosomal RNA gene, partial sequence
	Shirefua souther strain FN17 ToN tubosonial RNA gene narrial sequence
	sing state state i stat
	Escherichia coli ACNUU, complete genome
	Escherichia coli strain FM 37 16S ribosomal RNA gene nartial sequence
	Escherichia coli strain OZKT 165 ribosomai RNA gene, partial sequence
	Escherichia sp. S488 16S ribosomal RNA gene, partial sequence
	Escherichia sp. 555 165 ribosomai KNA gene, partial sequence
	Escherichia coli genome assembly FHI98, scaffold scaffold-29 contig-2.0.1 485219 forganism: Escherichia
	Exhaustic and a second s
	Escherichia coli genome assembly Prilos, scattoid scattoid-26_contig-0.1_87205_868516_[organism:Escherichia
	Escherichia coli genome assembly FHI75, scaffold scaffold-22 contig-4.0 1 444658 [organism: Escherichia
	Escharishing and a second his EUITS, coeffeld coeffeld 26 courts 0.1 1028, 1216600, Language Escharishing
	Escherichia con genome assembly Pri 75, scarloid scarloid-20_contig-0.1_1028_1510009_[organism: Escherichia
	Escherichia coli genome assembly FHI79, scaffold scaffold-12 contig-17.0 1 37303 [organism:Escherichia
	Eastharishin and an anomaly EUISO coeffold an ffold 18 control 2.0 1 672021 Langenian Eastharishin
	escherichia con genome assenioly Prila9, scariold scariold-18_config-5.0_1_675951_forganism: escherichia
	Escherichia coli genome assembly FHI92, scaffold scaffold-16 contig-8.1 144654 238240 [organism: Escherichia
	Escharichia agli agnoma assambly EU107, sonffold anoffold 46, agnitia 0.4, 143201, 875885, Jonaniam Escharichia
	escherichia con genome assentory PHP7, scartoid scartoid -40_contrg-0.4_145201_875885_[organism: Escherichia
	Shigella sonnei strain R5 16S ribosomal RNA gene, partial sequence
	Excharichia coli strain P0 16S ribosomal PNA paga partial sequence
	Lachterin der statil (2) (10) filosonia (20) gele, partai seigende
	Bacterium EM-2014-18 genomic DNA containing 16S-23S intergenic spacer region, isolate 18
	Bacterium EM-2014-19 genomic DNA containing 16S-23S intergenic spacer region, isolate 19
	Bacterium EM-2014-20 genomic DNA containing 165-255 intergenic spacer region, isolate 20
	Bacterium EM-2014-30 genomic DNA containing 16S-23S intergenic spacer region, isolate 30
	Destering EM 2014 (1) even is DNA contribute 1/6 225 internet point index (1)
	Bacterium EM-2014-41 genomic DNA containing 105-255 intergenic spacer region, isolate 41
	Bacterium EM-2014-44 genomic DNA containing 16S-23S intergenic spacer region, isolate 44
	Partonium EM 2014 48 ganomia DNA containing 165 235 intergandia magazi ragion isolata 48
	baceruni Est-2014-46 genomic EXXX containing 105-255 intergence spacer region, isolate 46
	Bacterium EM-2014-70 genomic DNA containing 16S-23S intergenic spacer region, isolate 70
	Bactarium FM-2014-76 panomic DNA containing 16S-23S intergenic spacer ragion isolate 76
	bacertain Ext-2014-70 genome Diver containing 103-253 intergence spacer region, isolate 70
	Bacterium EM-2014-112 genomic DNA containing 16S-23S intergenic spacer region, isolate 112
	Bacterium FM-2014-114 genomic DNA containing 16S-23S intergenic spacer region isolate 114
	Bartelan EM 2014 [2] second EM second and 10 200 integration space (region, isolate [1])
	bacterium ENF2014-121 genomic DNA containing 16S-23S intergenic spacer region, isolate 121
	Bacterium EM-2014-125 genomic DNA containing 16S-23S intergenic spacer region isolate 125
	Exception EX 2014 122 seconds DN14 containing 165 225 interprets speet region, sound 122
	Bacterium EM-2014-132 genomic DNA containing 16S-23S intergenic spacer region, isolate 132
	Bacterium EM-2014-134 genomic DNA containing 16S-23S intergenic spacer region isolate 134
	Barden and Evidence and a second
	Bacterium EM-2014-144 genomic DNA containing 168-238 intergenic spacer region, isolate 144
	Bacterium EM-2014-153 genomic DNA containing 16S-23S intergenic spacer region isolate 153
	Detection EM 2014 165 established bits of controlling 162 225 intergence entrolling include 125
	Bacterium EM-2014-165 genomic DNA containing 168-238 intergenic spacer region, isolate 165
	Bacterium EM-2014-166 genomic DNA containing 16S-23S intergenic spacer region, isolate 166
	Pastarium FM 2014 181 aanomia DNA containina 16S 23S internania spacer ragion, isolata 181
	bacertuin Executivitat genomic DAVA containing 105-255 intergenic spacer region, isolate 181
	Bacterium EM-2014-183 genomic DNA containing 16S-23S intergenic spacer region, isolate 183
	enterobacteria 8 leaves
	enterobacteria 8 leaves

0.0008

Brevibacillus borstelensis KT239000.1

Brevibacillus borstelensis strain HMR16 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|KT239000.1|</u> Length: 1408 Number of Matches: 1

Range 1: 46 to 1030 GenBank Graphics Vext Match 🔺 Previous Match						
Score 1676 bits	s(1858)	Expect 0.0	Identities 966/985(98%)	Gaps 4/985(0%)	Strand Plus/Plus	
Query	1	GAGTAACACGTA	GGCAACCTGCCCGT	AAGCTCGGGATAA	CATGGGGAAACTCATGCTAAT	60
Sbjct	46	GAGTAACACGTA	.GGCAACCTGCCCGT	AAGCTCGGGATAA	CATGGGGAAACTCATGCTAAT	105
Query	61	ACCGGATAGGGT	CTTCTCTCGCATGA	GAGGAGACGGAAA	GGTGGCGCAAGCTACCACTTA	120
Sbjct	106	ACCGGATAGGGT	CTTCTCTCGCATGA	GAGGAGACGGAAA	GGTGGCGCAAGCTACCACTTA	165
Query	121	CGGATGGGCCTG	CGGCGCATTAGCTA	GTTGGTGGGGTAA	CGGCCTACCAAGGCGACGATG	180
Sbjct	166	CGGATGGGCCTG	CGGCGCATTAGCTA	GTTGGTGGGGTAA	CGGCCTACCAAGGCGACGATG	225
Query	181	CGTAGCCGACCT	GAGAGGGTGACCGG	CCACACTGGGACT	GAGACACGGCCCAGACTCCTA	240
Sbjct	226	CGTAGCCGACCT	GAGAGGGTGACCGG	CCACACTGGGACT	GAGACACGGCCCAGACTCCTA	285
Query	241	CGGGAGGCAGCA	GTAGGGAATTTTCC		GTCTGATGGAGCAACGCCGCG	300
Sbjct	286	CGGGAGGCAGCA	GTAGGGAATTTTCC	ACAATGGACGAAA	GTCTGATGGAGCAACGCCGCG	345
Query	301	TGAACGATGAAG	GTCTTCGGATTGTA	AAGTTCTGTTGTC	AGAGACGAACAAGTACCGTTC	360
Sbjct	346	TGAACGATGAAG	GTCTTCGGATTGTA	AAGTTCTGTTGTC	AGAGACGAACAAGTACCGTTC	405
Query	361	GAACAGGGCGGT	ACCTTGACGGTACC	rgacgagaaagcc	ACGGCTAACTACGTGCCAGCA	420
Sbjct	406	GAACAGGGCGGT	ACCTTGACGGTACC	IGACGAGAAAGCC	ACGGCTAACTACGTGCCAGCA	465
Query	421	GCCGCGGTAATA	CGTAGGTGGCAAGC	GTTGTCCGGAATT	ATTGGGCGTAAAGCGCGCGCA	480
Sbjct	466	GCCGCGGTAATA	CGTAGGTGGCAAGC	GTTGTCCGGAATT	ATTGGGCGTAAAGCGCGCGCA	525
Query	481	GGCGGCTATGTA		GCCCGGGGCTCAA	CCCCGGTTCGCATCGGAAACT	540
Sbjct	526	GGCGGCTATGTA	AGTCTGGTGTTAAA	GCCCGGGGCTCAA	CCCCGGTTCGCATCGGAAACT	585
Query	541	GTGTAGCTTGAG	TGCAGAAGAGGAAA	GCGGTATTCCACG	TGTAGCGGTGAAATGCGTAGA	600
Sbjct	586	GTGTAGCTTGAG	TGCAGAAGAGGAAA	GCGGTATTCCACG	TGTAGCGGTGAAATGCGTAGA	645
Query	601	GATGTGGAGGAA	CACCAGTGGCGAAG	GCGGCTTTCTGGT	CTGTAACTGACGCTGAGGCGC	660
Sbjct	646	GATGTGGAGGAA	CACCAGTGGCGAAG	GCGGCTTTCTGGT	CTGTAACTGACGCTGAGGCGC	705
Query	661	GAAAGCGTGGGG	AGCAAACAGGATTA	GATACCCTGGTAG	TCCACGCCGTAAACGATGAGT	720
Sbjct	706	GAAAGCGTGGGG	AGCAAACAGGATTA	GATACCCTGGTAG	TCCACGCCGTAAACGATGAGT	765
Query	721	GCTAGGTGTTGG	GGGTTTCAATACCC	ICAGTGCCGCAGC	TAACGCAATAAGCACTCCGCC	780
Sbjct	766	GCTAGGTGTTGG	GGGTTTCAATACCC:	ICAGTGCCGCAGC	TAACGCAATAAGCACTCCGCC	825
Query	781	TGGGGAGTACGC	TCGCAAGAGTGAAA	CTCAAAGGATTTG.	ACGGGGGGCCCGCACAAGCGG-	839
Sbjct	826	TGGGGAGTACGC	TCGCAAGAGTGAAA	CTCAAAGGAATTG	ACGGGGGGCCCGCACAAGCGGT	885
Query	840	GGAGCATGTGGT	TTAATTCAAAGCAA	CGCGAAAAACCTT.	ACCA-GTCTTGACATCCCGCT	898
Sbjct	886	GGAGCATGTGGT	TTAATTCGAAGCAAG	CGCGAAGAACCTT	ACCAGGTCTTGACATCCCGCT	945
Query	899	GACCGTCCTaaa	aaaaGGGCTTCCCT		GACgggggggggGCATGGTTGTC	956
Sbjct	946	GACCGTCCTAGA	GATAGGGCTTCCCT	ICGGGGCAGCGGT	GACAGGTGGTGCATGGTTGTC	1005
Query	957	GTCCCCTCGGGT	CGGGAAATGTGGG	981		
Sbjct	1006	GTCAGCTCGTGT	CGTGAGATGTTGG	1030		



Bacillus licheniformis KP772335.1

Bacillus licheniformis strain OALB2 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|KP772335.1</u>| Length: 1445 Number of Matches: 1

Range 1:	45 to 98	39 GenBank	<u>Graphics</u>			V Next I	Match 🔺 Pre	evious Match	
Score 1645 bit	s(1824)		Expect 0.0	Identities 935/945(99%)	Gaps 4/945	5(0%)	Strand Plus/Plus		
Query	1	GGCGGI	ACGGGTGA	GTAACACGTGGG	TAACCTGCC	TGTAAGAC	TGGGATA	ACTCCGGGAAA	60
Sbjct	45	GGCGGZ	 ACGGGTGA	 GTAACACGTGGG	TAACCTGCC	TGTAAGAC	TGGGATA	 ACTCCGGGAAA	104
Query	61	CCGGGG	CTAATAC	CGGATGCTTGAT	TGAACCGCA	TGGTTCAA	TTATAAA	AGGTGGCTTTC	120
Sbjct	105	CCGGGG	CTAATAC	CGGATGCTTGAI	TGAACCGCA	ATGGTTCAA	TTATAAA.	AGGTGGCTTTC	164
Query	121	AGCTAC	CACTTGC	AGATGGACCCGC	GGCGCATT	AGCTAGTTO	GTGAGGT	AACGGCTCACC	180
Sbjct	165	AGCTAC	CACTTGC	AGATGGACCCGC	GGCGCATTA	GCTAGTTG	GTGAGGT	AACGGCTCACC	224
Query	181		ACGATGC	GTAGCCGACCTG	AGAGGGTG	TCGGCCAC	ACTGGGA	CTGAGACACGG	240
Sbjct	225	AAGGCO	SACGATGC	GTAGCCGACCTO	AGAGGGTGA	TCGGCCAC	ACTGGGA	.CTGAGACACGG	284
Query	241	CCCAG	ACTCCTAC	GGGAGGCAGCAG	TAGGGAAT	TTCCGCAA	TGGACGA	AAGTCTGACGG	300
Sbjct	285	CCCAG	ACTCCTAC	GGGAGGCAGCAG	TAGGGAATO	TTCCGCAA	TGGACGA	AAGTCTGACGG	344
Query	301	AGCAAC	GCCGCGT	GAGTGATGAAGG	TTTTCGGAI	CGTAAAAC	TCTGTTG	TTAGGGAAGAA	360
Sbjct	345	AGCAAC	GCCGCGT	GAGTGATGAAGG	TTTTCGGAI	CGTAAAAC	TCTGTTG	TTAGGGAAGAA	404
Query	361	CAAGT	ACCGTTCG	AATAGGGCGGTA	CCTTGACGO	TACCTAAC	CAGAAAG	CCACGGCTAAC	420
Sbjct	405	CAAGT	ACCGTTCG	AATAGGGCGGTA	CCTTGACGO	TACCTAAC	CAGAAAG	CCACGGCTAAC	464
Query	421	TACGTO	CCAGCAG	CCGCGGTAATAC	GTAGGTGGC	CAAGCGTTG	TCCGGAA	TTATTGGGCGT	480
Sbjct	465	TACGTO	GCCAGCAG	CCGCGGTAATAC	GTAGGTGGC	CAAGCGTTG	TCCGGAA	TTATTGGGCGT.	524
Query	481	AAAGCO	CGCGCAG	GCGGTTTCTTAA	GTCTGATG	GAAAGCCC	CCGGCTC	AACCGGGGAGG	540
Sbjct	525	AAAGCO	GCGCGCAG	GCGGTTTCTTAA	GTCTGATG	GAAAGCCC	CCGGCTC	AACCGGGGAGG	584
Query	541	GTCATI	GGAAACT	GGGGAACTTGAG	TGCAGAAGA	GGAGAGTG	GAATTCC	ACGTGTAGCGG	600
Sbjct	585	GTCATI	GGAAACT	GGGGAACTTGAG	TGCAGAAGA	GGAGAGTG	GAATTCC	ACGTGTAGCGG	644
Query	601	TGAAAT	GCGTAGA	GATGTGGAGGAA	CACCAGTGG	CGAAGGCG	ACTCTCT	GGTCTGTAACT	<mark>66</mark> 0
Sbjct	645	TGAAAI	GCGTAGA	GATGTGGAGGAA	CACCAGTGO	CGAAGGCG	ACTCTCT	GGTCTGTAACT	704
Query	661	GACGCI	GAGGCGC	GAAAGCGTGGGG	AGCGAACAG	GATTAGAT	ACCCTGG	TAGTCCACGCC	720
Sbjct	705	GACGCI	GAGGCGC	GAAAGCGTGGGG	AGCGAACAG	GATTAGAT	ACCCTGG	TAGTCCACGCC	764
Query	721	GTAAAC	GATGAGT	GCTAAGTGTTAA	AGGGTTTCC	GCCCTTTA	GTGCTGC	AGCAAACGCAT	780
Sbjct	765	GTAAAC	GATGAGT	GCTAAGTGTTAG	AGGGTTTCC	GCCCTTTA	GTGCTGC	AGCAAACGCAT	824
Query	781	TAAGCA	CTCCGCC	TGGGGAGTACGG	TCGCAAGAC	TGAAACTC	-AAGGAA	TTGACGGGGGGC	839
Sbjct	825	TAAGCA	CTCCGCC	TGGGGAGTACGG	TCGCAAGAC	TGAAACTC	AAAGGAA	TTGACGGGGGC	884
Query	840	CCGCCC	AAGCGGG	GGAGCATGTGGT	TTAATTCGA	AGCAACGC	GAA-AAC	CTTACCA-GTC	897
Sbjct	885	CCGCAC	AAGCGGT	GGAGCATGTGGT	TTAATTCGA	AGCAACGC	GAAGAAC	CTTACCAGGTC	944
Query	898	TTGACI	TCCTCTG	AC-ACCCTAAAA	ATAGGGCTT	CCCCTTCG	GGG 94:	1	
Sbjct	945	TTGACA	TCCTCTG	ACAACCCTAGAG	ATAGGGCTI	CCCCTTCG	GGG 98	9	



0.001

Enterococcus mundtii strain IHBB 9250 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb[KR085796.1]</u> Length: 1518 Number of Matches: 1

Range 1:	81 to 103	3 GenBank Graphics		V N	ext Match 🔺 Previous M	latch	
Score 1674 bit	s(1856)	Expect 0.0	Identities 944/953(99%)	Gaps 1/953(0%)	Strand Plus/Plus		
Query	1	CCGGGAAAAGAG	GAGTGGCGAACGGG	TGAGTAACACGT	GGTAACCTGCCCA	TCAGAAGG	60
Sbjct	81	CCGGGAAAAGAG	GAGTGGCGAACGGG	TGAGTAACACGT	GGTAACCTGCCCA	 TCAGAAGG	140
Query	61	GGATAACACTTO	GGAAACAGGTGCTAA	TACCGTATAACA	ATCGAAACCGCATG	GTTTCGTT	120
Sbjct	141	GGATAACACTTO	GGAAACAGGTGCTAA	TACCGTATAACAA	ATCGAAACCGCATG	GTTTCGTT	200
Query	121	TTGAAAGGCGCT		ATGGATGGACCCO	GCGGTGCATTAGCT.	AGTTGGTG	180
Sbjct	201	TTGAAAGGCGC	TTACGGTGCCGCTG	ATGGATGGACCCO	GCGGTGCATTAGCT.	AGTTGGTG	260
Query	181	AGGTAACGGCT	CACCAAGGCCACGA1	GCATAGCCGACC	IGAGAGGGTGATCG	GCCACATT	240
Sbjct	261	AGGTAACGGCT	CACCAAGGCCACGAI	GCATAGCCGACC	IGAGAGGGGTGATCG	GCCACATT	320
Query	241	GGGACTGAGACA	ACGGCCCAAACTCCI	ACGGGAGGCAGC	AGTAGGGAATCTTC	GGCAATGG	300
Sbjct	321	GGGACTGAGAC	ACGGCCCAAACTCCI	ACGGGAGGCAGC	AGTAGGGAATCTTC	GGCAATGG	380
Query	301	ACGAAAGTCTG		GTGAGTGAAGAAG	GTTTTCGGATCGT	AAAACTCT	360
Sbjct	381	ACGAAAGTCTG	ACCGAGCAACGCCGC	GTGAGTGAAGAAG	GTTTTCGGATCGT.	AAAACTCT	440
Query	361	GTTGTTAGAGA	AGAACAAGGGTGAGA	GTAACTGTTCAC	CCTTGACGGTATC	TAACCAGA	420
Sbjct	441	GTTGTTAGAGA	AGAACAAGGGTGAGA	GTAACTGTTCACC	CCTTGACGGTATC	TAACCAGA	500
Query	421	AAGCCACGGCTA	AACTACGTGCCAGCA	GCCGCGGTAATA	CGTAGGTGGCAAGC	GTTGTCCG	480
Sbjct	501	AAGCCACGGCTZ	ACTACGTGCCAGCA	GCCGCGGTAATAC	CGTAGGTGGCAAGC	GTTGTCCG	560
Query	481	GATTTATTGGGG		GGCGGTTTCTTA	AGTCTGATGTGAAA	GCCCCCGG	540
Sbjct	561	GATTTATTGGGG	CGTAAAGCGAGCGCA	GGCGGTTTCTTA	AGTCTGATGTGAAA	GCCCCCGG	620
Query	541		AGGGTCATTGGAAAC	TGGGAGACTTGA	GTGCAGAAGAGGAG	AGTGGAAT	600
Sbjct	621	CTCAACCGGGGG	AGGGTCATTGGAAAC	TGGGAGACTTGA	STGCAGAAGAGGAG	AGTGGAAT	680
Query	601	TCCATGTGTAG	CGGTGAAATGCGTAG	ATATATGGAGGA	ACACCAGTGGCGAA	GGCGGCTC	660
Sbjct	681	TCCATGTGTAG	CGGTGAAATGCGTAG	ATATATGGAGGA	ACACCAGTGGCGAA	GGCGGCTC	740
Query	661	TCTGGTCTGTA	ACTGACGCTGAGGCI	CGAAAGCGTGGGG	GAGCAAACAGGATT	AGATACCC	720
Sbjct	741	TCTGGTCTGTA	ACTGACGCTGAGGCI	CGAAAGCGTGGGG	GAGCAAACAGGATT	AGATACCC	800
Query	721	TGGTAGTCCAC	GCCGTAAACGATGAG	TGCTAAGTGTTG	GAGGGTTTCCGCCC	TTCAGTGC	780
Sbjct	801	TGGTAGTCCAC	SCCGTAAACGATGAG	TGCTAAGTGTTG	GAGGGTTTCCGCCC	TTCAGTGC	860
Query	781	TGCAGCTAACGO			ACCGCAAGGTTGAA	ACTCAAAG	840
Sbjct	861	TGCAGCTAACGO	CATTAAGCACTCCGC	CTGGGGAGTACG	ACCGCAAGGTTGAA	ACTCAAAG	920
Query	841	GAATTGACGGGG	GCCCGCACAAGCGG	GGGAGCATGGGG	TTAATTCGAAGCA	ACGCGAAA	900
Sbjct	921	GAATTGACGGGG	GCCCGCACAAGCGG	TGGAGCATGTGG	TTAATTCGAAGCA	ACGCGAAG	980
Query	901	AACCTTACCA-0	GTCTTGACATCCTTI	GACCCCTCTaaaa	aaaaaGCTTCCCC	т 952 I	
Sbjct	981	AACCTTACCAG	GTCTTGACATCCTTI	GACCACTCTAGA	GATAGAGCTTCCCC	т 1033	



Enterococcus hirae KT261200.1

Enterococcus hirae strain RCB988 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KT261200.1</u>| Length: 1437 Number of Matches: 1

Range 1:	59 to 96	5 GenBank Graphics		•	Next Match 🔺 Previous Match	
Score		Expect	Identities	Gaps	Strand	
1546 bit	s(1714)	0.0	892/910(98%)	4/910(0%)	Plus/Plus	
Query	1	AGTAACACGTGGG	TAACCTGCCCATC	AGAAGGGGATAAC	CACTTGGAAACAGGTGCTAATA	60
Sbjct	59	AGTAACACGTGGG	TAACCTGCCCATCA	AGAAGGGGATAAC	CACTTGGAAACAGGTGCTAATA	118
Query	61	CCGTATAACAATC	GAAACCGCATGGT		GCGCTTTCGGGTGTCGCTGAT	120
Sbjct	119	CCGTATAACAATC	GAAACCGCATGGT	TTTGATTTGAAAO	GCGCTTTCGGGTGTCGCTGAT	178
Query	121	GGATGGACCCGCG	GTGCATTAGCTAG	TTGGTGAGGTAAC	CGGCTCACCAAGGCCACGATGC	180
Sbjct	179	GGATGGACCCGCG	GTGCATTAGCTAG	TTGGTGAGGTAAC	CGGCTCACCAAGGCGACGATGC	238
Query	181	ATAGCCGACCTGA	GAGGGTGATCGGC	CACATTGGGACTG	GAGACACGGCCCAAACTCCTAC	240
Sbjct	239	ATAGCCGACCTGA	GAGGGTGATCGGCC	CACATTGGGACTO	GAGACACGGCCCAAACTCCTAC	298
Query	241	GGGAGGCAGCAGT		CAATGGACGAAAG	TCTGACCGAGCAACGCCGCGT	300
Sbjct	299	GGGAGGCAGCAGT	AGGGAATCTTCGGC	CAATGGACGAAAG	STCTGACCGAGCAACGCCGCGT	358
Query	301	GAGTGAAGAAGGT	TTTCGGATCGTAA		AGAGAAGAACAAGGATGAGAGT	360
Sbjct	359	GAGTGAAGAAGGT	TTTCGGATCGTAAA	ACTCTGTTGTT	AGAGAAGAACAAGGATGAGAGT	418
Query	361	AACTGTTCATCCC	TTGACGGTATCTA		CGGCTAACTACGTGCCAGCAGC	420
Sbjct	419	AACTGTTCATCCC	TTGACGGTATCTA	ACCAGAAAGCCAC	CGGCTAACTACGTGCCAGCAGC	478
Query	421	CGCGGTAATACGT	AGGTGGCAAGCGT	IGTCCGGATTTAT	TTGGGCGTAAAGCGAGCGCAGG	480
Sbjct	479	CGCGGTAATACGT	AGGTGGCAAGCGT	IGTCCGGATTTAI	TTGGGCGTAAAGCGAGCGCAGG	538
Query	481	CGGTTTCTTAAGT	CTGATGTGAAAGCO	CCCCGGCTCAACO	CGGGGAGGGTCATTGGAAACTG	540
Sbjct	539	CGGTTTCTTAAGT	CTGATGTGAAAGCO	CCCCGGCTCAACO	CGGGGAGGGTCATTGGAAACTG	598
Query	541	GGAGACTTGAGTG	CAGAAGAGGAGAGAG	IGGAATTCCATG	IGTAGCGGTGAAATGCGTAGAT	600
Sbjct	599	GGAGACTTGAGTG	CAGAAGAGGAGAG	IGGAATTCCATG:	IGTAGCGGTGAAATGCGTAGAT	658
Query	601	ATATGGAGGAACA	CCAGTGGCGAAGG(CGGCTCTCTGGT(CTGTAACTGACGCTGAGGCTCG	660
Sbjct	659	ATATGGAGGAACA	CCAGTGGCGAAGG	CGGCTCTCTGGT	CTGTAACTGACGCTGAGGCTCG	718
Query	661	AAAGCGTGGGGGA		GATACCCTGGTA(JTCCACGCCGTAAACGATGAGT	720
Sbjct	719	AAAGCGTGGGGGA	GCAAACAGGATTA	GATACCCTGGTA	GTCCACGCCGTAAACGATGAGT	778
Query	721	GCTAAGTGTTGGA	GGGTTTCCGCCCT	CAGTGCTGCAG	CTAACGCATTAAACACTCCGCC	780
Sbjct	779	GCTAAGTGTTGGA	GGGTTTCCGCCCT	TCAGTGCTGCAG		838
Query	781	TGGGGAGTACAAC				840
JUCT	039	CCCA A CUMCHCCCC				000
Query	041 041	-GGAGCATCTGTGGT		AACCCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA		025
Oueru	000	TTGACCCCTC	11AA-TICGAAGCA	AUGU-GAAGAA(JULIAUUAGGIUIIGAUATUUT	900
Sbict	956	TTGACCACTC 9	65			

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	firmicutes 2 leaves
	Enterococcus durans strain SJU18 16S ribosomal RNA gene, partial sequence Enterococcus durans strain SJU18 16S ribosomal RNA gene, partial sequence Enterococcus durans strain SJU18 16S ribosomal RNA gene, partial sequence
	Enterococcus lactis sarial COLDU-10/105 notional rAVA gene, partial sequence Enterococcus lactis narial 16S rRNA eene. strain az-548
	Enterococcus faecalis strain TS4E2 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain KS3S1 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium gene for 16S ribosomal RNA, partial sequence, isolate: VMCU54N Enterococcus faecium dmin D 16S ribosomal DNA gene, partial sequence.
	Enterococcus faecium strain D fos hostonal refor gene, partial sequence Enterococcus faecium strain AT15 16S ribosomal RNA eene, nartial sequence
	Enterococcus sp. 55B 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain S. 156 16S ribosomal RNA gene, partial sequence
	Generococcus faecium strain V9-200 IoS ribosomal RNA gene, partial sequence Generococcus faecium strain V9.158 IoS ribosomal RNA gene, partial sequence
	Enterococcus faecium strain V9-156 16S ribosonal RNA gene, partial sequence
	Enterococcus faecium strain V9-152 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain V9-150 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain V9-14/ 105 mosomal RNA gene, partial sequence Enterococcus faecium strain V6-112 16S ribosomal RNA gene, partial sequence
	 Enterococcus faecium strain V6-112 fos nosonial RNA gene, partial sequence Enterococcus faecium strain V6-110 I6S ribosonial RNA gene, partial sequence
	Enterococcus faecium strain E0-190 16S ribosomal RNA gene, partial sequence
	 Enterococcus faecium strain V3-101 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain EO-192 16S ribosomal RNA gene, partial sequence Enterococcus faecium strain V3 82 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain V3-82 fos hoosonal RNA gene, partial sequence
	 Enterococcus faecium strain E3-67 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain E3-66 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain V12-65 16S ribosomal RNA gene, partial sequence
	 Enterococcus faecium strain V12-25 ToS ribosomal RNA gene, partial sequence Enterococcus faecium strain V12-20 16S ribosomal RNA gene, partial sequence
	Lactobacillus helveticus strain GM2 16S ribosonal RNA gene, partial sequence
	Enterococcus faecium strain gp39 16S ribosomal RNA gene, partial sequence
	Enterococcus durans strain gp85 16S ribosomal RNA gene, partial sequence
	Enterococcus durans strain gp58 165 ribosomal KNA gene, partial sequence O Enterococcus lactis strain SCEN1403 165 ribosomal RNA gene, partial sequence
	Entrococcus lactis strain SCEN1402 16S ribosomal RNA gene, partial sequence
	Enterococcus lactis strain SCEN1401 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain Zil 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain 1PO IOS fibosomal KNA gene, partial sequence Enterococcus faecium strain VPT IOS ribosomal RNA gene, partial sequence
	Enterococcus faecium strain YP 16S ribosomal RNA gene, partial sequence
1 0000	Enterococcus faecium strain WVS333 16S ribosomal RNA gene, partial sequence
1.0008	Enterococcus faecium strain BIBT_VC_I 16S ribosomal RNA gene, partial sequence
	Enterococcus hirae strain CSCW12-12 16S ribosomal RNA gene, partial sequence Enterococcus facium strain PSB 5 16S ribosomal RNA gene, partial sequence
	Bacterium QHB36 16S ribosomal RNA gene, partial sequence
	Bacterium QHB33 16S ribosomal RNA gene, partial sequence Enteronogene facility entering SM67 16S ribosomal RNA gene, partial sequence
	 Uncultured Enterococcus sp. clone 58 16S ribosomal RNA gene, partial sequence
	 Uncultured Enterococcus sp. clone 13 16S ribosomal RNA gene, partial sequence Enterococcus facium cturia 74 21 16S ribosomal RNA gene, partial sequence
	 Enterococcus faecium strain Z4-21 fos fitosomal RNA gene, partial sequence Enterococcus faecium strain CA13 16S ribosomal RNA gene, complete sequence
	Enterococcus faecium strain CA03-1 16S ribosomal RNA gene, complete sequence DA46 16S ribosomal RNA gene, complete sequence
	Bacterium P2A5 165 ribosomal RNA gene, partial sequence Bacterium P2E9 16S ribosomal RNA gene, partial sequence
	Enterococcus durans strain KLDS 6.0933 16S ribosomal RNA gene, partial sequence
	Enterococcus durans strain KDLLJ2-1 16S ribosomal RNA gene, partial sequence O Enterococcus hirae strain RCB988 16S ribosomal RNA gene partia
	Bacterium MRG-FC-3 16S ribosomal RNA gene, partial sequence
	firmicutes 9 leaves Bacterium SID 16S ribocomal RNA cene nartial sequence
	Enterococcus faecium strain TPBLGL 30 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain SAMBN-4 16S ribosomal RNA gene, partial sequence Unaultured hostadium clane 2, 57, 1471 16S ribosomal RNA gene, partial sequence
	Enterococcus durans strain SMN1-2 16S ribosonal RNA gene, partial sequence
	Enterococcus hirae strain C07_1387_03 16S ribosomal RNA gene, partial sequence
	 Enterococcus sp. QAUEP07 IbS ribosomal RNA gene, partial sequence Enterococcus sp. OAUEF01 I6S ribosomal RNA gene, partial sequence
	Enterococcus sp. QAUEF03 16S ribosomal RNA gene, partial sequence
	Enterococcus sp. QAUEF04 16S ribosomal RNA gene, partial sequence
	Entrococcus sp. QAUEF05 16S ribosonal RNA gene, partial sequence
	Enterococcus sp. MRSA78B1_13_1A 16S ribosomal RNA gene, partial sequence Enterococcus on MBSA87B1_13_1A 16S ribosomal RNA gene, partial sequence
	Finetococcus sp. WRSA93B1_13_2E 16S ribosomal RVA gene, partial sequence
	Enterococcus sp. MRSA145B15_13_2E 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain IMAULI105/16S/nbosoma1/KNA gene, partial sequence Enterococcus faecium strain IMAULI167/16S/ribosoma1/RNA gene, nartial sequence
	Enterococcus durans strain IMAU11650 16S ribosomal RNA gene, partial sequence
	Enterococcus durans strain IMAU11652 16S ribosomal RNA gene, partial sequence Enterococcus facium strain IMAU11658 16S ribosomal RNA gene, partial assumeses
	Enterococcus facculus strain NK1 16S ribosomal RNA gene, partial sequence Enterococcus faccalis strain NK1 16S ribosomal RNA gene, partial sequence
	Enterococcus faecium strain NK8 16S ribosomal RNA gene, partial sequence Enterococcus faecium strain NK8 10S ribosomal RNA gene, partial sequence
	g Enterococcus durans strain IMAU 11458 (DM17-3) 105 ribosomal RNA gene, partial sequence g Enterococcus durans strain IMAU11321 (YM36-10) 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 4L1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 3L6 16S ribosomal RNA gene, partial sequence
8,6883	s uncumired nacierium come 21.5 ros nossoniai KNA gene, partiai sequénce firmicutes 2 leaves
	Enterococcus faecium strain 64/3, complete genome

Bacillus licheniformis DQ071560.1

Bacillus licheniformis strain MKU 1 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb[DQ071560.1]</u> Length: 1430 Number of Matches: 1

Range 1:	431 to 13	93 GenBank Graphics		V 1	Next Match 🔺 Previous Match	
Score		Expect Io	lentities	Gaps	Strand	
1703 bit	s(1888)	0.0 9	58/963(99%)	3/963(0%)	Plus/Minus	
Query	1	TTACAAACTCTCG	TGGTGTGACGGGC	GGTGTGTACAAG	GCCCGGGAACGTATTCACCGCG	60
Sbjct	1393	TTACAAACTCTCG	TGGTGTGACGGGC	GGTGTGTACAAG	GCCCGGGAACGTATTCACCGCG	1334
Query	61	GCATGCTGATCCG	CGATTACTAGCGA:	TTCCAGCTTCAC	GCAGTCGAGTTGCAGACTGCGA	120
Sbjct	1333	GCATGCTGATCCG	CGATTACTAGCGA	ITCCAGCTTCAC	GCAGTCGAGTTGCAGACTGCGA	1274
Query	121	TCCGAACTGAGAA	CAGATTTGTGGGA	FTGGCTTAGCCT 	CGCGGCTTCGCTGCCCTTTGTT	180
Sbjct	1273	TCCGAACTGAGAA	CAGATTTGTGGGA	ITGGCTTAGCCT	CGCGGCTTCGCTGCCCTTTGTT	1214
Query	181	CTGCCCATTGTAG	CACGTGTGTAGCC	CAGGTCATAAGG	GGCATGATGATTTGACGTCATC	240
Sbjct	1213	CTGCCCATTGTAG	CACGTGTGTAGCC	CAGGTCATAAGG	GGCATGATGATTTGACGTCATC	1154
Query	241	CCCACCTTCCTCC	GGTTTGTCACCGG	CAGTCACCTTAG	AGTGCCCAACTGAATGCTGGCA	300
Sbjct	1153	CCCACCTTCCTCC	GGTTTGTCACCGG	CAGTCACCTTAG	AGTGCCCAACTGAATGCTGGCA	1094
Query	301	ACTAAGATCAAGG	GTTGCGCTCGTTG	CGGGACTTAACC	CAACATCTCACGACACGAGCTG	360
Sbjct	1093	ACTAAGATCAAGG	GTTGCGCTCGTTG	CGGGACTTAACC	CAACATCTCACGACACGAGCTG	1034
Query	361	ACGACAACCATGC	ACCACCTGTCACT	CTGCCCCCGAAG	GGGAAGCCCTATCTCTAGGGTT	420
Sbjct	1033	ACGACAACCATGC	ACCACCTGTCACT	CTGCCCCCGAAG	GGGAAGCCCTATCTCTAGGGTT	974
Query	421	GTCAGAGGATGTC	AAGACCTGGTAAG(GTTCTTCGCGTT	GCTTCGAATTAAACCACATGCT	480
Sbjct	973	GTCAGAGGATGTC.	AAGACCTGGTAAG	GTTCTTCGCGTT	GCTTCGAATTAAACCACATGCT	914
Query	481	CCACCGCTTGTGC	GGGCCCCCGTCAA:	TTCCTTTGAGTT	TCAGTCTTGCGACCGTACTCCC	540
Sbjct	913	CCACCGCTTGTGC	GGGCCCCCGTCAA	TTCCTTTGAGTT	TCAGTCTTGCGACCGTACTCCC	854
Query	541 853		TAATGCGTTTGCTC TAATGCGTTTGCTC	GCAGCACTAAAG	GGCGGAAACCCTCTAACACTTA	600 794
Query	601	GCACTCATCGTTT	ACGGCGTGGACTA	CCAGGGTATCTA	ATCCTGTTCGCTCCCCACGCTT	660
~ - Sbjct	793	 GCACTCATCGTTT	 ACGGCGTGGACTAG	 CCAGGGTATCTA	ATCCTGTTCGCTCCCCACGCTT	734
Query	661	TCGCGCCTCAGCG	TCAGTTACAGACCA	AGAGAGTCGCCT	TCGCCACTGGTGTTCCTCCACA	720
Sbjct	733	TCGCGCCTCAGCG	TCAGTTACAGACC	AGAGAGTCGCCT	TCGCCACTGGTGTTCCTCCACA	674
Query	721	TCTCTACGCATTT	CACCGCTACACGT	GGAATTCCACTC	TCCTCTTCTGCACTCAAGTTCC	780
Sbjct	673	TCTCTACGCATTT	CACCGCTACACGT	GAATTCCACTC	TCCTCTTCTGCACTCAAGTTCC	614
Query	781	CCAGTTTCCAATG	ACCCTCCCCGGTT	GAGCCGGGGGGCT	TTCACATCAAACTTAAAAAACC	840
Sbjet	013	CCAGTTTCCAATG	ACCCTCCCCGGTTC	JAGCCGGGGGGC'I	TTCACATCAAACTTAAAAAAACC	000
Sbict	553	GCCTGCGCGCGCGCT	TTACGCCCAATAA	IIIIIIII IIII IIIIIIII IIII TTCCGGACAACG	CTTGCCACCTACGTATTACCGC	494
Query	900	GGCTGCTGGCACG	TAGTTAGCCGGGGG	CTTTCTGGTTAG	GTACCGTC-AGGTACC-CCCTA	957
Sbjct	493	GGCTGCTGGCACG	IIIIIIIIIIIIIII TAGTTAGCCGTGG0	 CTTTCTGGTTAG		434
Query	958	TTC 960				
Sbjct	433	TTC 431				

	Bacillus licheniformis strain MKU I 16S ribosomal RNA gene, partial sequence	■IciQuery_396
	 Bacillus icheniformis strain XIV 2011 ISS ribosomal RNA gene, partial sequence Bacillus icheniformis WX-02 genome Bacillus icheniformis WX-02 genome Bacillus icheniformis RNA gene, partial sequence Bacillus icheniformis RNA gene, partial sequence Bacillus icheniformis train TS (08 165 nbosomal RNA gene, partial sequence Bacillus icheniformis train TS (08 165 nbosomal RNA gene, partial sequence Bacillus icheniformis train TS (08 165 nbosomal RNA gene, partial sequence Bacillus icheniformis train ILTW 8 165 nbosomal RNA gene, partial sequence Bacillus icheniformis train ILTW 8 165 nbosomal RNA gene, partial sequence Bacillus icheniformis train ILTW 8 156 nbosomal RNA gene, partial sequence Bacillus icheniformis train ILTW 78 5247(8) fost nbosomal RNA gene, partial sequence Bacillus icheniformis train CITT MFB 5247(8) fost nbosomal RNA gene, partial sequence Bacillus icheniformis train ZIY-557 165 ribosomal RNA gene, partial sequence Bacillus icheniformis train XV12 165 ribosomal RNA gene, partial sequence Bacillus icheniformis strain RNA yerl, partial RNA gene, partial sequence Bacillus icheniformis strain RNA yerl, partial sequence Bacillus icheniformis strain RNA gene, partial sequence Bacillus icheniformis strain RNA gene, partial sequence Bacillus icheniformis strain RNA gene, partial sequence Bacillus icheniformis strain CL-58 165 ribosomal RNA gene, partial sequence Bacillus icheniformis strain CL-58 165 ribosomal RNA gene, partial sequence Bacillus icheniformis strain KP120 165 ribosomal RNA gene, partial sequence Bacillus icheniformis strain KP120 165 ribosomal RNA gene, partial sequence Bacillus icheniformis strain XP120 165 ribosomal RNA gene, partial sequence Bacillus ic	

Lysinibacillus fusiformis KP872952.1

Lysinibacillus fusiformis strain XKS50.1 16S ribosomal RNA gene, partial sequence Sequence ID: <u>KP872952.1</u> Length: 1450 Number of Matches: 1

Range 1: 1 to 1450 GenBank Graphics Vext Match A Previous Match						
Score 2678 bit	s(1450)	Expect 0.0	Identities 1450/1450(100%)	Gaps 0/1450(0%)	Strand Plus/Plus	
Ouerv	1	CCCGGGGGGGT	CCTATACATGCAAGTC	GAGCGAACAGAAAAG	GAGCTTGCTCCTTTGACGT	60
Sbjct	1	CCCGGGGGGGT	CCTATACATGCAAGTC	GAGCGAACAGAAAAG	GAGCTTGCTCCTTTGACGT	60
Query	61	TAGCGGCGGA	CGGGTGAGTAACACGT	GGGCAACCTACCCTA	TAGTTTGGGATAACTCCGG	120
Sbjct	61	TAGCGGCGGA		 GGGCAACCTACCCTA	 TAGTTTGGGATAACTCCGG	120
Query	121	GAAACCGGGGG	CTAATACCGAATAATC	TCTTTTGCTTCATGG	TGAAAGACTGAAAGACGGT	180
Sbjct	121	GAAACCGGGGG	CTAATACCGAATAATC	TCTTTTGCTTCATGG	TGAAAGACTGAAAGACGGT	180
Query	181	TTCGGCTGTC	CTATAGGATGGGCCC	GCGGCGCATTAGCTA	GTTGGTGAGGTAACGGCTC	240
Sbjct	181	TTCGGCTGTC	SCTATAGGATGGGCCC	GCGGCGCATTAGCTA	GTTGGTGAGGTAACGGCTC	240
Query	241	ACCAAGGCGA	CGATGCGTAGCCGACC	TGAGAGGGTGATCGG	CCACACTGGGACTGAGACA	300
Sbjct	241	ACCAAGGCGA	CGATGCGTAGCCGACC	TGAGAGGGTGATCGG	CCACACTGGGACTGAGACA	300
Query	301	CGGCCCAGAC	fcctacgggaggcagc:	AGTAGGGAATCTTCC:	ACAATGGGCGAAAGCCTGA	. 360
Sbjct	301	CGGCCCAGAC	TCCTACGGGAGGCAGC	AGTAGGGAATCTTCC	ACAATGGGCGAAAGCCTGA	360
Query	361	TGGAGCAACG	CCGCGTGAGTGAAGAA	GGTTTTCGGATCGTA	AAACTCTGTTGTAAGGGAA	420
Sbjct	361	TGGAGCAACG	CGCGTGAGTGAAGAA	GGTTTTCGGATCGTA	AAACTCTGTTGTAAGGGAA	420
Query	421	GAACAAGTAC	AGTAGTAACTGGCTGT.	ACCTTGACGGTACCT	TATTAGAAAGCCACGGCTA	480
Sbjct	421	GAACAAGTAC	AGTAGTAACTGGCTGT	ACCTTGACGGTACCT	TATTAGAAAGCCACGGCTA	480
Query	481	ACTACGTGCC	AGCAGCCGCGGTAATA	CGTAGGTGGCAAGCG	TTGTCCGGAATTATTGGGC	540
Sbjct	481	ACTACGTGCC	AGCAGCCGCGGTAATA	 CGTAGGTGGCAAGCG		540
Query	541	GTAAAGCGCGG	GCAGGCGGTCCTTTA	AGTCTGATGTGAAAG	CCCACGGCTCAACCGTGGA	600
Sbjct	541	GTAAAGCGCGG	GCAGGCGGTCCTTA	AGTCTGATGTGAAAG	CCCACGGCTCAACCGTGGA	600
Query	601	GGGTCATTGG	AACTGGGGGACTTGA	GTGCAGAAGAGGAAA	GTGGAATTCCAAGTGTAGC	660
Sbjct	601	GGGTCATTGG	AACTGGGGGACTTGA	GTGCAGAAGAGGAAA	GTGGAATTCCAAGTGTAGC	660
Query	661	GGTGAAATGC	TAGAGATTTGGAGGA	ACACCAGTGGCGAAG	GCGACTTTCTGGTCTGTAA	720
Sbjct	661	GGTGAAATGC		 ACACCAGTGGCGAAG	 GCGACTTTCTGGTCTGTAA	720
Query	721	CTGACGCTGA	GCGCGAAAGCGTGGG	GAGCAAACAGGATTA	GATACCCTGGTAGTCCACG	780
Sbjct	721	CTGACGCTGA	GCGCGAAAGCGTGGG	GAGCAAACAGGATTA	GATACCCTGGTAGTCCACG	780
Query	781	CCGTAAACGAT	GAGTGCTAAGTGTTA	GGGGGTTTCCGCCCC	TTAGTGCTGCAGCTAACGC	840
Sbjct	781	CCGTAAACGAT	GAGTGCTAAGTGTTA	GGGGGTTTCCGCCCC	TTAGTGCTGCAGCTAACGC	840
Query	841	ATTAAGCACTC	CGCCTGGGGAGTACG	GTCGCAAGACTGAAA	CTCAAAGGAATTGACGGGG	; 900
Overv	901	GCCCGCACAC	CGCCTGGGGGGGGAGTACG	TTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT	CTCAAAGGAATTGACGGGG	, 900 : 960
Sbict	901	GCCCGCACAA	CGGTGGAGCATGTGG	TTTAATTCGAAGCAA	CGCGAAGAACCTTACCAG	; 960
Query	961	TCTTGACATCO	CGTTGACCACTGTAG	AGATATAGTTTCCCC	TTCGGGGGCAACGGTGACA	1020
Sbjct	961	TCTTGACATCO	CGTTGACCACTGTAG	 AGATATAGTTTCCCC'	 TTCGGGGGGCAACGGTGACA	1020
Query	1021	GGTGGTGCATC	GTTGTCGTCAGCTCG	TGTCGTGAGATGTTG	GGTTAAGTCCCGCAACGAG	; 1080
Sbjct	1021	GGTGGTGCATO	GTTGTCGTCAGCTCG	TGTCGTGAGATGTTG	GGTTAAGTCCCGCAACGAG	; 1080
Query Sbjct	1081 1081	CGCAACCCTTC	SATCTTAGTTGCCATC	ATTTAGTTGGGCACT ATTTAGTTGGGCACT	CTAAGGTGACTGCCGGTGA	1140 1140
Query	1141	CAAACCGGAGG	AAGGTGGGGATGACG	TCAAATCATCATGCC	CCTTATGACCTGGGCTACA	1200
Sbjct	1141	CAAACCGGAGG	AAGGTGGGGATGACG	IIIIIIIIIIIIII TCAAATCATCATGCC	CCTTATGACCTGGGCTACA	1200
Query	1201	CACGTGCTAC	ATGGACGATACAAAC	GGTTGCCAACTCGCG.	AGAGGGAGCTAATCCGATA	1260
Sbjct	1201	CACGTGCTAC	ATGGACGATACAAAC	GGTTGCCAACTCGCG	AGAGGGAGCTAATCCGATA	1260
Query	1261	AAGTCGTTCTC	AGTTCGGATTGTAGG	CTGCAACTCGCCTAC	ATGAAGCCGGAATCGCTAG	; 1320
Sbjct	1261	AAGTCGTTCTC	AGTTCGGATTGTAGG	CTGCAACTCGCCTAC	ATGAAGCCGGAATCGCTAG	; 1320
Query	1321	TAATCGCGGAT	CAGCATGCCGCGGTG	AATACGTTCCCGGGC	CTTGTACACACCGCCCGTC	: 1380
Sbjct	1321	TAATCGCGGAT	CAGCATGCCGCGGTG	AATACGTTCCCGGGC	CTTGTACACACCGCCCGTC	1380
Sbjct	1381	ACACCACGAGA	GTTTGTAACACCCGA 	MGTCGGTGAGGTAAC AGTCGGTGAGGTAAC	CTITGEAGCCAGCCGCCGA 	1440 L440
Query	1441	AGGTGATGAT	1450			
Sbjct	1441	AGGTGATGAT	1450			



0.0001

Bacillaceae bacterium SE17 16S ribosomal RNA gene, partial sequence Bacillaceae bacterium SE41 16S ribosomal RNA gene, partial sequence Lysimbacillus xylanil yticus strain GT18 165 robosomal RNA gene, partial sequence Lysimbacillus sphaericus strain BL1 165 ribosomal RNA gene, partial sequence Lysimbacillus fusiformis strain BL2 165 ribosomal RNA gene, partial sequence Lysinibacillus sphaericus strain BL7 16S ribosomal RNA gene, partial sequence Bacillus sp. PZ_7 16S ribosomal RNA gene, partial sequence Lysinibacillus sp. enrichment culture clone MJJ-11 16S ribosomal RNA gene, partial sequence Usinibacillus fusiformis strain R3 165 ribosonal RNA gene, partial sequence Qacillas p. 81 165 ribosonal RNA gene, partial sequence Usinibacillus fusiformis strain TL 165 ribosonal RNA gene, partial sequence Uysinhacillus rusiformis strai ILI ios nossomal KNA gene, partial sequence Uysinhacillus fusiformis strai ILI oli 66 ifoshosmal RNA gene, partial sequence Uysinhacillus fusiformis strain N169 16S ribosomal RNA gene, partial sequence Uysinhacillus fusiformis strain SI oli 65 ribosomal RNA gene, partial sequence Uysinhacillus fusiformis strain SI oli 66 ribosomal RNA gene, partial sequence Bacillales bacterium Cul 0304 16S ribosomal RNA gene, partial sequence Bachtliels hadernum Cull (1994) IOS Infosomal RNA gene, partal sequence Uysinhaedluss p. 1909 IOS ribosomal RNA gene, partal sequence Uysinhaedluss p. SSI.22 IOS ribosomal RNA gene, partal sequence Uysinhaedlus fusiformis strain 44A IOS ribosomal RNA gene, partal sequence Uysinhaedlus fusiformis strain L106 IOS ribosomal RNA gene, partal sequence Uysinhaedlus fusiformis strain L106 IOS ribosomal RNA gene, partal sequence Usyninacius instorms si ran L106 105 ritosoma KvA gene, partal sequence @Cobacillus settorhempolitius strati KS141 165 ritosomal RVA gene, partal sequence % Firmientes bacterium K171 165 ritosomal RVA gene, partal sequence % jusimbacillus bacterium K171 165 ritosomal RVA gene, partal sequence % jusimbacillus py 2016 firitosomal RNA gene, contral sequence % jusimbacillus macroides strain fwzy1 165 ritosomal RNA gene, partal sequence Lysinibacillus macroides strain fwzy182 16S ribosomal RNA gene, partial sequence Vysinhacillus fusiformis partial 165 rRNA gene, isota PA feele, partial se Lysinhacillus fusiformis partial 165 rRNA gene, isotae AVSI Oysinhacillus macroides partial 165 rRNA gene, isotae AVSI Proteus mirabilis strain BCr 165 ribosomal RNA gene, partial sequence Lysinhacillus p. WI, 10-223 165 ribosomal RNA gene, complete sequence Uysimbacilus sp. 52-165 ribosomal RNA gene, partial sequence Uncultured Bacillus sp. clone XT58 165 ribosomal RNA gene, partial sequence Usimbacillus fusiformis strain 164B (BP1) 165 ribosomal RNA gene, partial sequence Usimilacius usuotinis Vata (UP) (UF) (US) 100 noteenaa eve gava, paraa page Unculured bacterium gene for 165 rohosomal RNA, paraial sequence, clone: 20/B1 Usimihacillus fusiformis strain S-1 16S rohosomal RNA gene, paraial sequence Usimihacillus sp. XJIC-134-1RF1 16S ribosomal RNA gene, paraial sequence Usinibacillus sp. BAB-292 165 ribosomal RNA gene, partial sequence Usinibacillus sp. BAB-692 165 ribosomal RNA gene, partial sequence Usinibacillus sp. BAB-637 165 ribosomal RNA gene, partial sequence Lysinibacillus sphaericus strain B1-CDA 16S ribosomal RNA gene, partial sequence Lysinibacillus sylanilyticus strain BD3 16S ribosomal RNA gene, partial sequence

Aneurinibacillus migulanus (Bacillus brevis) NR_113764.1

Aneurinibacillus migulanus strain NBRC 15520 16S ribosomal RNA gene, partial sequence Sequence ID: <u>ref[NR_113764.1]</u> Length: 1464 Number of Matches: 1 > <u>See 1 more title(s)</u>

Range 1:	28 to 144	11 <u>GenBank</u> <u>Graph</u>	ics		🔻 Next Match	n 🔺 Previous Mato	:h
Score 2531 bit	ts <mark>(2806)</mark>	Expect 0.0	Identities 1411/1414(99%)	Gaps 2/1414(0%	5t b) Pl	rand us/Plus	
Query	18	ATGCA-GTCG	AGCGGACCAATGAA	GAGCTTGCTCTT	CGGCGGTI	AGCGGCGGAC	GGGTGA
Sbjct	28	ATGCAAGTCG	AGCGGACCAATGAA	GAGCTTGCTCTT	CGGCGGT	AGCGGCGGAC	GGGTGA
Query	77	GTAACACGTA	GGCAACCTGCCTGT	ACGACTGGGATA	ACTCCGGG	AAACCGGAGC	TAATAC
Sbjct	88	GTAACACGTA	GGCAACCTGCCTGT	ACGACTGGGATA	ACTCCGGG	GAAACCGGAGC	TAATAC
Query	137	CGGATACTTC	TTTCAGACCGCATG	GTCTGAAAGGGA	AAGACCTI	TGGTCACGTA	CAGATG
Sbjct	148	CGGATACTTC	TTTCAGACCGCATG	GTCTGAAAGGGA	AAGACNTT	TGGTCACGTA	CAGATG
Query	197	GGCCTGCGGC	GCATTAGCTAGTTG	GTGGGGTAACGG	CCTACCAZ	GGCGACGATG	CGTAGC
Sbjct	208	GGCCTGCGGC	GCATTAGCTAGTTG	GTGGGGGTAACGG	CCTACCAR	AGGCGACGATG	CGTAGC
Query	257	CGACCTGAGA	GGGTGATCGGCCAC	ACTGGGACTGAG	ACACGGCC	CAGACTCCTA	CGGGAG
Sbjct	268	CGACCTGAGA	GGGTGATCGGCCAC	ACTGGGACTGAG	ACACGGCC	CAGACTCCTA	.CGGGAG
Query	317	GCAGCAGTAG	GGAATCTTCCGCAA	TGGACGAAAGTC	rgacggag	CAACGCCGCG	TGAACG
Sbjct	328	GCAGCAGTAG	GGAATCTTCCGCAA	TGGACGAAAGTC	rgacggad	CAACGCCGCG	TGAACG
Query	377	ATGAAGGTTT	TCGGATCGTAAAGT	TCTGTTGTTAGG	GAAGAACO	GCCGGGATGA	CCTCCC
Sbjct	388	ATGAAGGTTT	TCGGATCGTAAAGT	TCTGTTGTTAGG	GAAGAACO	GCCGGGATGA	CCTCCC
Query	437	GGTCTGACGG		GCCCCGGCTAAC			GTAATA
Sbjct	448	GGTCTGACGG	TACCTAACGAGAAA	GCCCCGGCTAAC	FACGTGCC	AGCAGCCGCG	GTAATA
Query	497						TTCTTA
Sbjct	508	CGTAGGGGGC	AAGCGTTGTCCGGA	ATTATTGGGCGT	AAAGCGCC	CGCAGGCGGC	TTCTTA
Query	557	AGTCAGGTGTC	FAAAGCCCACGGCT		CCACTTO	AAACTGGGAA	GCTTGA
Sbjct	568	AGTCAGGTGTC	GAAAGCCCACGGCT	CAACCGTGGAGGG	GCCACTTO	AAACTGGGAA	GCTTGA
Query	617	GTGCAGGAGAG	GAGAGCGGAATTC	CACGTGTAGCGG	rgaaatgo	GTAGAGATGT	GGAGGA
Sbjct	628	GTGCAGGAGAG	GAGAGCGGAATTC	CACGTGTAGCGG	rgaaatgo	GTAGAGATGT	GGAGGA
Query	677	ACACCCGTGGC	CGAAGGCGGCTCTC	TGGCCTGTAACTO	FACGCTG#	GGCGCGAAAG	CGTGGG
Sbjct	688	ACACCCGTGGG	CGAAGGCGGCTCTC	TGGCCTGTAACTO	GACGCTGE	GGCGCGAAAG	CGTGGG
Query	737	GAGCGAACAG	GATTAGATACCCTG	GTAGTCCACGCCC	STAAACG1	TGAGTGCTAG	GTGTTG
Sbjct	748	GAGCGAACAG	JATTAGATACCCTG	GTAGTCCACGCCC	TAAACGI	TGAGTGCTAG	GTGTTG
Query	797	GGGACTCCAAT	fcctcagtgccgca	GCTAACGCAATA	AGCACTCC	GCCTGGGGAG	TACGGC
Sbjct	808	GGGACTCCAA	ICCTCAGTGCCGCA	GCTAACGCAATA	AGCACTCO	GCCTGGGGAG	TACGGC
Query	857	CGCAAGGCTGZ			GACAAGC	GGTGGAGCAT	GTGGTT
SDJCT	868	CGCAAGGCTGA	AACTCAAAGGAAT	TGACGGGGGACCCC	JCACAAGO	GGIGGAGCAI	GTGGTT
Query	917				JACATCCC		
SDJCT	928	TAATTCGAAGO	AACGCGAAGAACC	TTACCAGGGCTTC	JACATCCC	GCTGACCCTC	CTAGAG
Query	977					GTUGTCAGCT	
Jorg	1027	AFAGGAGCTC		GIGACAGGTGGT(CATGGT	GIUGTUAGCT	CGTGTC
Query	1049	GTGAGATGTTC				TAGTTGCCA	GCATTT
SUJCT	1007	A CERCAGATOTTO	GGTTAAGTUUUGU	AACGAGCGCAACC		T TAGTTGCCA	CCECTT
guery	1100	AGTTGGGCAC				GIGGGGGATGA	
SUJUT	1157	AGI IGGGCAC	CIAGGGAGACTGC	CGICGACAAGACO		CARCCARCE	CCCCCA
Spict	1160	ATCATCATGCC					
Ouerv	1217	GCCAACTCGCC	CCITALGTCCTGG	CCCTTAAAACCA	TCTCAG	TCGGATTGCA	GGCTGC
~ Sbict	1228	GCCAACTCCCC	AGAGTGCGCGAAT				GGCTGC
011070	1277	AACTCCCCCTCC	TATCAACCCCAAT	CCCTACTATCC	CCATCACI		
Query	12//						
sbjct	1288	AACTCGCCTGC	CAT'GAAGCCGGAAT	CGCTAGTAATCG(GGATCAG	CATGCCGCGG	TGAATA
Query	1337	CGTTCCCGGGT	CTTGTACACACCG	CCCGTCACACCAC	CGAGAGT1	TGCAACACCC	GAAGTC
Sbjct	1348	CGTTCCCGGGI	CTTGTACACACCG	CCCGTCACACCAC	GAGAGTI	TGCAACACCC	GAAGTC
Query	1397	GGTGAGGTAAC	CCGCAA-GAGCCAG	CCGCCGAAGG 1	L429		
Sbjct	1408	GGTGAGGTAA	CGCAAGGAGCCAG	CCGCCGAAGG 1	L441		



Bacillus subtilis EF488088.1

Bacillus subtilis strain QD9 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb[EF488088.1]</u> Length: 1412 Number of Matches: 1

Range 1: 366 to 1319 GenBank Graphics Vext Match A Previous Match							
Score	(1004)	Expect	Identities	Gaps	Strand		
1700 bits	s(1884)	0.0	950/954(99%)	1/954(0%)	Plus/Minus		
Query	1	AAACTCTCGTG	GTGTGACGGGCGGTG	TGTACAAGGCCC	GGGAACGTATTCACC	GCGGCAT	60
Sbjct	1319	AAACTCTCGTG	GTGTGACGGGCGGTG	TGTACAAGGCCC	GGGAACGTATTCACC	GCGGCAT	1260
Query	61	GCTGATCCGCG	ATTACTAGCGATTCC	AGCTTCACGCAG	TCGAGTTGCAGACTG	CGATCCG	120
Sbjct	1259	GCTGATCCGCG	ATTACTAGCGATTCC	AGCTTCACGCAG	TCGAGTTGCAGACTG	CGATCCG	1200
Query	121	AACTGAGAACA	GATTTGTGGGATTGG	CTTAACCTCGCG	GTTTCGCTGCCCTTT	GTTCTGT	180
Sbjct	1199	AACTGAGAACA	GATTTGTGGGATTGG	CTTAACCTCGCG	GTTTCGCTGCCCTTT	GTTCTGT	1140
Query	181	CCATTGTAGCA	CGTGTGTGTAGCCCAGG	TCATAAGGGGCA	TGATGATTTGACGTC	ATCCCCA	240
Sbjct	1139	CCATTGTAGCA	CGTGTGTAGCCCAGG	IIIIIIIIIIII TCATAAGGGGCA	 TGATGATTTGACGTC	ATCCCCA	1080
Query	241	CCTTCCTCCGG	TTTGTCACCGGCAGT	CACCTTAGAGTG	CCCAACTGAATGCTG	GCAACTA	300
Sbjct	1079	CCTTCCTCCGG	TTTGTCACCGGCAGT	CACCTTAGAGTG	CCCAACTGAATGCTG	 GCAACTA	1020
Query	301	AGATCAAGGGT	TGCGCTCGTTGCGGG	ACTTAACCCAAC	ATCTCACGACACGAG	CTGACGA	360
Sbjct	1019	AGATCAAGGGT	TGCGCTCGTTGCGGG	ACTTAACCCAAC	ATCTCACGACACGAG	 CTGACGA	960
Query	361	CAACCATGCAC	CACCTGTCACTCTGC	CCCCGAAGGGGA	CGTCCTATCTCTAGG	ATTGTCA	420
Sbjct	959	CAACCATGCAC	CACCTGTCACTCTGC	CCCCGAAGGGGA	CGTCCTATCTCTAGG	ATTGTCA	900
Query	421	GAGGATGTCAA	GACCTGGTAAGGTTC	TTCGCGTTGCTT	CGAATTAAACCACAT	GCTCCAC	480
Sbjct	899	GAGGATGTCAA	GACCTGGTAAGGTTC	TTCGCGTTGCTT	CGAATTAAACCACAT	GCTCCAC	840
Query	481	CGCTTGTGCGG	GCCCCCGTCAATTCC	TTTGAGTTTCAG	TCTTGCGACCGTACT	CCCCAGG	540
Sbjct	839	CGCTTGTGCGG	GCCCCCGTCAATTCC	 TTTGAGTTTCAG	IIIIIIIIIIIIIIII TCTTGCGACCGTACT [,]	CCCCAGG	780
Query	541	CGGAGTGCTTA	ATGCGTTAGCTGCAG	CACTAAGGGGCG	GAAACCCCCTAACAC'	TTAGCAC	600
Sbjct	779	 CGGAGTGCTTA	 ATGCGTTAGCTGCAG	 CACTAAGGGGGCG(GAAACCCCCTAACAC	 TTAGCAC	720
Query	601	TCATCGTTTAC	GGCGTGGACTACCAG	GGTATCTAATCC	IGTTCGCTCCCCACG	CTTTCGC	660
Sbjct	719	 TCATCGTTTAC	GGCGTGGACTACCAG	GGTATCTAATCC	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	 CTTTCGC	660
Query	661	TCCTCAGCGTC	AGTTACAGACCAGAG	AGTCGCCTTCGC	CACTGGTGTTCCTCC	ACATCTC	720
Sbjct	659	TCCTCAGCGTC	AGTTACAGACCAGAG	AGTCGCCTTCGC	CACTGGTGTTCCTCC	ACATCTC	600
Query	721	TACGCATTTCA	CCGCTACACGTGGAA	TTCCACTCTCCT	CTTCTGCACTCAAGT	TCCCCAG	780
Sbjct	599	TACGCATTTCA	CCGCTACACGTGGAA	TTCCACTCTCCT	CTTCTGCACTCAAGT	TCCCCAG	540
Query	781	TTTCCAATGAC	CCTCCCCGGTTGAGC	CGGGGGGCTTTCA	САТСАААСТТААААА	ACCGCCT	840
Sbjct	539	TTTCCAATGAC	CCTCCCCGGTTGAGC	CGGGGGGCTTTCA	CATCAGACTTAAGAA	ACCGCCT	480
Query	841	GCGAGCCCTTT.	ACGCCCAATAATTCC	GGACAACGCTTG	CCACCTACGTATTAC	CGCGGCT	900
Sbjct	479	GCGAGCCCTTT.	ACGCCCAATAATTCC	GGACAACGCTTG	CCACCTACGTATTAC	CGCGGCT	420
Query	901	GCTGGCACGTA	GTTAGCCGGGGCTTTC	TGGTTAGGTACC	GTC-AGGTACCGCCC	T 953	
Sbjct	419	GCTGGCACGTA	GTTAGCCGTGCTTTC	TGGTTAGGTACC	GTCAAGGTACCGCCC'	і Т 366	

Origonal Statistics (Complete genome
 Backisha subjicits strin SVA, Complete genome
 Backisha subjicits strin SVA, STAC, complete genome
 Backisha subjicits strin SVA, STAC, complete genome
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 Backisha subjicits strin SVA, Statist SVA, Back, partial sequence
 Backisha subjicits strin SVA, Statist SVA, Back, partial sequence
 Backisha subjicits strin SVA Statist SVA pere, partial sequence
 Backisha subjicits strin SVA Statist SVA pere, strint SSRIII
 Backisha subjicits partial IS (RNA pere, strint SSRIII)
 Backisha subjicits strin SNI IS (RNA pere, strint SSRIII)
 Backisha subjicits strin SNI IS (RNA pere, strint SSRIII)
 Backisha subjicits strin SNI IS (RNA pere, partial sequence)
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0.0001

Bacillus subtilis strain PR38 165 ribosomal RNA gene, partial sequence
 Bacillus tequilensis strain B-47 165 ribosomal RNA gene, partial sequence
 Bacillus tequilensis estimation and the sequence
 Bacillus sp. SSKSD8 165 ribosomal RNA gene, partial sequence
 Bacillus sp. SSKSD8 165 ribosomal RNA gene, partial sequence

8- The phylogenetic analysis of bacteria isolated from books and shelves in libraries and archive storerooms samples.

Pseudomonas jessenii LN774645.1

Pseudomonas jessenii partial 16S rRNA gene, isolate 1111MAR14N4 Sequence ID: <u>emb|LN774645.1|</u> Length: 1191 Number of Matches: 1

Range 1:	1 to 1028	GenBank Graphics		V Nex	kt Match 🔺 Previous Match	
Score 1775 bits	(1968)	Expect 0.0	Identities 1013/1028(99%)	Gaps 3/1028(0%)	Strand Plus/Plus	
Query	16	AATGCCTAGGA	ATCTGCCTATTAGT	GGGGGACAACGTTT	CGAAAGGAACGCTAATACCGC	: 75
Sbjct	1	AATGCCTAGGA	ATCTGCCTATTAGT		GAAAGGAACGCTAATACCGC	60
Query	76	ATACGTCCTAC	GGGAGAAAGCAGGG	GACCTTCGGGCCTTC	GCGCTAATAGATGAGCCTAGG	; 135
Sbjct	61	ATACGTCCTAC	GGGAGAAAGCAGGG	GACCTTCGGGCCTTC	CGCTAATAGATGAGCCTAG	; 120
Query	136	TCGGATTAGCI	AGTTGGTGAGGTAA	TGGCTCACCAAGGC	JACGATCCGTAACTGGTCTGA	195
Sbjct	121	TCGGATTAGCI	CAGTTGGTGAGGTAA	TGGCTCACCAAGGCO	JACGATCCGTAACTGGTCTGA	180
Query	196	GAGGATGATCA	AGTCACACTGGAACT	GAGACACGGTCCAG	ACTCCTACGGGAGGCAGCAGI	255
Sbjct	181	GAGGATGATCA	AGTCACACTGGAACT	GAGACACGGTCCAG	ACTCCTACGGGAGGCAGCAGI	240
Query	256	GGGGAATATTG	GACAATGGGCGAAA	GCCTGATCCAGCCA	IGCCGCGTGTGTGAAGAAGGI	315
Sbjct	241	GGGGAATATTO	GACAATGGGCGAAA	GCCTGATCCAGCCAT	IGCCGCGTGTGTGAAGAAGGI	300
Query	316	CTTCGGATTG	TAAAGCACTTTAAGT	TGGGAGGAAGGGTC	JTTGCCTAATACGTGACGGTI	375
Sbjct	301	CTTCGGATTGT	CAAAGCACTTTAAGT	TGGGAGGAAGGGTC	3TTGCCTAATACGTGACGGCI	360
Query	376	TTGACGTTAC	GACAGAATAAGCAC	CGGCTAACTCTGTGC	CAGCAGCCGCGGTAATACAG	; 435
Sbjct	361	TTGACGTTACC	CGACAGAATAAGCAC	CGGCTAACTCTGTGC	CAGCAGCCGCGGTAATACAG	420
Query	436	AGGGTGCAAGO	GTTAATCGGAATTA	CTGGGCGTAAAGCGG	CGCGTAGGTGGTTTGTTAAGI	495
Sbjct	421	AGGGTGCAAGC	GTTAATCGGAATTA	CTGGGCGTAAAGCGC	CGCGTAGGTGGTTTGTTAAGT	480
Query	496	TGGATGTGAAZ	ATCCCCGGGCTCAAC	CTGGGAACTGCATCO		555
Sbjct	481	TGGATGTGAAA	ATCCCCGGGCTCAAC	CTGGGAACTGCATCO	CAAAACTGGCAAGCTAGAGTA	540
Query	556	CGGTAGAGGGI	GGTGGAATTTCCTG	TGTAGCGGTGAAATO	CGTAGATATAGGAAGGAACA	615
Sbjct	541	CGGTAGAGGGI	GGTGGAATTTCCTG	TGTAGCGGTGAAATC	GTAGATATAGGAAGGAACA	600
Query	616	TCAGTGGCGAA			AGGTGCGAAAGCGTGGGGAG	675
Sbjct	601	TCAGTGGCGAA	GGCGACCACCTGGA	CTGATACTGACACTO	AGGTGCGAAAGCGTGGGGAG	660
Query	676	C-AACAGGATI	AGATACCCTGGTAG	TCCACGCCGT-AACG	JATGTCAACTAGCCGTTGGGA	733
Sbjct	661	CAAACAGGATT	AGATACCCTGGTAG	TCCACGCCGTAAACG	ATGTCAACTAGCCGTTGGGA	720
Query	734	TCCTTGAGATC	TTAGTGGCGCAGCT	AACGCATTAAGTTG <i>A</i>	ACCGCCTGGGGAGTACGGCCG	793
Sbjct	721	TCCTTGAGATO	TTAGTGGCGCAGCT.	AACGCATTAAGTTGA	1CCGCCTGGGGAGTACGGCCG	780
Query	794	CAAGGTTAAAA	CTCAAATGAATTGA	CGGGGGGCCCGCACAA	AGCGGTGGAGCATGTGGTTTA	853
Sbjct	781	CAAGGTTAAAA	ACTCAAATGAATTGA	CGGGGGGCCCGCACAA	AGCGGTGGAGCATGTGGTTTA	840
Query	854	ATTCGAAGCAA	ACGCGAAAAACCTTA	CCAGGCCTTGACATC	CAGTGAACTTTCCAGAAATG	913
Sbjct	841	ATTCGAAGCAA	CGCGAAGAACCTTA	CCAGGCCTTGACATC	CAGTGAACTTTCCAGAGATG	900
Query	914	GATTGGTGCC1	TCGGGAACACTGAA	ACAGGGGGCTGCATGG	CTGTCGTCAGCTCGGGTCGG	973
Sbjct	901	GATTGGTGCCI	"TCGGGAACACTGAG	ACAGGTGCTGCATGG	CTGTCGTCAGCTCGTGTCGT	960
Query	974	GAAATGTTGGG	FTTAAGTCCCGTAAC			1032
SDJCt	961 1022	GAGATGTTGGG	TTAAGTCCCGTAAC	GAGUGUAACCCTTGT	JUUTTAGTTAUCAGCACGTAA	1020
Query	1021	AGGGGGGGC 1	0.26			
ວມງບບ	TUZT	AGGIGGC 1	020			



Bacillus cereus KP192930.1

Bacillus cereus strain SHZ3.1 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KP192930.11</u> Length: 1419 Number of Matches: 1

Range 1: 334 to 1385 GenBank Graphics Vext Match 🛦 Previous Match								
Score 1824 bits	s(2022)	Expect 0.0	Identities 1039/1052(99%)	Gaps 4/1052(0	s)%) P	trand lus/Minus		
Query	1	CGGGTGTTAC	AAACTCTCGTGGTG	GACGGGCGG	TGTGTACA	AGGCCCGGGAA	CGTATTC	60
Sbjct	1385	CGGGTGTTAC	AAACTCTCGTGGTG	GACGGGCGG	TGTGTACA	AGGCCCGGGAA	CGTATTC	1326
Query	61	ACCGCGGCAT	GCTGATCCGCGATT	ACTAGCGATT	CCAGCTTC	ATGTAGGCGAG	TTGCAGC	120
Sbjct	1325	ACCGCGGCAT	GCTGATCCGCGATTA	ACTAGCGATT	CCAGCTTO	ATGTAGGCGAG	TTGCAGC	1266
Query	121	CTACAATCCG	AACTGAGAACGGTT	TATGAGATT	AGCTCCAC	CTCGCGGTCTT	GCAGCTC	180
Sbjct	1265	CTACAATCCG	AACTGAGAACGGTT	TATGAGATT	AGCTCCAC	CTCGCGGTCTT	GCAGCTC	1206
Query	181	TTTGTACCGT	CCATTGTAGCACGT	GTGTAGCCCA	GGTCATAA	GGGGCATGATG	ATTTGAC	240
Sbjct	1205	TTTGTACCGT	CCATTGTAGCACGT	GTGTAGCCCA	GGTCATAA	GGGGCATGATG	ATTTGAC	1146
Query	241	GTCATCCCCA	CCTTCCTCCGGTTT	GTCACCGGCA	GTCACCTI	AGAGTGCCCAA	CTTAATG	300
Sbjct	1145	GTCATCCCCA	CCTTCCTCCGGTTT	GTCACCGGCA	GTCACCTI	AGAGTGCCCAA	CTTAATG	1086
Query	301	ATGGCAACTA		GCTCGTTGCG	GGACTTAA		ACGACAC	360
Sbjct	1085	ATGGCAACTA	AGATCAAGGGTTGC	GCTCGTTGCG	GGACTTAA	CCCAACATCTC	ACGACAC	1026
Query	361	GAGCTGACGA	CAACCATGCACCAC	CTGTCACTCT	GCTCCCGA		TATCTCT	420
Sbjct	1025	GAGCTGACGA	CAACCATGCACCAC	CTGTCACTCT	GCTCCCGA	AGGAGAAGCCC	TATCTCT	966
Query	421	AGGGTTTTCA	GAGGATGTCAAGAC	CTGGTAAGGT	TCTTCGCG	TTGCTTCGAAT	TAAACCA	480
Sbjct	965	AGGGTTTTCA	GAGGATGTCAAGAC	CTGGTAAGGT	TCTTCGCG	TTGCTTCGAAT	TAAACCA	906
Query	481	CATGCTCCAC	CGCTTGTGCGGGCCC	CCCGTCAATT	CCTTTGAG	TTTCAGCCTTG	CGGCCGT	540
Sbjct	905	CATGCTCCAC	CGCTTGTGCGGGCCC	CCCGTCAATT	CCTTTGAG	TTTCAGCCTTG	CGGCCGT	846
Query	541	ACTCCCCAGG	CGGAGTGCTTAATG	CGTTAACTTC	AGCACTAA		CCTCTAA	600
Sbjct	845	ACTCCCCAGG	CGGAGTGCTTAATG	CGTTAACTTC	AGCACTAA	AGGGCGGAAAC	CCTCTAA	786
Query	601	CACTTAGCAC	TCATCGTTTACGGC(GTGGACTACC	AGGGTAT	CTAATCCTGTT1	GCTCCCC	660
Sbjct	785	CACTTAGCAC	TCATCGTTTACGGC	GTGGACTACC.	AGGGTAT	CTAATCCTGTTI	GCTCCCC	726
Query	661	ACGCTTTCGC	GCCTCAGTGTCAGT	FACAGACCAG	AAAGTCGO	CCTTCGCCACTG	GTGTTCC	720
Sbjct	725	ACGCTTTCGC	GCCTCAGTGTCAGT:	TACAGACCAG	AAAGTCG	CCTTCGCCACTG	GTGTTCC	666
Query	721	TCCATATCTC	TACGCATTTCACCG	CTACACATGG.	AATTCCAC	CTTTCCTCTTCI	GCACTCA	780
Sbjct	665	TCCATATCTC	TACGCATTTCACCG	CTACACATGG	AATTCCA	CTTTCCTCTTCI	GCACTCA	606
Query	781	AGTCTCCCAG	TTTCCAATGACCCT	CCACGGTTGA	GCCGTGG	GCTTTCACATCA	AACTTAA	840
Sbjct	605	AGTCTCCCAG	TTTCCAATGACCCT	CCACGGTTGA	GCCGTGG	GCTTTCACATCA	GACTTAA	546
Query	841	AAAACCACCT		CCCAATAATT	CCGGATAZ	ACGCTTGCCACC	TACGTAT	900
Sbjct	545	GAAACCACCT	GCGCGCGCTTTACG	CCCAATAATT	CCGGATA	ACGCTTGCCACC	TACGTAT	486
Query	901	TACCGCGGCT	GCTGGCACGTAGTT	AGCCG-GGCT'	TTCTGGT	TAGGTACCGTCA		959
Sbjct	485	TACCGCGGCT	GCTGGCACGTAGTT	AGCCGTGGCT	TTCTGGT	TAGGTACCGTCA	AGGTGCC	426
Query	960	AGCTTATTCA	ACTAGCACTTGTTC	TTCCCTAAC-2	ACAAAAT1	TTTACAACCCG-	AAGCCTT	1017
Sbjct	425	AGCTTATTCA	ACTAGCACTTGTTC	TTCCCTAACA	ACAAAATT	TTACGACCCGA	AAGCCTT	366
Query	1018	CCTCACTCCC		ICCAAATT	1048			
Sbjct	365	CATCACTCAC	GCGGCGTTGCTCCG	ICAGAATT	334			

Icl Query_84899	
	Bacillus thuringiensis strain VCRC B548 16S ribosomal RNA gene, partial sequence
۲	Bacillus tavanancis strain SH25.1 105 tibosonial RNA gene, partial sequence
	Bacillus for the final for the first state of the f
	Bacillus cereus partia 165 rRVA guit, suan D10
	Bacillus cereus partial 16S rRNA gene, strain B14
	Bacillus thuringiensis partial 16S rRNA gene. strain B16
	Bacillus thuringiensis partial 16S rRNA gene, strain B19
	Bacillus thuringiensis partial 16S rRNA gene, strain B20
	Bacillus cereus partial 16S rRNA gene, strain B22
	Bacillus cereus partial 16S rRNA gene, strain B23
	Bacillus cereus partial 10S rKNA gene, strain B24
	Bacillus Cercuis partial 105 fK/vA gene, strain B23
	Bacillus carrens partial 165 IRVA gene, strain D27
	Bacillus cerens partial 165 IRNA gene, strain B20
	Bacillus cereus partial 16S rRNA gene, strain B34
	Bacillus cereus partial 16S rRNA gene, strain B35
	Bacillus cereus partial 16S rRNA gene, strain B37
	Bacillus thuringiensis partial 165 rRNA gene, strain B40
	Bacillus cereus partial 10S rKNA gene, strain B43
	Bacillus cereus partial 105 rKNA gene, strain B44
	Bacillus cereus partial 165 IRNA gene, strain B40
	Bacillus Cercus patital 105 IRVA gene, strain Dro
	Bacillus cereus partial 165 rRNA gene strain B51
	Bacillus cereus partial 16S rRNA gene strain B53
	Bacillus cereus partial 16S rRNA gene, strain B57
	Bacillus cereus partial 16S rRNA gene, strain B59
	Bacillus cereus partial 16S rRNA gene, strain B60
	Bacillus cereus partial 16S rRNA gene, strain B61
	Bacillus cereus partial 105 fK/NA gene, strain B05
	Bacillus cereus partial 105 INNA gene, strain B67
	Bacillus cereus partial 165 rRNA gene strain B68
	Bacillus cereus partial 16S rRNA gene, strain B70
	Bacillus cereus partial 16S rRNA gene, strain B71
	Bacillus cereus partial 16S rRNA gene, strain B74
	PBacillus cereus partial 165 rRNA gene, strain B76
	Bacillus cereus partial 16S rRNA gene, strain B//
	Bacillus cereus partial 105 rKNA gene, strain B/9
	Bacillus celeus partial 105 fKNA gene, strain D81
	Bacillus cereus partial 105 INVA gene, strain B83
	Bacillus cereus partial 165 rRNA gene, strain B84
	Bacillus cereus partial 16S rRNA gene, strain B86
	Bacillus cereus partial 16S rRNA gene, strain B88
	Bacillus cereus partial 16S rRNA gene, strain B90
	Bacillus cereus strain F3-6 16S ribosomal RNA gene, partial sequence
	Bacillus cereus strain F3-34 10S ribosomal RNA gene, partial sequence
	Bacillus sp. CS3 105 nbosomal KNA gene, partial sequence
	V DACHIUS SD UL 14 105 TIDOSOMALININA GENE DATIAL SECTIONCE

Pantoea sp. CR30 165 ribosomal RNA gene, partial sequence Pantoea sp. CR30 165 ribosomal RNA gene, partial sequence firmicutes | 47 leaves

Bacillus altitudinis KT758615.1

Range 1: 27 to 1077 GenBank Graphics Vext Match 🔺 Previous Match					
Score 1817 bit	s(2014)	Expect Identitie 0.0 1036/10	as Gaps 052(98%) 3/1052(0%)	Strand) Plus/Plus	
Query	1	CCGGATGTTAGCGGCG	3ACGGGTGAGTAACACGTGGG	TAACCTGCCTGTAAGACTGGGA'	г 60
Sbjct	27	CCGGATGTTAGCGGCGG	ACGGGTGAGTAACACGTGGG	TAACCTGCCTGTAAGACTGGGA	Г Г 86
Query	61	AACTCCGGGAAACCGG	AGCTAATACCGGATAGTTCCT	TGAACCGCATGGTTCAAGGATG	A 120
Sbjct	87	AACTCCGGGAAACCGG	AGCTAATACCGGATAGTTCCT	TGAACCGCATGGTTCAAGGATG	A 146
Query	121	AAGACGGTTTCGGCTG	CACTTAYAGATGGACCCGCG	GCGCATTAGCTAGTTGGTGAGG	r 180
Sbjct	147	AAGACGGTTTCGGCTG	CACTTACAGATGGACCCGCG	GCGCATTAGCTAGTTGGTGAGG	Г Г 206
Query	181	AACGGCTCACCAAGGCO	ACGATGCGTAGCCGACCTGA	GAGGGTGATCGGCCACACTGGG	A 240
Sbjct	207	AACGGCTCACCAAGGC	ACGATGCGTAGCCGACCTGA	GAGGGTGATCGGCCACACTGGG	A 266
Query	241	CTGAGACACGGCCCAG	ACTCCTACGGGAGGCAGCAGT	AGGGAATCTTCCGCAATGGACG	A 300
Sbjct	267	CTGAGACACGGCCCAG	ACTCCTACGGGAGGCAGCAGT	AGGGAATCTTCCGCAATGGACG	A 326
Query	301	AAGTCTGACGGAGCAAG	CGCCGCGTGAGTGATGAAGGT	TTTCGGATCGTAAAGCTCTGTT	G 360
Sbjct	327	AAGTCTGACGGAGCAAC	CGCCGCGTGAGTGATGAAGGT	TTTCGGATCGTAAAGCTCTGTT	G 386
Query	361	TTAGGGAAGAACAAGT	CAAGAGTAACTGCTTGCACC	TTGACGGTACCTAACCAGAAAG	C 420
Sbjct	387	TTAGGGAAGAACAAGT	3CAAGAGTAACTGCTTGCACC	TTGACGGTACCTAACCAGAAAG	C 446
Query	421	CACGGCTAACTACGTG	CAGCAGCCGCGGTAATACGT	AGGTGGCAAGCGTTGTCCGGAA	г 480
Sbjct	447	CACGGCTAACTACGTGC	CAGCAGCCGCGGTAATACGT	AGGTGGCAAGCGTTGTCCGGAA	I I 506
Query	481	TATTGGGCGTAAAGGGG	TCGCAGGCGGTTTCTTAAGT	CTGATGTGAAAGCCCCCGGCTC	A 540
Sbjct	507	TATTGGGCGTAAAGGGC	CTCGCAGGCGGTTTCTTAAGT	CTGATGTGAAAGCCCCCGGCTC	A 566
Query	541	ACCGGGGGAGGGTCATTO	GAAACTGGGAAACTTGAGTG	CAGAAGAGGAGAGTGGAATTCC	A 600
Sbjct	567	ACCGGGGGAGGGTCATTO	GAAACTGGGAAACTTGAGTG	CAGAAGAGGAGAGTGGAATTCC	A 626
Query	601	CGTGTAGCGGTGAAATG	CGTAGAGATGTGGAGGAACA		3 <mark>660</mark> I
Sbjct	627	CGTGTAGCGGTGAAATG	CGTAGAGATGTGGAGGAACA	CCAGTGGCGAAGGCGACTCTCTC	686 €
Query	661	GTCTGTAACTGACGCTG	AGGAGCGAAAGCGTGGGGAG	CGAACAGGATTAGATACCCTGG	с 720 I
Sbjct	687	GTCTGTAACTGACGCTG	AGGAGCGAAAGCGTGGGGAG	CGAACAGGATTAGATACCCTGG	c 746
Query	721	AGTCCACGCCGTAAACG	ATGAGTGCTAAGTGTTAGGG	GGTTTCCGCCCCTTAGTGCTGC	¥ 780
Sbjct	747	AGTCCACGCCGTAAACG	ATGAGTGCTAAGTGTTAGGG	GGTTTCCGCCCCTTAGTGCTGCA	4 806
Query	781	GCTAACGCATTAAGCAC	TCCGCCTGGGGAGTACGGTC	GCAAGACTGAAACTCAAAGGAA	C 840
Sbjct	807	GCTAACGCATTAAGCAC	TCCGCCTGGGGAGTACGGTC	GCAAGACTGAAACTCAAAGGAA	C 866
Query	841	TGACGGGGGGCCCGCACA	AGCGGTGGAGCATGTGGTTT.	AATTCGAAGCAACGCGAAAAAC	C 900
Sbjct	867	TGACGGGGGGCCCGCACA	AGCGGTGGAGCATGTGGTTT.	AATTCGAAGCAACGCGAAGAAC	926
Query	901	TTACCAGGTCTTGACAT	CCTCTGACACCCTAAAGATA	GGGCTTTCCCTTC-GGGAAAGA(3 959 I
Sbjct	927	TTACCAGGTC-TGACA1	CCTCTGACACCCTAGAGATA	GGGCTTTCCCTTCGGGGACAGA	∋ 985
Query	960	TGACAGGGGGGGCATGGI	TGTCGTGAGCTCGGGTCGGG.		C 1019
Sbjct	986	TGACAGTGGTGCATGGI	TGTCGTCAGCTCTGGTCGTG.	AGATGTTGGGTTAAGTCCCGCA	c 1045
Query	1020	GAGCGCACCCTTGATCT	A-TTGCCAGCATTTA 105	0	
Sbjct	1046	GAGCGCAACCCTGATCI	AGTTGCCAGCATTTA 107	7	

Bacillus altitudinis strain HQB822 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KT758615.1</u>| Length: 1111 Number of Matches: 1

	«fimicutes 2 leaves	Buillin on 1001 100 -3
<u> </u>	frmicutes 3 leaves	Bacillas sp. 35B2 165 nb
	 Bacilus pumius strain SH-B11, complete genome Bacilus stratosphericus strain SHP1-12 16S ribosomal RNA cene, nartial sequence 	
	Bacillus aerophilus strain MER_TA_109 16S ribosomal RNA gene, partial sequence	
	 Bacilus aerophilus strain MER 81 16S ribosomal RNA gene, partial sequence Bacilus aerophilus strain ASMS8 5 16S ribosomal RNA gene, partial sequence 	
	 Bacillus acrophilus strain ASCOS2 7mm 165 ribosonial RNA gene, partial sequence 	
	 Bacillus pumilus strain JGTA-R5 15S ribosonal RNA gene, partial sequence 	
	 Bacillus pumilus strain N-3-1-2 16S ribosomal RNA gene, partial sequence Bacillus pumilus strain GP, 8 correlate oneone 	
	 Bacillus sp. strain WCH11 16S ribosomal RNA gene, partial sequence 	
	 Bacillus sp. strain YQH26 16S ribosomal RNA gene, partial sequence 	
	 Bacillus pumilus strain ZAQ1 16S ribosomal RNA gene, partial sequence Bacillus pumilus strain YT1 16S ribosomal RNA gene, partial sequence 	
	Bacillus safensis strain F77116 16S ribosonul RNA gene, partial sequence	
	 Bacillus pumilus strain CCM3 16S ribosomal RNA gene, partial sequence Buillus pumilus Area (Science (Science)) 	
	 Bacillus pumlus strain MB4 NIOT 165 ribosomal RNA gene, partial sequence 	
	 Bacillus pumilus strain PFR4 16S ribosomal RNA gene, partial sequence 	
	 Uncultured Bacillus sp. clone T22F7d15 16S nbosomal RNA gene, partial sequence Bacillus numilia aeros for 16S rPNA, martial sequence, straire CMCS89 	
	 Bacilus punius strain B10 16S ribosonal RNA gene, partial sequence 	
	 Bacillus pumilus strain SL9021 16S ribosomal RSA gene, partial sequence 	
	 Bacillus sp. LAMA 781 165 ribosomal RNA gene, partial sequence Bacillus sp. F64 165 ribosomal RNA gene, partial sequence 	
	 Bacillus sp. F65 16S ribosonial RNA gene, partial sequence 	
	 Bacillus sp. 2098 16S ribosomal RNA gene, partial sequence Bacillus and A2017 16S characterized RNA gene, partial sequence 	
	 Bacillus aprochilas strain 20E 16S ribosomal RNA cene, nartial sequence 	
	Bacillus sp. C4-1 16S ribosomal RNA gene, partial sequence	
	 Bacillus pumilus strain GR31 16S ribosomal RNA gene, partial sequence 	
	 Bacilus punitus strain GK55 105 ribosomal KNA gene, partial sequence Bacilus stratosphericus strain H76 16S ribosomal RNA gene, natial sequence 	
	 Bacillus aerophilus strain IARI-THW-17 16S ribosomal RNA gene, partial sequence 	
	 Bacillus pumilus strain JUN-15 16S ribosomal RNA gene, partial sequence Bacillus nu IU/N1 16S ribosomal RNA gene partial sequence 	
	 Bacillus sp. CZB7 16S ribosomal RNA gene, partial sequence 	
	Bacillus sp. BG-7-E4 16S ribosomal RNA gene, partial sequence	
	 Bacillus stratosphericus strain MCCC 1A00112 16S ribosomal RNA gene, partial sequence Bavillus stratosphericus strain BEB30 16S ribosomal RNA gene, partial sequence 	
	 Bacillus abitadinis strain AB4 16S ribosonal RNA gene, partial sequence 	
	 Uncultured Bacillus sp. clone DVBSD_D305 16S ribosomal RNA gene, partial sequence 	
	 Uncultured Bacillus sp. clone DVBSD D306 16S ribosomal RNA gene, partial sequence Uncultured Bacillus on clone DVBSD D311 16S ribosomal RNA gene, partial sequence 	
	 Racillus aeronhilus strain R-16 16S ribosomal RNA oene partial sequence 	
	 Bacillus pumilus strain B-38 16S ribosomal RNA gene, partial sequence 	
	Bacillus stratosphericus strain JQZST-2 16S ribosomal RNA gene, partial sequence	
	Uncultured Bacillus sp. clone HXQX-92 16S ribosomal RNA gene, partial sequence	
	Bacillus sp. S11713 16S ribosomal RNA gene, partial sequence	
	Bacillus pumilus strain BK14-3239 16S ribosomal RNA gene, partial sequence	
	Bacilius pumitus strain 16/aa 16S ribosomal KNA gene, partial sequence	
	Bacillus on P42(2014) 16S ribosomal PNA agree partial sequence	
	 Incultured bacterium clone ncd2808c10c1 16S ribosomal RNA eene nartial sequence 	
	Bacillus altitudinis strain VIT-RI4 16S ribosomal RNA gene, partial sequence	
	Uncultured Bacillus sp. gene for 16S ribosomal RNA, partial sequence, clone: KZtp6-42	
	Bacillus pumilus strain DZBT01 16S ribosomal RNA gene, partial sequence	
	Bacillus stratosphericus strain CL RH01 16S ribosomal RNA gene, partial sequence	
	Bacillus altitudinis strain 4BS5 16S ribosomal RNA gene, partial sequence	
	Bacilius altitudinis strain 556 165 ribosomal RNA gene, partial sequence	
	Bacillus pumitus strain FeKB-FL1404 10S ribosomal KNA gene, partial sequence	
	 Bacillus en L 1(2014) 16S ribosomal PNA gene, partial sequence 	
	 Bacillus sp. 1 PPL-SC1 16S ribosomal RNA eene, nartial sequence 	
	Geobacillus stearothermophilus strain PPL-SSC5 16S ribosomal RNA gene, partial sequence	
	Bacillus pumilus gene for 16S ribosomal RNA, partial sequence, strain: NKCM3201	
	Bacillus stratosphericus strain AMET 1601 16S ribosomal RNA gene, partial sequence	
	 Bacillus stratosphericus strain F-83 16S ribosomal RNA gene, partial sequence 	
	Bacilius pumilus strain Klu7 165 ribosonial RNA gene, partial sequence	
	Dacinus sp. JB4 105 nbosomai KNA gene, partial sequence Azomicilium linofatium partial 165 (PNA anna isolata Azom 12	
	Regiling nemiling strain SRR07 16S physical DNA ages exercise annual sectors	
	Bacillus sp CanS,107 16S ribosomal RNA gene, partial sequence	
	 Bacillus dratosphericus drain I9DMVR 16S ribosomal RNA eene martial source 	
	Geobacillus stearothermonbilus strain SBANSCu4 16S ribosomal RNA eene. nartial sequence	e
	Bacterium XX2 16S ribosomal RNA gene, partial sequence	7.2
	Bacterium LM4-3 16S ribosomal RNA gene, partial sequence	
	Bacterium F1-4 16S ribosomal RNA gene, partial sequence	
	Construction 1 10 Language	

Bacillus pumilus KP322017.1

Bacillus pumilus strain MK16 16S ribosomal RNA gene, partial sequence sequence ID: gb|KP322017.1| Length: 1469 Number of Matches: 1

Range 1: (Range 1: 64 to 1120 GenBank Graphics Vext Match 🔺 Previous Match					
Score 1799 bits	s(1994)	Expect 0.0	Identities 1038/1057(98%)	Gaps 6/1057(0%)	Strand Plus/Plus	
Query	1	CCCGGATGTTA	GCGGCGGACGGGTG	GTAACACGTGGGT	AACCTGCCTGTAAGACTGGGA	60
Sbjct	64	CCCGGATGTTA	GCGGCGGACGGGTGZ	AGTAACACGTGGGT	AACCTGCCTGTAAGACTGGGA	123
Query	61	TAACTCCGGGA	AACCGGAGCTAATAC	CCGGATAGTTCCTT	GAACCGCATGGTTCAAGGATG	120
Sbjct	124	TAACTCCGGGA	AACCGGAGCTAATAC	CGGATAGTTCCTT	GAACCGCATGGTTCAAGGATG	183
Query	121	AAAGACGGTTT	CGGCTGTCACTTAC	GATGGACCCGCGG	CGCATTAGCTAGTTGGTGGGG	180
Sbjct	184	AAAGACGGTTT	CGGCTGTCACTTAC	GATGGACCCGCGG	CGCATTAGCTAGTTGGTGGGG	243
Query	181	TAATGGCTCAC	CAAGGCGACGATGC	TAGCCGACCTGAG	AGGGTGATCGGCCACACTGGG	240
Sbjct	244	TAATGGCTCAC	CAAGGCGACGATGC	TAGCCGACCTGAG	AGGGTGATCGGCCACACTGGG	303
Query	241	ACTGAGACACG	GCCCAGACTCCTAC	GGAGGCAGCAGTA	GGGAATCTTCCGCAATGGACG	300
Sbjct	304	ACTGAGACACG	GCCCAGACTCCTAC	GGAGGCAGCAGTA	.GGGAATCTTCCGCAATGGACG	363
Query	301	AAAGTCTGACG	GAGCAACGCCGCGT	GAGTGATGAAGGTT	TTCGGATCGTAAAGCTCTGTT	360
Sbjct	364	AAAGTCTGACG	GAGCAACGCCGCGT	GAGTGATGAAGGTT	TTCGGATCGTAAAGCTCTGTT	423
Query	361	GTTAGGGAAGA	ACAAGTGCGAGAGT	ACTGCTCGCACCT	TGACGGTACCTAACCAGAAAG	420
Sbjct	424	GTTAGGGAAGA	ACAAGTGCGAGAGT	ACTGCTCGCACCT	TGACGGTACCTAACCAGAAAG	483
Query	421	CCACGGCTAAC	TACGTGCCAGCAGC	GCGGTAATACGTA	GGTGGCAAGCGTTGTCCGGAA	480
Sbjct	484	CCACGGCTAAC	TACGTGCCAGCAGC	GCGGTAATACGTA	.GGTGGCAAGCGTTGTCCGGAA	543
Query	481	TTATTGGGCGT	AAAGGGCTCGCAGGG	GGTTTCTTAAGTC	TGATGTGAAAGCCCCCGGCTC	540
Sbjct	544	TTATTGGGCGT	AAAGGGCTCGCAGG	GGTTTCTTAAGTC	TGATGTGAAAGCCCCCGGCTC	603
Query	541	AACCGGGGAGG	GTCATTGGAAACTG	GAAACTTGAGTGC	AGAAGAGGAGAGTGGAATTCC	600
Sbjct	604	AACCGGGGAGG	GTCATTGGAAACTG	GAAACTTGAGTGC	AGAAGAGGAGAGTGGAATTCC	663
Query	601	ACGTGTAGCGG	TGAAATGCGTAGAGA	ATGTGGAGGAACAC	CAGTGGCGAAGGCGACTCTCT	660
Sbjct	664	ACGTGTAGCGG	TGAAATGCGTAGAGA	ATGTGGAGGAACAC	CAGTGGCGAAGGCGACTCTCT	723
Query	661	GGTCTGTAACT	GACGCTGAGGAGCG		GAACAGGATTAGATACCCTGG	720
Sbjct	724	GGTCTGTAACT	GACGCTGAGGAGCGA	AAGCGTGGGGAGC	GAACAGGATTAGATACCCTGG	783
Query	721	TAGTCCACGCC	GTAAACGATGAGTG	CTAAGTGTTAGGGG	GTTTCCGCCCCTTAGTGCTGC	780
Sbjct	784	TAGTCCACGCC	GTAAACGATGAGTG	CTAAGTGTTAGGGG	GTTTCCGCCCCTTAGTGCTGC	843
Query	781	AGCTAACGCAT	TAAGCACTCCGCCT	GGGGAGTACGGTCG	CAAGACTGAAACTCAAAGGAA	840
Sbjct	844	AGCTAACGCAT	TAAGCACTCCGCCT	GGGAGTACGGTCG	CAAGACTGAAACTCAAAGGAA	903
Query	841	TTGACGGGGGC	CCGCACAAGCGGTG	GAGCATGTGGTTTA	ATTCGAAGCAACGCGAAAAAC	900
Sbjct	904	TTGACGGGGGG	CCGCACAAGCGGTG	GAGCATGTGGTTTA	ATTCGAAGCAACGCGAAGAAC	963
Query	901	CTTACCA-GTC	TTGACATCCTCTGA	с-ассстааааата	GGGCTTTCCCTTC-GGGACAG	957
Sbjct	964	CTTACCAGGTC	TTGACATCCTCTGA	CAACCCTAGAGATA	AGGGCTTTCCCTTCGGGGACAG	1023
Query	958	AGTGACAgggg	gggCATGGTTGTCG	rccgctcgggtcgg	GAAATGTTGGGTTAA-TCCCC	1016
Sbjct	1024	AGTGACAGGTG	I IIIIIIIIIIIII GTGCATGGTTGTCG	II IIII III FCAGCTCGTGTCGI	GAGATGTTGGGTTAAGTCCGC	1083
Query	1017	CACGAGCGC-A	CCCTTGATC-TAAT	IGCCAGCATCCA	1051	
Sbjct	1084	AACGAGCGCAA	CCCTTGATCTTAGT	IIIIIIIIIIII IGCCAGCATTCA	1120	

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🌳 🔶 Bacillus	umilus SAFR-032, complete genome
🌵 🏓 Bacilli	s pumilus strain W-N-5-3-2 16S ribosomal RNA gene, partial sequence
- 🔶 Bacte	rium BEL C12 16S ribosomal RNA gene, partial sequence
- Bac	llus pumilus 16S ribosomal RNA gene, partial sequence
Bac	llus pumilus strain m363 16S ribosomal RNA gene, partial sequence
Bac	llus pumilus strain m414 16S ribosomal RNA gene, partial sequence
Bac	illus pumilus strain O48 16S ribosomal RNA gene, partial sequence
Bac	illus pumilus strain YHE-31 16S ribosomal RNA gene partial sequence
Ba	illus pumilus strain VHH-2 16S ribosomal RNA gene, partial sequence
	ultured bacterium clone IF 31 16S ribosomal RNA gene, partial sequence
	illus pumilus strain K AP75 16S ribosomal RNA gene, partial sequence
	illus sp EIAT-22482 16S ribosomal RNA gene, partial sequence
	illus sp. FJA F-22462 105 fibosofiai KIVA gene, partial sequence
	thus pumilus strain HIDD 9209 105 fibosomal DNA gene, partial sequence
	thus pumitus strain HIDD 9900 105 fibosomal RNA gene, partial sequence
	thus pumilus strain IHDB 1400 165 ribosomai KINA gene, partial sequence
	tarium ABLIS 165 ribesemel BNA cone norticl sequence
	terium ARD18 105 ribosomai RNA gene, partial sequence
Dat	terium AK025 105 ribosomai KNA gene, partial sequence
Bac	illus pumitus strain LP02 01 165 ribosomal RNA gene, partial sequence
Bac	illus pumitus strain PW 116 165 ribosomal KNA gene, partial sequence
Bac	illus sp. Asnod6C12 partial 168 rKNA gene, isolate Asnod6C12
Bac	terium LX22 16S ribosomal RNA gene, partial sequence
Bac	illus pumilus partial I6S rRNA gene, isolate M11
Bac	illus sp. 801B1_12AR2A 16S ribosomal RNA gene, partial sequence
Bac	illus sp. 984B6_12ECASO 16S ribosomal RNA gene, partial sequence
Bac	illus sp. 991B6_12ER2A 16S ribosomal RNA gene, partial sequence
Bac	illus sp. 985B6_12ACASO 16S ribosomal RNA gene, partial sequence
Bac	illus sp. 988B6_12ACASO 16S ribosomal RNA gene, partial sequence
e Bac	illus sp. 995B6_12AR2A 16S ribosomal RNA gene, partial sequence
🗧 Bac	illus sp. 999B6_12AR2A 16S ribosomal RNA gene, partial sequence
Bac	illus pumilus 16S ribosomal RNA gene, partial sequence
Bac	illus safensis strain LOCK 1018 16S ribosomal RNA gene, partial sequence
Bac	illus sp. 7Kp1a 16S ribosomal RNA gene, partial sequence
Bac	illus sp. 13K7a2 16S ribosomal RNA gene, partial sequence
Bac	illus pumilus strain Bp108 16S ribosomal RNA gene, partial sequence
Bac	illus safensis strain SB-13 16S ribosomal RNA gene, partial sequence
🗢 Bac	illus safensis strain L26 16S ribosomal RNA gene, partial sequence
🔶 Un	ultured bacterium clone 958-4 16S ribosomal RNA gene, partial sequence
🔶 Una	ultured bacterium clone 958-5 16S ribosomal RNA gene, partial sequence
🔶 Uno	ultured bacterium clone 958-7 16S ribosomal RNA gene, partial sequence
🔶 Uno	ultured bacterium clone 958-12 16S ribosomal RNA gene, partial sequence
🔶 Uno	ultured bacterium clone 958-18 16S ribosomal RNA gene, partial sequence
• Uno	ultured bacterium clone 335-2 16S ribosomal RNA gene, partial sequence
Una Una	ultured bacterium clone 335-5 16S ribosomal RNA gene, partial sequence
i Una	ultured bacterium clone 335-18 16S ribosomal RNA gene, partial sequence
	ultured bacterium clone 335-21 16S ribosomal RNA gene, partial sequence
	alantea bacteriani cione bob-ari rob ribosoniari ra tra gene, partial sequence

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Bacillus stratosphericus KJ672335.1

Bacillus stratosphericus strain MUGA150 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KJ672335.1</u>| Length: 1404 Number of Matches: 1

Range 1:	ange 1: 326 to 1371 GenBank Graphics Vext Match 🔺 Previous Match					
Score 1781 bits	s(1974)	Expect 0.0	Identities 1026/1046(98%)	Gaps 7/1046(0%)	Strand Plus/Minus	
Query	1	TCTCGTGGT	TGACGGGCGGTGTC		ACGTATTCACCGCGGCATGCTG	60
Sbjct	1371	TCTCGTGGT	GTGACGGGCGGTGTC	GTACAAGGCCCGGGZ	ACGTATTCACCGCGGCATGCTG	1312
Query	61	ATCCGCGAT		GCTTCACGCAGTCGZ	AGTTGCAGACTGCGATCCGAACT	120
Sbjct	1311	ATCCGCGAT	TACTAGCGATTCCAC	GCTTCACGCAGTCGA	AGTTGCAGACTGCGATCCGAACT	1252
Query	121	GAGAACAGA	TTTGTGGGATTGGC	TAAACCTTGCGGTC1	FCGCAGCCCTTTGTTCTGTCCAT	180
Sbjct	1251	GAGAACAGA	TTTGTGGGATTGGC	TAAACCTTGCGGTC1	FCGCAGCCCTTTGTTCTGTCCAT	1192
Query	181	TGTAGCACG	IGTGTAGCCCAGGT	CATAAGGGGCATGAI	IGATTTGACGTCATCCCCACCTT	240
Sbjct	1191	TGTAGCACG	IGTGTAGCCCAGGT	CATAAGGGGCATGAI	IGATTTGACGTCATCCCCACCTT	1132
Query	241	CCTCCGGTT	IGTCACCGGCAGTC	ACCTTAGAGTGCCCZ	ACTGAATGCTGGCAACTAAGAT	300
Sbjct	1131	CCTCCGGTT	IGTCACCGGCAGTC	ACCTTAGAGTGCCCZ	ACTGAATGCTGGCAACTAAGAT	1072
Query	301	CAAGGGTTG	CGCTCGTTGCGGGA	CTTAACCCAACATCI	FCACGACACGAGCTGACGACAAC	360
Sbjct	1071	CAAGGGTTG	CGCTCGTTGCGGGA	CTTAACCCAACATCI	FCACGACACGAGCTGACGACAAC	1012
Query	361	CATGCACCA	CCTGTCACTCTGTCC	CCCGAAGGGAAAGTC	CCTATCTCTAGGGTTGTCAGAGG	420
Sbjct	1011	CATGCACCAG	CCTGTCACTCTGTCC	CCCGAAGGGAAAGCC	CCTATCTCTAGGGTTGTCAGAGG	952
Query	421	ATGTCAAGA	CCTGGTAAGGTTCT	ICGCGTTGCTTCGAZ	ATTAAACCACATGCTCCACCGCT	480
Sbjct	951	ATGTCAAGAG	CCTGGTAAGGTTCT	ICGCGTTGCTTCGAZ	ATTAAACCACATGCTCCACCGCT	892
Query	481	TGTGCGGGC	CCCCGTCAATTCCT	ITGAGTTTCAGTCTI	IGCGACCGTACTCCCCAGGCGGA	540
Sbjct	891	TGTGCGGGC	CCCCGTCAATTCCT	TGAGTTTCAGTCTI	IGCGACCGTACTCCCCAGGCGGA	832
Query	541	GTGCTTAAT	GCGTTAGCTGCAGC	ACTAAGGGGCGGAAZ	ACCCCCTAACACTTAGCACTCAT	600
Sbjct	831	GTGCTTAAT	GCGTTAGCTGCAGC	ACTAAGGGGCGGAAZ	ACCCCCTAACACTTAGCACTCAT	772
Query	601	CGTTTACGG		GTATCTAATCCTGT		660
Sbjct	771	CGTTTACGG	CGTGGACTACCAGG	GTATCTAATCCTGT:	TCGCTCCCCACGCTTTCGCTCCT	712
Query	661	CAGCGTCAG		GTCGCCTTCGCCAC	IGGTGTTCCTCCACATCTCTACG	720
Sbjct	711	CAGCGTCAG	TTACAGACCAGAGAG	GTCGCCTTCGCCAC	IGGTGTTCCTCCACATCTCTACG	652
Query	721	CATTTCACCO	GCTACACGTGGAAT	FCCACTCTCCTCTT	CTGCACTCAAGTTTCCCAGTTTC	780
Sbjct	651	CATTTCACCO	GCTACACGTGGAAT	FCCACTCTCCTCTT	CTGCACTCAAGTTTCCCAGTTTC	592
Query	781	CAATGACCC	FCCCCGGTTGAGCC	GGGGGCTTTCACAT	CAAACTTAAAAAACCGCCTGCGA	840
Sbjct	591	CAATGACCC	rccccggttgagcc	GGGGGCTTTCACAT	CAGACTTAAGAAACCGCCTGCGA	532
Query	841	GCCCTTTAC	SCCCGATAATTCCCC	GGACAACGCTTGCC	ACCTACGTATTACCGCGGCTGCT	900
Sbjct	531	GCCCTTTAC	IIII IIIIIIII GCCCAATAATTCCCC	 GGACAACGCTTGCCA	ACCTACGTATTACCGCGGCTGCT	472
Query	901	GGCACGTAT	TTAGCCGTGGCTTT	CTGGTTAGGTACCG	ICAAGGGGCAAGCAGTTACTCTT	960
Sbjct	471	GGCACGTAG			I I I I I I I I I I I I I I I I I I I	412
Query	961	GCACTTGTT	CTTCCCTACACAA	AAACTTTACAATCC-	-AAAACCTTCATCCCTC-CGCGG	1016
Sbjct	411	GCACTTGTT	<mark> </mark> CTTCCCTAACAACA(GAGCTTTACGATCC	JAAAACCTTCATCACTCACGCGG	352
Query	1017	CGTTGCT-C	GTCAAATTCCTC	CAT 1039		
Sbjct	351	CGTTGCTCC	 GTCAGACTTTCGTC	 CAT 326		

	Bacillus aerophilus strain MER_18 16S ribosomal RNA gene, partial sequence	▼ sci/Query_134
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	firmicutes [7 leaves	
	Bacterium ST3(2015) strain ST3 16S ribosomal RNA gene, partial sequence Bacilus aerophilus strain IHBB 11116 16S ribosomal RNA gene, partial sequence	
	Bacillus pumilus strain MD-A 19 16S ribosomal RNA gene, partial sequence	
	 Endophytic bacterium SV686 165 rhosomal RNA gene, partial sequence Bacillus on UVERATS 165 rhosomal RNA gene, partial sequence 	
	Bacillus sp. ADM/sz 105 ribosonal RNA gene, partial sequence	
	Bacillus so, ADNK45 105 ribosomal RNA gene, partial sequence Bacillus so, ADNK21 105 ribosomal RNA gene, partial sequence	
	Bacilla screus strain FMDD 3 165 ribosonia RNA gene, partial sequence	
	Bacillus pumilus strain DB9-2 16S ribosomal RNA gene, parial sequence Bacillus pumilus train W1A to 1.1 16S interesting PDA gene activity constraints	
	Bacillus punitus statu (xxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxx	
	Uncultured bacterium clono 958-6 165 ribosomal RNA gene, partial sequence	
	Character Determiner Soft 2002, https://doi.org/10.1116/j.ecu.partial.sequence Bacillus pumilius strain NS-31165 ribosoma IRAA gene, partial sequence	
	Bacillus sp. 16Kp3 165 ribosomal RNA gene, partial sequence	
	Bachus 9, 3 M (24 ros nobsonia) (NY gene, partial sequence	
	Bacillus sp. 21K5e 16S ribosomal RNA gene, partial sequence	
	Bacillus sg, 11K 19 105 mbsonial RNA gene, partial sequence Bacillus sfratoshericus sfrata ISBN 1165 mbsonial RNA gene, partial sequence	
	Bacillus punilus partial 16S rRNA gene, strain GRC, isolate 20	
	Bacilus pundus strain CRAIS IOS indosomal RNA gene, partial sequence Bacilus avendulus etrain CI 19 IOS ribosomal RNA eren partial sequence	
	Bacillus aerius strain CLA 165 ribosomal RNA gene, pariai sequence	
	Bacillus pumilus strain CL3 16S ribosomai RNA gene, partai sequence Bacillus detucontricus partial 16S (PAN sense strain B80	
	Bacillus acrius strain SBANSCOR 165 friedman RNA gene, partial sequence	
	 Bacilus punitus strain CanR-21 165 ribosomal RNA gene, partial sequence Bacilus polynais train PLC 11156 ribosomal RNA gene, partial sequence 	
	Bacillus safenis strain RSC 11 105 ribosomal RNA gene, partial sequence	
	Bacillus sa fensis strain RIMT. 109 16S ribosomal RNA gene, partial sequence Bacillus esfemic train RIMT. 101 16S ribosomal RNA gene, partial sequence	
	Bacillus altitudinis strain Bacillus III for hostination RA gene, partial sequence Bacillus altitudinis strain Bacillus III for hostinational RA gene, partial sequence	
	Bacillus pumilus strain ASR-31 165 ribosomal RNA gene, partial sequence Bacillus paraphilus crimi IUII 61 165 ribosomal RNA gene partial sequence	
	Bacturas arcopanius sinain nenos 11100 105 missioniai Acce gene, partial sequence Bacturas arcopanius sinain 111108 9411 1053 missioniai RNA gene, partial sequence	
	 Bacillus attitudinis strain SBB27 165 ribosomal RNA gene, partial sequence Ministrum aunification for descent reaction accurate sequence 	
	Bacillus stratosional revo gene, parta sequence Bacillus stratosional RCVA (gene, parta) sequence	
	 Bacterium M1(2015b) strain M1 16S ribosomal RNA gene, partial sequence 	
	Bachus annualmis strain Kr0727 to S hosomal KNA gene, partial sequence Bachus annualmis strain Kr0727 to S hosomal KNA gene, partial sequence	
	Bacillus sp. Q11 16S ribosomal RNA gene, partial sequence	
	• Dacinis annuolinis strain 11.05 ribixoma KAA gene, partiai sequence	
	Bacillus sp. CICC 23870 16S ribosomal RNA gene, partial sequence	
	 Bacillas sp. W5(2015) 165 ribosomal RNA gene, partial sequence Bacillas quering textural IXAL/DD1/SC ideograf RNA series partial sequence 	
	Endophylic bacterium SV8871655 inosonia Rox gene, partal sequence Endophylic bacterium SV8871655 inosonia Rox gene, partal sequence	
	 Uncultured Bacillus sp. clone YXY-11 IoS ribosomia RNA gene, partial sequence Residue de statemente ettatis (RES) (St. St. Statement, RNA gene, partial sequence 	
	Bacillus stratiospiericus strain RVOSC 1020 1030 into strational RVA gene, partai sequence Bacillus pumilus isolate XXXV-1116S ribosomal RVA eene, partai sequence	
	Bacillus sp. JXRH25 16S ribosomal RNA gene, partial sequence	
	Bacillus sp. JAL24 16S rhosomal KNA gene, partial sequence Bacillus sp. DBL7331 16S rhosomal RNA erene, partial sequence	
	Uncultured Bacillus sp. clone DDGJ11 16S ribosomal RNA gene, partial sequence	
	 Bacilus pumilus strain BS25 165 ribosomal RNA gene, partial sequence Bacilus reformation and the sequence 	
	 Bacillus stratophericus strain LGR-6 165 ribosonal RNA gene, partial sequence 	
o	 Bacillus actius strain LOS-116S ribosomal RNA energy aprila sequence Bacillus activitatio IDB / Col 116S ribosomal RNA pana, partial sequence 	
	Bachia pumitus suain FT-50.14 ToS indoorna KVA gene, partai sequence Bachia pumitus strain GS-305 IS6 ribosonal RVA eene, partai sequence	
	 Bacillus pumilus strain GW-605 16S ribosomal RNA gene, partial sequence 	
	Bacillas 9, 185-9 165 ribosonial KNA gene, partial sequence	
	 Bacillus altitudinis strain IIIB B 15457 16S ribosomal RNA gene, partial sequence 	
	 Bacillus actius strain LLS.IE(B)-1105 ribosomal RNA gene, partial sequence Bacillus activitation 1231 (2011) 115 ribosomal RNA gene, partial sequence 	
	Bachts parallels strating 57 Prior 105 Prosonal RAA gene, partial sequence Bachts parallels prior strating 6729 165 Priosonal RAA gene, partial sequence	
	Bacillus altitudinis strain BCX-57 16S ribosomal RNA gene, partial sequence	
	 Bacillus statospiericus statu LAMA 11/2 105 ribosomai Riva gene, partiai sequence Bacillus statospiericus statu LAMA 1114 165 ribosomai RNA eene, partiai sequence 	
	Bacillus sp. LTW29 16S ribosomal RNA gene, partial sequence	
	 Bacillus sp. WP-A-6 16S ribosomal KNA gene, partial sequence Bacillus sp. hb58 16S ribosomal RNA gene, partial sequence 	
	firmicutes 4 leaves	
	Bacillus sp. hb117 16S ribosomal RNA gene, partial sequence	
	Bacillus altitudinis strain -Y 15 16S ribosomal RNA gene, partial sequence	
	Bacillus pumilus strain JK-SX001 16S ribosomal RNA gene, partial sequence	
	 Bacillus aerophilus strain Vanz 10S ribosomai RNA gene, partial sequence formientes 12 leaves 	
	740402-042312-0423	
Bacillus weihenstephanensis strain A2-25c-6b 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KC527665.1</u>] Length: 1252 Number of Matches: 1

Range 1:	161 to 11	97 GenBank Grap	hics		V Next	Match 🔺 Previous N	1atch	
Score 1750 bits	s(1940)	Expect 0.0	Identities 1016/1038(9	Gaps 98%) 7/1038(0%)	Strand Plus/Minus		
Query	1	CAAACTCTCG	TGGTGTGAC	GGGCGGTGTGTACA		GGAACGTATTCA	CCGCGGCA	60
Sbjct	1197	CAAACTCTCG	GGTGTGAC	GGGCGGTGTGTACA	AGGCCCG	GGAACGTATTCA	CCGCGGCA	1139
Query	61	TGCTGATCCG	CGATTACTA	GCGATTCCAGCTTC	ATGTAGG	CGAGTTGCAGCC	TACAATCC	120
Sbjct	1138	TGCTGATCCG	CGATTACTA	GCGATTCCAGCTTC	ATGTAGG	CGAGTTGCAGCC	TACAATCC	1079
Query	121	GAACTGAGAA	CGGTTTTAT	GAGATTAGCTCCAC	CTCGCGG	CTTGCAGCTCT	TTGTACCG	180
Sbjct	1078	GAACTGAGAA	CGGTTTTAT	GAGATTAGCTCCAC	CTCGCGG	ICTTGCAGCTCT	TTGTACCG	1019
Query	181	TCCATTGTAG			GGGGCAT	GATGATTTGACG	TCATCCCC	240
Sbjct	1018	TCCATTGTAG	CACGTGTGT	AGCCCAGGTCATAA	GGGGCAT	GATGATTTGACG	TCATCCCC	959
Query	241	ACCTTCCTCC	GGTTTGTCA		AGAGTGC	CCAACTTAATGA	TGGCAACT	300
Sbjct	958	ACCTTCCTCC	GGTTTGTCA	CCGGCAGTCACCTT	AGAGTGC	CCAACTTAATGA	TGGCAACT	899
Query	301	AAGATCAAGG	GTTGCGCTC	GTTGCGGGACTTAA	CCCAACA	CTCACGACACG	AGCTGACG	360
Sbjct	898	AAGATCAAGG	GTTGCGCTC	GTTGCGGGACTTAA	CCCAACA	ICTCACGACACG	AGCTGACG	839
Query	361	ACAACCATGC	ACCACCTGT		AGGAGAA	GCCCTATCTCTA	GGGTTTTC	420
Sbjct	838	ACAACCATGC	ACCACCTGT	CACTCTGCTCCCGA	AGGAGAA	GCTCTATCTCTA	GAGTTTTC	779
Query	421	AGAGGATGTC	AAGACCTGG	TAAGGTTCTTCGCG		GAATTAAACCAC	ATGCTCCA	480
Sbjct	778	AGAGGATGTC	AAGACCTGG	TAAGGTTCTTCGCG	TGCTTC	GAATTAAACCAC	ATGCTCCA	719
Query	481	CCGCTTGTGC	GGGCCCCCG	ICAATTCCTTTGAG	TTTCAGC	CTTGCGGCCGTA	CTCCCCAG	540
Sbjct	718	CCGCTTGTGC	GGGCCCCCG	TCAATTCCTTTGAG	TTTCAGC	CTTGCGGCCGTA	CTCCCCAG	659
Query	541	GCGGAGTGCT	TAATGCGTT	AACTTCAGCACTAA		AAACCCTCTAAC	ACTTAGCA	600
Sbjct	658	GCGGAGTGCT	TAATGCGTT	AACTTCAGCACTAA	AGGGCGGZ	AAACCCTCTAAC	ACTTAGCA	599
Query	601	CTCATCGTTT			TAATCCT			660 530
SDJCt	598	CCCCTCATCGTTT	TACGGCGTGG.	GACCAGADAGECCC	CTTCCC	ACTECTECE	CCATATCE	720
Sbict	538	CGCCTCAGIG	TCAGTTACA	GACCAGAAAGICGC	CTTCGCC	ACTGGTGTTCC	CCATATCT	479
Query	721	CTACGCATTT	CACCGCTAC	ACATGGAATTCCAC	TTTCCTC	TTCTGCACTCA	AGTCTCCCA	780
Sbjct	478	 CTACGCATTT	CACCGCTAC	 ACATGGAATTCCAC	 TTTCCTC	 TTCTGCACTCAZ	GTCTCCCA	419
Query	781	GTTTCCAATG	ACCCTCCAC	GGTTGAGCCGTGGG	CTTTCAC	атсааасттаа-	AAACCACC	839
Sbjct	418	GTTTCCAATG	ACCCTCCAC	GGTTGAGCCGTGGG	CTTTCAC	ATCAGACTTAA	JAAACCACC	359
Query	840	TGCGCGCGCT	TTACGCCCA	ATAATTCCGGA-AA	CGCTTGC	CACCTACGTAT	ACCGCGGC	898
Sbjct	358	TGCGCGCGCT	TTACGCCCA	ATAATTCCGGATAA	CGCTTGC	CACCTACGTATI	ACCGCGGC	299
Query	899	TGCTGGCACG	TAATTAGCC	G-GGCTTTCTGGTT. 	AGGTACC	GTCAGGGGGCCAG	NTTATTCA	957
Sbjct	298	TGCTGGCACG	TAGTTAGCC	GTGGCTTTCTGGTT.	AGGTACC	GTCAGGTGCCAG	CTTATTCA	239
Query	958	ACTAGCATTT	GTTCTTCCT	-ACACAAATTTTAC	-AACCGA	AGCTTCATCACI	C-CNCGCG	1014
Sbjct	238	ACTAGCACTT	GTTCTTCCT	AACACAGAGTTTAC	GACCCGA	AGCTTCATCACI	CACGCGCG	179
Sbjct	178	TGCTCGTCAA	AATTCOTC ACTTCGTC	161				



Acinetobacter lofwii KT387352.1

Acinetobacter Iwoffii strain CL_102 16S ribosomal RNA gene, partial sequence sequence ID: $\underline{gb|KT387352.1|}$ Length: 1437 Number of Matches: 1

Range 1: 4	425 to 13	56 GenBank Graphics	Vext Next 1	Match 🔺 Previous Match	
Score 1640 bits	(1818)	Expect Identities 0.0 923/932(99%)	Gaps S 0/932(0%) P	i trand Ilus/Minus	
Query	1	GTACAAGGCCCGGGAACGTATTCA	CGCGGCATTCTGATCC	GCGATTACTAGCGAT	CCG 60
Sbjct	1356	GTACAAGGCCCGGGAACGTATTCA	CGCGGCATTCTGATCC	GCGATTACTAGCGAT	CCG 1297
Query	61	ACTTCATGGAGTCGAGTTGCAGAC	CCAATCCGGACTACGA	TCGGCTTTTTGAGATI	TAGC 120
Sbjct	1296	ACTTCATGGAGTCGAGTTGCAGAC	CCAATCCGGACTACGA	TCGGCTTTTTGAGAT	TAGC 1237
Query	121	ATCCTCTCGCGAGGTAGCAACCCT	TGTACCGACCATTGTA	GCACGTGTGTAGCCC	IGGT 180
Sbjct	1236	ATCCTCTCGCGAGGTAGCAACCCT	TGTACCGACCATTGTA	GCACGTGTGTAGCCC	IGGT 1177
Query	181	CGTAAGGGCCATGATGACTTGACG	CGTCCCCGCCTTCCTC	CAGTTTGTCACTGGC	AGTA 240
Sbjct	1176	CGTAAGGGCCATGATGACTTGACG	CGTCCCCGCCTTCCTC	CAGTTTGTCACTGGC	AGTA 1117
Query	241	TCCTTAAAGTTCCCGGCTTAACCCC	CTGGCAAATAAGGAAA	AGGGTTGCGCTCGTT	GCGG 300
Sbjct	1116	TCCTTAAAGTTCCCGGCTTAACCCC	GCTGGCAAATAAGGAAA	AGGGTTGCGCTCGTT	GCGG 1057
Query	301	GACTTAACCCAACATCTCACGACA	CGAGCTGACGACAGCCA	TGCAGCACCTGTATG	TAAG 360
Sbjct	1056	GACTTAACCCAACATCTCACGACAC	CGAGCTGACGACAGCCA	TGCAGCACCTGTATG	TAAG 997
Query	361	TTCCCGAAGGCACCAATCCATCTC	GGAAAGTTCTTACTAT	GTCAAGACCAGGTAAG	GTT 420
Sbjct	996	TTCCCGAAGGCACCAATCCATCTC	GGAAAGTTCTTACTAT	GTCAAGACCAGGTAAG	GTT 937
Query	421	CTTCGCGTTGCATCGAATTAAACCA	CATGCTCCACCGCTTG	TGCGGGCCCCCGTCA	ATTC 480
Sbjct	936	CTTCGCGTTGCATCGAATTAAACCA	CATGCTCCACCGCTTG	TGCGGGCCCCCGTCA	ATTC 877
Query	481	ATTTGAGTTTTAGTCTTGCGACCG	ACTCCCCAGGCGGTCT	ACTTATCGCGTTAGC	rgcg 540
Sbjct	876	ATTTGAGTTTTAGTCTTGCGACCG	ACTCCCCAGGCGGTCT	ACTTATCGCGTTAGC	IGCG 817
Query	541	CCACTAAAGCCTCAAAGGCCCCCAA	GGCTAGTAGACATCGT	TTACGGCATGGACTAC	CCAG 600
Sbict	816	CCACTAAAGCCTCAAAGGCCCCAA	CGGCTAGTAGACATCGT	TTACGGCATGGACTAC	CCAG 757
Query	601	GGTATCTAATCCTGTTTGCTCCCCA	TGCTTTCGCACCTCAG	TGTCAGTATTAGGCCA	AGAT 660
Sbjct	756	GGTATCTAATCCTGTTTGCTCCCCA	ATGCTTTCGCACCTCAG	TGTCAGTATTAGGCCA	AGAT 697
Query	661	GGCTGCCTTCGCCATCGGTATTCC	CCAGATCTCTACGCAT	TTCACCGCTACACCTG	GAA 720
Sbjct	696	GGCTGCCTTCGCCATCGGTATTCC	CCAGATCTCTACGCAT	TTCACCGCTACACCTG	GAA 637
Query	721	TTCTACCATCCTCTCCCATACTCT	GCCAACCAGTATCGAA	TGCAATTCCCAAGTTA	AGC 780
Sbjct	636	TTCTACCATCCTCTCCCATACTCT	GCCAACCAGTATCGAA	TGCAATTCCCAAGTTA	AGC 577
Query	781	TCGGGGATTTCACATTTGACTTAA	TGGCCACCTACGCGCG	CTTTACGCCCAGAAAA	ATCC 840
Sbjct	576	TCGGGGATTTCACATTTGACTTAAT	TGGCCACCTACGCGCG	CTTTACGCCCAGTAAA	ATCC 517
Query	841	GATTAACGCTTGCACCCTCTGTAT	ACCGCGGCTGCTGGCA		TTA 900
Sbjct	516	GATTAACGCTTGCACCCTCTGTATT	ACCGCGGCTGCTGGCA	CAGAGTTAGCCGGTGC	TTA 457
Query	901	TTCTGCAAATAACGTCCACTATCCA	AAAATAT 932		
Sbjct	456	TTCTGCGAGTAACGTCCACTATCCA	AGAGTAT 425		

Uncultured bacterium clone A-38 16S ribosomal RNA gene, partial sequence		
Ť	Acinetobacter Iwoffin strain JCM 6840 165 ribosomal RNA gene, partial sequence	
	Gamma proteobacterium F3 a5 16S ribosomal RNA gene, partial sequence	
	Prolinoborus fasciculus strain MB-11 16S ribosomal RNA gene, partial sequence	
	Prolinoborus fasciculus strain MG-3 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain MR-13 ToS ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain 263ZG5 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain WG-70 16S ribosomal KNA gene, partial sequence	
	Acinetobacter Iwoffii strain WR-168 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain WB-203 I6S ribosomal RNA gene, partial sequence Prolinohogus fasciculus strain WL-242 I6S ribosomal RNA gene, partial sequence	
	Prolinoborus fasciculus strain WTB-120 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain AM-72 16S ribosomal RNA gene, partial sequence Prolinghouse forginal usataria TP, 142 16S ribosomal RNA gene, partial sequence	
	Profinoborus fasciculus strain 1B-142 16S ribosomal RNA gene, partial sequence Acinetobacter lwoffii strain AB-185 16S ribosomal RNA gene, partial sequence	
	Acinetobacter sp. LHG-tBW4 partial 16S rRNA gene, strain LHG-tBW4	
	Acinetobacter Iwoffii strain M0209 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain MTB-4-1 16S ribosomal RNÅ gene, partial sequence	
	Acinetobacter Iwoffii strain MTA-4 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain HA 522 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone XJDN-640 16S ribosomal RNA gene, partial sequence Bacterium N2 3 16S ribosomal RNA gene, partial sequence	
	Endophytic bacterium SV900 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain SBB32 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain WD206 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain LAHP2-7 16S ribosomal RNA gene, partial sequence	
	Cincultured bacterium clone 958-19 165 ribosomal KNA gene, partial sequence	
	Acinetobacter Iwoffii strain CL 102 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain WD224 16S ribosomal RNA gene, partial sequence Discultured Acinetobacter on clone TCCC 11167 16S ribosomal RNA gene partial sequence.	
	Prolinoborus fasciculus strain IHBB 9208 16S ribosomal RNA gene, partial sequence	
	Acinetobacter sp. Sn-19 16S ribosomal KNA gene, partial sequence	
	Acinetobacter lwoffi strain MTA-33 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain MTA-2 16S ribosomal RNA gene, partial sequence	
	Acinetobacter sp. Tibet-YD4820-5 16S ribosomal RNA gene, partial sequence	
	Acinetobacter sp. LHG-2BW3 partial 16S rRNA gene, strain LHG-2BW3	
	Acinetobacter Iwoffii strain AB-207 105 fibosonia RNA gene, partial sequence	
	Acinetobacter lwoffii strain TB-128 16S ribosomal RNA gene, partial sequence	
	Acinetotoscler livollu strain AL-11 16S ritosomal RNA gene, partial sequence Prolinohomis fasciculus strain WR-169 16S ritosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain WL-228 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain WL-199 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwofii strain WG-92 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain 265ZG8 16S ribosomal RNA gene, partial sequence	
	Acinetobacter sp. Car-01 10S motsonial R/As gene, partial sequence	
	Prolinoborus fasciculus strain MB-14 16S ribosomal RNA gene, partial sequence	
	Profinoborus fasciculus straim MTA-24-1 105 ribosomal KNA gene, partial sequence	
	Gamma proteobacterium F3_a10 16S ribosomal RNA gene, partial sequence	
	Bacterium C50(2013) 16S ribosomal RNA gene, partial sequence	• Acinetobacter Iwoffii partial 16S rPNA sense strain Marcailla. D786
	Acinetobacter lwoffii strain T24 16S nibosomal RNA gene, partial sequence	reasonable inviti pittan 105 Row gene, stant matselle P/80
	Acinetobacter lwoffi strain WD238 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone 3R2 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain KAR20 16S ribosomal RNA gene, complete sequence	
	Acinetobacter Iwoffii strain ZJY-//8 16S ribosomal RNA gene, partial sequence Acinetobacter Iwoffii strain SDI1 16S ribosomal RNA eene, partial sequence	
	Acinetobacter Iwoffii partial 16S rRNA gene, isolate 10	
	Acinetobacter Iwoffii strain 263AY7 16S ribosomal RNA gene, partial sequence Definohomic facionalis ctrain WR, 139.16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain MTB-12 16S ribosonal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain ML-39 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain 272XG8 16S ribosomal RNA oene nartial sequence	
	Acinetobacter Iwoffii strain AB-204 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain AR-180 16S ribosomal RNA gene, partial sequence	
	Acinetobacter Iwoffii strain TB-122 16S ribosomal RNA gene, partial sequence	
	Prolinoborus fasciculus strain AL-25 105 ribosomal KNA gene, partial sequence	
	Prolinoborus fasciculus strain WL-241 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain WB-216-2 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain WM-65 16S ribosomal RNA gene, partial sequence	
	Acinetobacter IwoIIII strain WG-82 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain MR-18-2 16S ribosomal RNA gene, partial sequence	
	Acinetobacter lwoffii strain ML-9 16S ribosomal RNA gene, partial sequence	
	Prolinoborus fasciculus strain MB-24 16S ribosomal RNA gene, partial sequence	
	Profunoborus fasciculus strain MM-111 16S ribosomal RNA gene, partial sequence	
	s profilioporus fasciculus strain MO-20 105 ribosomal KNA gene, partial sequence	

g-proteobacteria | 3 leaves Prolinoborus fasciculus partial 16S rRNA gene, isolate 56 SP635

Bacillus licheniformis DQ071560.1

Bacillus licheniformis strain MKU 1 16S ribosomal RNA gene, partial sequence sequence ID: gb[DQ071560.1] Length: 1430 Number of Matches: 1

Range 1:	437 to 13	88 <u>GenBank</u> <u>Graphi</u>	<u>cs</u>		Next Match 🔺 Previous Ma	tch	
Score 1701 bits	s(1886)	Expect 0.0	Identities 949/952(99%)	Gaps 1/952(0%)	Strand Plus/Minus		
Query	1	AACTCTCGTGG	TGTGACGGGCGGTG	GTACAAGGCCC	GGGAACGTATTCACCG	CGGCATG	60
Sbjct	1388	AACTCTCGTGG	TGTGACGGGCGGTG	GTACAAGGCCC	GGGAACGTATTCACCG	CGGCATG	1329
Query	61	CTGATCCGCGA	TTACTAGCGATTCC	AGCTTCACGCAG	TCGAGTTGCAGACTGC	GATCCGA	120
Sbjct	1328	CTGATCCGCGA	TTACTAGCGATTCCA	AGCTTCACGCAG	TCGAGTTGCAGACTGC	GATCCGA	1269
Query	121	ACTGAGAACAG	ATTTGTGGGATTGG	CTTAGCCTCGCG	GCTTCGCTGCCCTTTG	TTCTGCC	180
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Sbjct	1208	CATTGTAGCAC	GTGTGTAGCCCAGG	CATAAGGGGCA	TGATGATTTGACGTCA	TCCCCAC	1149
Query	241	CTTCCTCCGGT	TTGTCACCGGCAGT	CACCTTAGAGTG	CCCAACTGAATGCTGG	СААСТАА	300
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Query	301	GATCAAGGGTT	GCGCTCGTTGCGGGZ	ACTTAACCCAAC	ATCTCACGACACGAGC	TGACGAC	360
Sbjct	1088	GATCAAGGGTT	GCGCTCGTTGCGGG	ACTTAACCCAAC	ATCTCACGACACGAGC	TGACGAC	1029
Query	361	AACCATGCACC	ACCTGTCACTCTGC	CCCGAAGGGGA	AGCCCTATCTCTAGGG	TTGTCAG	420
Sbjct	1028	AACCATGCACC	ACCTGTCACTCTGC	CCCGAAGGGGA	AGCCCTATCTCTAGGG	TTGTCAG	969
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Sbjct	968	AGGATGTCAAG	ACCTGGTAAGGTTC	TCGCGTTGCTT	CGAATTAAACCACATG	CTCCACC	909
Query	481	GCTTGTGCGGG	CCCCCGTCAATTCC	TTGAGTTTCAG	TCTTGCGACCGTACTC	CCCAGGC	540
Sbjct	908	GCTTGTGCGGG	CCCCCGTCAATTCC	TTGAGTTTCAG	TCTTGCGACCGTACTC	CCCAGGC	849
Query	541	GGAGTGCTTAA	TGCGTTTGCTGCAG	CACTAAAGGGCG	GAAACCCTCTAACACT	TAGCACT	600
Sbjct	848	GGAGTGCTTAA	TGCGTTTGCTGCAG	CACTAAAGGGCG	GAAACCCTCTAACACT	TAGCACT	789
Query	601	CATCGTTTACG	GCGTGGACTACCAG	GTATCTAATCC	TGTTCGCTCCCCACGC	TTTCGCG	660
Sbjct	788	CATCGTTTACG	GCGTGGACTACCAGO	GTATCTAATCC	TGTTCGCTCCCCACGC	TTTCGCG	729
Query	661	CCTCAGCGTCA	GTTACAGACCAGAGA	GTCGCCTTCGC	CACTGGTGTTCCTCCA	CATCTCT	720
Sbjct	728	CCTCAGCGTCA	GTTACAGACCAGAGA	GTCGCCTTCGC	CACTGGTGTTCCTCCA	CATCTCT	669
Query	721	ACGCATTTCAC	CGCTACACGTGGAAT	TCCACTCTCCT	CTTCTGCACTCAAGTT	CCCCAGT	780
Sbjct	668	ACGCATTTCAC	CGCTACACGTGGAAI	TCCACTCTCCT	CTTCTGCACTCAAGTT	CCCCAGT	609
Query	781	TTCCAATGANC	CTCCCCGGTTGAGCO	GGGGGGCTTTCA		CCGCCTG	840
Sbjct	608	TTCCAATGACC	CTCCCCGGTTGAGCO	GGGGGGCTTTCA	САТСАААСТТАААААА	CCGCCTG	549
Query	841	CGCGCGCTTTA	CGCCCAATAATTCCC	GACAACGCTTG	CCACCTACGTATTACC	GCGGCTG	900
Sbjct	548	CGCGCGCTTTA	CGCCCAATAATTCCC	GACAACGCTTG	CCACCTACGTATTACC	GCGGCTG	489
Query	901	CTGGCACGTAA	TTAGCCG-GGCTTTC	TGGTTAGGTAC	CGTCAAGGTACCGCC	951	
Sbjct	488	CTGGCACGTAG	TTAGCCGTGGCTTTC	TGGTTAGGTAC	CGTCAAGGTACCGCC	437	

Icl Query_174773 Bacillus lichenif	ormis strain MKU 1 16S	ribosomal RNA gen	e, partial sequence
- Y			

	Desilies of THOOR LCC - is and DNA and and it is a second
	Bachius sp. 11008 105 hoosomal KIVA gene, partial sequence
	Basillar listenic WK 02 senses
1	Bacillus licheniformis etrain New 2 5 165 schearmal PNA gang matial accuracy
	Bachius inchemitorimis suami rvaniz-5 105 ritosonial RIVA gene, partial sequence
	Primicules bacterium 30B-AC140 105 fibosomial KNA gene, partial sequence
	Bachius inchemiorims strain KKB1 105 thoosonial KINA gene, partial sequence
	Bacilius licheniformis strain V 1M2K0/ 105 fibosomai KNA gene, partial sequence
	Bacillus sonorensis strain ZJY-908 IOS ribosomal KNA gene, partial sequence
	Bacillus sp. BAB-5080 10S ribosomal RNA gene, partial sequence
	Bacillus sp. BAB-5144 10S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain KP050r 16S ribosomal RNA gene, partial sequence
	Bacillus sp. KP057r 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain KP093r 16S ribosomal RNA gene, partial sequence
	Bacillus sp. KP129r 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis partial 16S rRNA gene, strain IRQBAS18
	Bacillus sp. BAB-5495 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain NJ4 16S ribosomal RNA gene, partial sequence
	Bacillus sp. NZ3-1 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain NCIM2715 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain DMB31 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain FI44 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain FI159 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain FI47 16S ribosomal RNA gene, partial sequence
	Bacterium JDSHGTTB03-8 16S ribosomal RNA gene, partial sequence
	Infinitutes 17 leaves
	Bacillus sp. B27(2015) 16S ribosomal RNA gene, partial sequence
	Bacillus sp. B31(2015) 16S ribosomal RNA gene, partial sequence
	Bacillus sp. B-1-40 16S ribosomal KNA gene, partial sequence
	Bacillus sonorensis partial 16S rRNA gene, isolate M33
	Bacillus licheniformis gene for 16S ribosomal RNA, partial sequence, isolate: MS 8516
	Bacillus lichenitornus strain DB2 16S ribosomal RNA gene, partial sequence
	Bacillus sp. ADMK71 16S ribosomal RNA gene, partial sequence
	Bacillus sp. Bac108R 10S ribosomal RNA gene, partial sequence
	Bacillus sp. ADMK1 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis gene for 16S ribosomal RNA, partial sequence, isolate: MS 8519
	Bacillus licheniformis gene for 165 ribosomal KNA, partial sequence, isolate: MS /50/
	Bacilius sp. C-3-11 10S ribosomai KNA gene, partial sequence
	Bacijus lichenitormis strain F1152 105 ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain 1-A-E-54 105 ribosomal RNA gene, partial sequence
	Bacinus inchemitorimis surain ri Port 105 noosomal RNA gene, partial sequence
	Bacillus lichenitormis strain NJ4-1 105 ribosomal KNA gene, partial sequence
	Bactinus licineniformis strain N9 105 ribosomal KNA gene, partial sequence
	For the sequence

firmicutes | 15 leaves firmicutes | 21 leaves Bacillus sp. Bac49R 16S ribosomal RNA gene, partial sequence

Bacillus megaterium KU550043.1

Bacillus megaterium strain EB-238 16S ribosomal RNA gene, partial sequence sequence ID: gb[KU550043.1] Length: 1406 Number of Matches: 1

Range 1:	je 1: 395 to 1361 GenBank Graphics Vext Match 🛦 Previous Match					
Score 1721 bits	s(1908)	Expect 0.0	Identities 962/967(99%)	Gaps 0/967(0%)	Strand Plus/Minus	
Query	1	ACTCTCGTGGT	GTGACGGGCGGTG	TGTACAAGGCCCG	GGAACGTATTCACCGCGGCA	TGC 60
Sbjct	1361	ACTCTCGTGGT	GTGACGGGCGGTG'	TGTACAAGGCCCG	GGAACGTATTCACCGCGGCA	.TGC 1302
Query	61	TGATCCGCGAT	TACTAGCGATTCC	AGCTTCATGTAGG	CGAGTTGCAGCCTACAATCC	GAA 120
Sbjct	1301	TGATCCGCGAT	TACTAGCGATTCC	AGCTTCATGTAGG	CGAGTTGCAGCCTACAATCC	GAA 1242
Query	121	CTGAGAATGGT	TTTATGGGATTGG	CTTGACCTCGCGG	TCTTGCAGCCCTTTGTACCA	TCC 180
Sbjct	1241	CTGAGAATGGT	TTTATGGGATTGG	CTTGACCTCGCGG	TCTTGCAGCCCTTTGTACCA	TCC 1182
Query	181	ATTGTAGCACG	TGTGTAGCCCAGG'	TCATAAGGGGCAT	GATGATTTGACGTCATCCCC	ACC 240
Sbjct	1181	ATTGTAGCACG	TGTGTAGCCCAGG	TCATAAGGGGCAT	GATGATTTGACGTCATCCCC	ACC 1122
Query	241	TTCCTCCGGTT	TGTCACCGGCAGT	CACCTTAGAGTGC	CCAACTAAATGCTGGCAACT	AAG 300
Sbjct	1121	TTCCTCCGGTT	TGTCACCGGCAGT	CACCTTAGAGTGC	CCAACTAAATGCTGGCAACT	AAG 1062
Query	301	ATCAAGGGTTG	CGCTCGTTGCGGG	ACTTAACCCAACA	TCTCACGACACGAGCTGACG	ACA 360
Sbjct	1061	ATCAAGGGTTG	CGCTCGTTGCGGG	ACTTAACCCAACA	TCTCACGACACGAGCTGACG	ACA 1002
Query	361	ACCATGCACCA	CCTGTCACTCTGT	CCCCCGAAGGGGA	ACGCTCTATCTCTAGAGTTG	TCA 420
Sbjct	1001	ACCATGCACCA	CCTGTCACTCTGT	CCCCCGAAGGGGA	ACGCTCTATCTCTAGAGTTG	TCA 942
Query	421	GAGGATGTCAA	GACCTGGTAAGGT'	ICTTCGCGTTGCT	TCGAATTAAACCACATGCTC	CAC 480
Sbjct	941	GAGGATGTCAA	GACCTGGTAAGGT	TCTTCGCGTTGCT	TCGAATTAAACCACATGCTC	CAC 882
Query	481	CGCTTGTGCGG	GCCCCCGTCAATT	CCTTTGAGTTTCA	AGTCTTGCGACCGTACTCCCC	AGG 540
Sbjct	881	CGCTTGTGCGG	GCCCCCGTCAATT	CCTTTGAGTTTCA	AGTCTTGCGACCGTACTCCCC	AGG 822
Query	541	CGGAGTGCTTA		AGCACTAAAGGGC		CAC 600
SDJCT	821	CGGAGTGCTTA	ATGCGTTAGCTGC.	AGCACTAAAGGGC	GGAAACCCTCTAACACTTAG	CAC /62
Query	601	TCATCGTTTAC	GGCGTGGACTACC	AGGGTATCTAATC	CTGTTTGCTCCCCACGCTTT	CGC 660
Sbjet	761	TCATCGTTTAC	GGCGTGGACTACC	AGGGTATCTAATC	CTGTTTGCTCCCCACGCTTT	CGC 702
Query	661	GCCTCAGCGTC	AGTTACAGACCAA	AAAGCCGCCTTCG	CCACTGGTGTTCCTCCACAT	CTC 720
Sbjct	701	GCCTCAGCGTC	AGTTACAGACCAA	AAAGCCGCCTTCG	GCCACTGGTGTTCCTCCACAT	CTC 642
Query	721					CAG 780
Sbjet	041	TACGCATTTCA	CCGCTACACGTGG			CAG 582
Query	781					
Sujet	041	CCCCCCCCCC	ACCCCCALGGITGA	GCCGTGGGCTTTC		CCT 522
Query	641 501					
Oueru	901	TCCTCCCCCTT	AUGUUUAATAATT			1000 402
Spict	461	TGCTGGCACGT				 100
Ouery	961	TCTTCTT	7	TITCIGGIIMGGI	LOODI CANDI LOODI CANDI LOODI	IAC 402
Sbjct	401	IIIIIII TCTTGTA 39	, 5			
2	-					

1	QIcl/Query 200683
p	Bacillus flexus strain ysm11 16S ribosomal RNA gene, partial sequence
	Bacillus sp. YH4 16S ribosomal RNA gene, partial sequence
	Dacings 50, 117 105 noosomai KNA gene, partial sequence
	Printing 2 leaves
	firminites 2 large
	Bacillus arvabhattai strain KRM 11 16S ribosomal RNA gene nartial sequence
	Bacillus sp. NI(2014) 16S ribosomal RNA gene, partial sequence
	Bacillus megaterium strain KNUSC3019 16S ribosomal RNA gene, partial sequence
	Bacillus megaterium strain EB-238 16S ribosomal RNA gene, partial sequence
	Bacillus aryabhattai strain KNUSC3049 16S ribosomal KNA gene, partial sequence
	Dacillus aryaonattai strain 110 105 moosomal KNA gene, partial sequence
	Bacillus areals su am 10100-2 105 notsomai reive gene, partial sequence
	Bacillus sp. 35(2015) 16S nbosomal RNA gene, partial sequence
	Bacillus megaterium gene for 16S ribosomal RNA, partial sequence, strain: BEb-50
	Bacillus flexus strain TWSL_13 16S ribosomal RNA gene, partial sequence
	Bacilius sp. BAB-3130 165 mbosomal KNA gene, partial sequence
	Azotobacter chroococcum partial 105 rKNA gene, isolate Azoto GaK
	Bacillus flexus strain CO31 103 flotoomal RVA gene, partial sequence
	Bacillus mezaterium strain BGAS 39E 16S ribosomal RNA gene, partial sequence
	Bacillus flexus strain NB4-9 16S nbosomal RNA gene, partial sequence
	Bacillus sp. 40 pH8 16S ribosomal RNA gene, partial sequence
	Bacillus megaterium strain M9-1B 16S ribosomal RNA gene, complete sequence
	Bacillus megatenum strain L24 165 nbosomai KNA gene, partial sequence
	Bacillus megaterium suain 1.50 ros rioosomai RNA gene, partai sequence
	Bacillus megaterium strain IARI-SWC 20 16S nbosomal RNA zene, partial sequence
	Bacterium XF013 16S ribosomal RNA gene, partial sequence
	Bacillus flexus strain NB3-6 16S ribosomal RNA gene, partial sequence
	Bacillus megaterium strain NM36 16S ribosomal KNA gene, partial sequence
	Dacinus sp. PKSULU 105 noosomai KNA gene, partial sequence
	Bacillus en VSSVII0 16S mbosonial RNA gene narrial sequence
	Bacillus sp. BAB-5149 16S ribosomal RNA gene, partial sequence
	Bacillus megaterium strain BLZ01 16S ribosomal RNA gene, partial sequence
	Bacillus megaterium gene for 16S nbosomal RNA, partial sequence, strain: BEb-46
	V Bacillus sp. W3-1 105 mosomai KNA gene, partal sequence
	ulacinus megaterium suam 170-0 105 noosomal AVA gene, partial sequence
	Unentimed Bavillus on game for 165 mbosomal RNA martial segmence clone: 4X1
	Bacillus arvabhattai strain PB5 165 ribosomal RNA gene, partial sequence
	^O Bacterium C0989 16S ribosomal RNA gene, partial sequence
	firmicutes 18 leaves
	Maniferences 15 Jeaves
	Bacillus en enrichment culture clone CDB/011 16S ribosomal RNA gene nartial sequence
	Bacillus megaterium strain B9-9B 16S ribosomal RNA gene, complete sequence
	Bacillus licheniformis strain NZ2-3 16S ribosomal RNA gene, partial sequence
0.000	Geobacillus stearothermophilus strain NB4-3 16S ribosomal KNA gene, partial sequence
9.0002	Bacilius tiexus strain SBANSUa10 16S ribosomal KNA gene, partial sequence
	Baching Sp. 17A 1-27/01 105 motosomai ruve gene, partial sequence
	firmientas 10 lasvas
	dimension of the second s

Staphylococcus succiuns KJ888125.1

Staphylococcus succinus strain MI-6 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KJ888125.1</u>| Length: 1381 Number of Matches: 1

Range 1:	40 to 98	88 <u>GenBank</u>	Graphics				V Next M	1atch 🔺 Pre	vious Match	
Score 1660 bit	s(1840))	Expect 0.0	Identities 940/949(99%)	Gaps 3/949(0%	6)	Strand Plus/Plus		
Query	1	AACACO	TGGGTAA	CCTACCTATA	AGACTG	GAATAAC	TTCGGG	AAACCGG	AGCTAATGCCG	60
Sbjct	40	AACACO	TGGGTAA	CCTACCTATA	AGACTG	GAATAAC	IIIII TTCGGG	AAACCGG	AGCTAATGCCG	99
Query	61	GATAAC	ATATAGA	ACCGCATGGT	TCTATA	TGAAAG	ATGGTT	TTGCTAT	CACTTATAGAT	120
Sbjct	100	GATAAC	CATATAGA	ACCGCATGGT	TCTATA	GTGAAAG	ATGGTT'	TTGCTAT	CACTTATAGAT	159
Query	121	GGACCO	GCGCCGT	ATTAGCTAGT	TGGTAA	GTAATG	GCTTAC		ACGATACGTAG	180
Sbjct	160	GGACCO	GCGCCGT	ATTAGCTAGT	TGGTAA	GTAATG	GCTTAC	CAAGGCG	ACGATACGTAG	219
Query	181	CCGACO	TGAGAGG	GTGATCGGCC	ACACTG	GAACTGA	GACACG	GTCCAGA	CTCCTACGGGA	240
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Sbjct	280	GGCAGO	AGTAGGG	AATCTTCCGC	AATGGG	GAAAGC	CTGACG	GAGCAAC	GCCGCGTGAGT	339
Query	301	GATGA		GGATCGTAAA	ACTCTG	TATTAG	GGAAGA	ACAAATG	CGTAAGTAACT	360
Sbjct	340	GATGA	AGGTTTTC	GGATCGTAAA	ACTCTG	TATTAG	GGAAGA	ACAAATG	CGTAAGTAACT	399
Query	361	GTGCGC	ATCTTGA	CGGTACCTAA	TCAGAA	AGCCACG	GCTAAC	TACGTGC	CAGCAGCCGCG	420
Sbjct	400	GTGCGC	ATCTTGA	CGGTACCTAA	TCAGAA	AGCCACG	GCTAAC	TACGTGC	CAGCAGCCGCG	459
Query	421	GTAATA	ACGTAGGT	GGCAAGCGTT	ATCCGG	ATTATT	GGGCGT	AAAGCGC	GCGTAGGCGGT	480
Sbjct	460	GTAATA	ACGTAGGT	GGCAAGCGTT	ATCCGG	ATTATT	GGGCGT	AAAGCGC	GCGTAGGCGGT	519
Query	481	TTCTT	AGTCTGA	TGTGAAAGCC	CACGGC	CAACCG	IGGAGG	GTCATTG	GAAACTGGGAA	540
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Query	541	ACTTGA	AGTGCAGA	AGAGGAAAGT	GGAATTO	CATGTG		TGAAATG	CGCAGAGATAT	600
Sbjct	580	ACTTGA	AGTGCAGA	AGAGGAAAGT	GGAATTO	CATGTG	TAGCGG	TGAAATG	CGCAGAGATAT	639
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Sbjct	640	GGAGGA	ACACCAG	TGGCGAAGGC	GACTTTC	TGGTCT	GTAACT	GACGCTG	ATGTGCGAAAG	699
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Sbjct	700	CGTGGG	GATCAAA	CAGGATTAGA	TACCCTO	GTAGTC	CACGCC	GTAAACGA	ATGAGTGCTAA	759
Query	721	GTGTTA	GGGGGTT	TCCGCCCCTT	AGTGCT	CAGCTA	ACGCAT	FAAGCAC	ICCGCCTGGGG	780
Sbjct	760	GTGTTA	GGGGGTT	TCCGCCCCTT	AGTGCT	CAGCTA	ACGCAT	FAAGCAC	ICCGCCTGGGG	819
Query	781	AGTACO	ACCGCAA	GGTTGAAACT	CAAAGGA	ATTGAC	GGGGAC	CCGCACAZ	AGCGGTGGAGC	840
Sbjct	820	AGTACG	ACCGCAA	GGTTGAAACT	CAAAGGA	ATTGAC	GGGGAC	CCGCACA	AGCGGTGGAGC	879
Query	841	ATGTGG	TTTAATT	CGAAGCAACG	CGAAAAA	CCTTAC	C-AATC	TTGACAT	CCTTTG-AAAC	898
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Query	899	TCTAAA	GATAAAG	CCTTCCCCTT	CGGGGGG		GACAgg	gggggCA	946	
Sbjct	940	TCTAGA	GATAGAG	CCTTCCCCTT	CGGGGGG	CAAAGT	GACAGG	IGGTGCA	988	

	Staphylococcus succinus strain IARI-SL-31 16S ribosomal RNA gene, partial sequence	
	 Uncumured bacterium cione C1 13 ToS infosomat RNA gene, pa Saphylococcus succinus strain EN4 16S ribosomal RNA gene, partial sequence 	artai sequence
	 Staphylococcus succinus subsp. succinus partial 165 rRNA gene, isolate 0911MAR22V8 	Icl/Query_172539
	 Staphylococcus stacunus strain AMG-DI 16S ribosomal RNA gene, complete sequence Uncultured Staphylococcus sp. clone Gexpansi.Q1 16S ribosomal RNA gene, partial sequence 	
	Staphylococcus sp. NP24 16S ribosomal RNA gene, partial sequence Staphylococcus succinus strain EM17 16S ribosomal RNA gene, partial sequence	
	Staphylococcus succinus strain T16 16S ribosomal RNA gene, partial sequence bacteria 12 Jeaves	
	Uncultured bacterium clone nbw766g04c1 16S ribosomal RNA gene, partial sequence	
	Staphylococcus succinus strain EK2 16S ribosomal RNA gene, partial sequence	
	Staphylococcus succinus strain \$2 16S ribosomal RNA gene, partial sequence	
	Saphylococcus sylosus strain 118 165 nbosomal KNA gene, partial sequence Uncultured bacterium clone nbw1178/05c1 165 nbosomal RNA gene, partial sequence	
	Firmicutes [3 leaves Staphylococcus succinus strain IHB B 6872 16S ribosomal RNA gene, partial sequence	
	 Uncultured bacterium clone ncd980c06c1 16S ribosonial RNA gene, partial sequence Staebylococcus succinus strain 57A (C3P2) 16S ribosonial RNA gene, partial sequence 	
	Uncultured bacterium clone nbw316a/5c1 16S ribosomal RNA gene, partial sequence Sanhvlococcus succinus subso succinus strain M1-10 16S ribosomal RNA eene, partial sequence	
	Uncultured bacterium clone nbw776g02c1 16S nbosomal RNA gene, partial sequence Granicutes [2] bases	
	Staphylococcus sp. MT5 16S ribosomal RNA gene, partial sequence	
	Staphylococcus succinus strain 99A (BR46) 16S ribosonal RNA gene, partial sequence	
	Stenotrophomonas sp. L5 16S ribosomal RNA gene, partial sequence	
	Staphylococcus succinus strain GCA867 16S ribosomal RNA gene, partial sequence	
	Shapriykooccus sp. M.208-2 165 ribosomal RNA gene, partial sequence Uncultured bacterium clone nbw228d02c1 165 ribosomal RNA gene, partial sequence	
	Uncultured Furniscutes bacterium clone b5 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone nbw315g07c1 16S ribosomal RNA gene, partial sequence	
	Uncultured Staphylococcus sp. clone bottae 10 16S ribosomal RNA gene, partial sequence Staphylococcus sp. TM6 3 16S ribosomal RNA gene, partial sequence	
0000	Staphylococcus succinus strain M40 16S ribosonal RNA cene, partial sequence	ultured bacterium clone nbw250g10c1 16S ribosomal RNA gene, partial sequence
.0005	Uncultured bacterium clone nbw777/d8c116S ribosomal RNA gene, partial sequence Sanhvlococcus succinus strain 1 (C3S1) 16S ribosomal RNA eene, partial sequence	
Growieutes 12 leaves	e nbw316a06c1 165 ribosomal RNA gene, partial sequence	
Uncultured bacterium clone D128 ERTV 4 16S ribo	somal RNA gene, partial sequence train EGN 2011 1.65 mbacamel RNA areas portial companyo	
Staphylococcus succinus strain MI-6 16S ribosomal R	NA gene partial sequence	
Staphylococcus succinus NIPHL090904/K5 16S ribo	over perce, parata sequence Unarband bestering along above 700/011 16C above no 180 k anna motial annument	
1	Staphyleoceus succinis strain JDM2 2A 165 ribosonal RNA gene, partial sequence	
Saphylococcus sp. 614B2	12EGalle 165 ribosomal RNA gene, partial sequence	
Staphylococcus sp. Be-4 1	as movemu RAA gene, partial sequence 65 movemu RAA gene, partial sequence	
Staphylococcus succinus s	ando, ascentas sertam FA-1 too mosemaa RAA gene, partial sequence train ASW2C 16S rhosemal RAA gene, partial sequence	
Uncultured bacterium clon	zan roke-1-2- tos mosoma KSA gene, partal sequence e ned950g08c1 165 ribosomal RNA gene, partal sequence	
bacteria 4 leaves		B14
	Suppyoeccus succuts strain SI-I5-1 I6S ribosomal RNA, partial sequence, Bacterium Y	KNA gene, partai sequence
	* lamacules [4 kaves	Staphylococcus equorum subsp. equorum partial 16S rRNA gene, isolate 0312MAR
Bees	v ♦ bacteria 17 kaves	Staphylococcus gallinarum strain JDM2_3A 16S ribosomal RNA gene, partial sequen
firmicites 2 leaves		

Bacillus pumilus KU230023.1

Bacillus pumilus strain m350 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb[KU230023.1]</u> Length: 1396 Number of Matches: 1

Range 1:	nge 1: 399 to 1340 GenBank Graphics Vext Match 🔺 Previous Match					
Score 1663 bits	s(1844)	Expect 0.0	Identities 934/942(99%)	Gaps 0/942(0%)	Strand Plus/Minus	
Query	1	TGTGACGGGCG	GTGTGTACAAGGCCC	GGGAACGTATTC	ACCGCGGCATGCTGATCCGCGA	60
Sbjct	1340	 TGTGACGGGCG	GTGTGTACAAGGCCC	GGGAACGTATTC		1281
Query	61	TTACTAGCGAT	TCCAGCTTCACGCAG	TCGAGTTGCAGA	ACTGCGATCCGAACTGAGAACAG	120
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Query	121	ATTTATGGGAT	TGGCTAAACCTTGCG	GTCTTGCAGCCC	TTTGTTCTGTCCATTGTAGCAC	180
Sbjct	1220	ATTTATGGGAT	TGGCTAAACCTTGCG	GTCTTGCAGCCC	TTTGTTCTGTCCATTGTAGCAC	1161
Query	181	GTGTGTAGCCC	AGGTCATAAGGGGCA	TGATGATTTGAC	GTCATCCCCACCTTCCTCCGGT	240
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Query	241	TTGTCACCGGC.	AGTCACCTTAGAGTG	CCCAACTGAATG	CTGGCAACTAAGATCAAGGGTT	300
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Query	301	GCGCTCGTTGC	GGGACTTAACCCAAC	ATCTCACGACAC	GAGCTGACGACAACCATGCACC	360
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Query	361	ACCTGTCACTC	TGTCCCCGAAGGGAA	AGCCCTATCTCI	AGGGTTGTCAGAGGATGTCAAG	420
Sbjct	980	ACCTGTCACTC	TGTCCCCGAAGGGAA	AGCCCTATCTCI	CAGGGTTGTCAGAGGATGTCAAG	921
Query	421	ACCTGGTAAGG	TTCTTCGCGTTGCTT	CGAATTAAACCA	ACATGCTCCACCGCTTGTGCGGG	480
Sbjct	920	ACCTGGTAAGG	TTCTTCGCGTTGCTT	CGAATTAAACCA	ACATGCTCCACCGCTTGTGCGGG	861
Query	481	CCCCCGTCAAT	TCCTTTGAGTTTCAG	ICTTGCGACCGI	ACTCCCCAGGCGGAGTGCTTAA	540
Sbjct	860	CCCCCGTCAAT	TCCTTTGAGTTTCAG	ICTTGCGACCGI	CACTCCCCAGGCGGAGTGCTTAA	801
Query	541	TGCGTTAGCTG	CAGCACTAAGGGGCG	GAAACCCCCTAA	CACTTAGCACTCATCGTTTACG	600
Sbjct	800	TGCGTTAGCTG	CAGCACTAAGGGGCG	GAAACCCCCTAA	CACTTAGCACTCATCGTTTACG	741
Query	601	GCGTGGACTAC	CAGGGTATCTAATCC	TGTTCGCTCCCC	CACGCTTTCGCTCCTCAGCGTCA	660
Sbjct	740	GCGTGGACTAC	CAGGGTATCTAATCC	IGTTCGCTCCCC	CACGCTTTCGCTCCTCAGCGTCA	681
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Query	721	CGCTACACGTG	GAATTCCACTCTCCT	CTTCTGCACTCA	AGTTTCCCAGTTTCCAATGACC	780
Sbjct	620	CGCTACACGTG	GAATTCCACTCTCCT	CTTCTGCACTCA	AGTTTCCCAGTTTCCAATGACC	561
Query	781	CTCCCCGGTTG.	AGCCGGGGGGCTTTCA	CATCAAACTTAA	AAAACCGCCTGCGAGCCCTTTA	840
Sbjct	560	CTCCCCGGTTG	AGCCGGGGGGCTTTCA	CATCAGACTTAA	AGAAACCGCCTGCGAGCCCTTTA	501
Query	841	CGCCCAATAAT	TCCGGACAACGCTTG	CCACCTACGTAI	TACCGCGGCTGCTGGCACGTAA	900
Sbjct	500	CGCCCAATAAT	TCCGGACAACGCTTG	CCACCTACGTAT	TACCGCGGCTGCTGGCACGTAG	441
Query	901	TTAGCCGGGGC	TTTCTGGGTTAGTAC	CGTCAAGGGGCG	GAGCA 942	
Sbjct	440	TTAGCCGTGGC	TTTCTGGTTAGGTAC	CGTCAAGGTGCG	GAGCA 399	

	Bacillus sp. WMR1 165 nbosomal RNA gene, partial sequence	
	 Bacilias sp. N17(2014) 16S ribosomal RNA gene, partial sequence 	Bacilia in SW-1216S elocional PVA anna a
		Bacillus sp. G121 16S ribosomal RNA eene, nartial sequence
		Bacillus sp. M403(2014) 16S ribosomal RNA gene, partial sequence
		Bacillus sp. M408 16S ribosomal RNA gene, partial sequence
		Bacilius pumilus strain ASU2 165 nbosomal KNA gene, partial sequence Bacilius safoncis etnin C11 165 nbosomal RNA gene, partial sequence
		Bacillus safensis strain GA-5 16S ribosonal RNA gene, partial sequence
		Bacillus sp. CT 7-03 16S ribosomal RNA gene, partial sequence
		Bacillus safensis strain KP1 16S ribosomal RNA gene, partial sequence
		Bacillus safensis stram YR1 16S ribosomal RNA gene, partial sequence
		 Dachus Salensis Stain D31 105 mbosonal KNA gene, partial sequence Ravillar numiliar strain E.1.1.1165 riboronal DSA anna matial assumes
		 Bacillus so, RKAT-0008 partial 165 rRNA cene, isolate RKAT-0008
		Bacillus sp. BAB-5103 16S ribosomal RNA gene, partial sequence
		 Bacillus sp. BAB-5275 16S ribosomal RNA gene, partial sequence
		Bacillus satensis strain 14 165 ribosomal RNA gene, partial sequence
		 Bacillus safensis drain IHBB 9177-16S ribosomal RNA cene, nartial sequence
		Bacillus rumilus strain ASR-35 165 ribosomal RNA eene, partial sequence
		Bacillus pumilus strain C1C3702 16S ribosomal RNA gene, partial sequence
		Bacillus pumilus strain SBANHCa15 16S ribosomal RNA gene, partial sequence
		Bacillus cercus strain UB14 105 ribosonial KNA gene, partial sequence
		 Bacillus rumilus strain UK35 105 ribosonial KNA gene, partial sequence Bacillus rumilus strain BS131 16S ribosonial RNA cone, nartial sequence
		Bacillus safensis strain L26 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone 958-7 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone 335-2 16S ribosomal RNA gene, partial sequence
		Recillus numilus strain RV21 165 ribosomal RNA gene, partial sequence
		Bacillus safensis strain JCR-44 16S ribosomal RNA cene, nartial sequence
		Bacillus altitudinis partial 16S rRNA gene, isolate HLIRP1
		Bacillus sp. ADMK36 16S nbosomal RNA gene, partial sequence
		Bacillus safensis strain MCCB 308 165 ribosomal KNA gene, partial sequence
		Redhis an RAR 5572 16S ribusonal RNA anno matial sequence
		 Bacillus altitudinis partial 16S rRNA gene, isolate LK4HaP15
		Bacillus pumilus strain KMDH10 16S ribosomal RNA gene, partial sequence
102		Bacillus pumilus strain ST15 16S ribosomal RNA gene, partial sequence
003		 Baculus antiaanus sciam AA0-1 105 filosomai KNA gene, partial sequence Incultured bacterium clone 335,5 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone 958-18 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone 958-4 16S ribosomal RNA gene, partial sequence
		Bacilius sp. NZ2-5 16S ribosomal RNA gene, partial sequence
		 Bacillus sp. CRI14 105 fibosonial RNA gene, partial sequence Bacillus rumilus strain CL21 16S ribosonial RNA eene, nartial sequence
		Bacillus pumilus strain SBANHO9 16S ribosomal RNA gene, partial sequence
		Bacillus safensis strain C1AC3701 16S ribosomal RNA gene, partial sequence
		Bacterium ARb28 16S ribosomal RNA gene, partial sequence
		Azogririllum linoferum partial 165 rRNA gene, justala sequence
		firmicutes 15 leaves
		Bacillus pumilus strain mv49b 16S ribosomal RNA gene, partial sequence
		Bacillus sp. ADMK74 16S ribosomal RNA gene, partial sequence Bacillus on ADMK72 16S ribosomal DNA gene, partial sequence
		Racillus safensis grain JCR-65 16S ribosomal RNA cene, partial sequence
		Bacillus safensis strain DB3 16S ribosomal RNA gene, partial sequence
		Bacillus pumilus strain AA5-5 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone 335-18 16S ribosomal RNA gene, partial sequence
		Orcultured bacterium clone 958-5 16S ribosomal RNA gene, partial sequence
		Bacillus pumilus strain WD204 16S ribosomal RNA gene, partial sequence
		Bacillus pumilus strain RAS2-5 16S ribosomal RNA gene, partial sequence
		Bacillus sp. CL15 165 nbosomal RNA gene, partial sequence
		Recillus safencis drain CM3701 16S ribosonal RNA cone, nartial sequence
		Bacillus safensis strain OU93 16S ribosomal RNA gene, partial sequence
		Bacillus safensis strain IHBB 11005 16S ribosomal RNA gene, partial sequence
		Bacillus pumilus strain SBB12 16S ribosomal RNA gene, partial sequence
		 Bacillus y MDL/14 I6S ribosomal RNA gene, partial sequence Bucillus y MDL/14 I6S ribosomal RNA gene partial sequence
		Bacillus pumilus strain HN-10 16S ribosonal RNA cene, partial sequence
		Bacillus sp. BAB-5134 16S ribosonal RNA gene, partial sequence
		Bacillus sp. RKAT-16 partial 16S rRNA gene, isolate RKAT-16
		Endoptivite bacterium SV/15/16S nitosomal RNA gene, partial sequence
_		THINK URS 19 KAVES

Bacillus licheniformis KT200463.1

Bacillus licheniformis strain KP062r 16S ribosomal RNA gene, partial sequence sequence ID: $\underline{gb[KT200463.1]}$ Length: 1201 Number of Matches: 1

Range 1:	Range 1: 68 to 1014 GenBank Graphics Vext Match A Previous Match						
Score	-(100.4)	Expect	Identities	Gaps	Strand		
1700 bit	s(1884)	0.0	945/947(99%)	0/947(0%)	Plus/Plus		
Query	1	CGGTGTGTACAA	GGCCCGGGAACGT	ATTCACCGCGGCAT(GCTGATCCGCGAT'	TACTAGCG	60
Sbjct	68	CGGTGTGTACAA	GGCCCGGGAACGT	ATTCACCGCGGCAT	GCTGATCCGCGAT'	TACTAGCG	127
Query	61	ATTCCAGCTTCA	.CGCAGTCGAGTTG(CAGACTGCGATCCGA	AACTGAGAACAGA'	TTTGTGGG	120
Sbjct	128	ATTCCAGCTTCA	CGCAGTCGAGTTG	CAGACTGCGATCCG	AACTGAGAACAGA	TTTGTGGG	187
Query	121	ATTGGCTTAGCC	TCGCGGCTTCGCT	GCCCTTTGTTCTGC	CCATTGTAGCACG	TGTGTAGC	180
Sbjct	188	ATTGGCTTAGCC	TCGCGGCTTCGCT	GCCCTTTGTTCTGC	CCATTGTAGCACG	TGTGTAGC	247
Query	181	CCAGGTCATAAG	GGGCATGATGATT	IGACGTCATCCCCA	CCTTCCTCCGGTT	TGTCACCG	240
Sbjct	248	CCAGGTCATAAG	GGGCATGATGATT	IGACGTCATCCCCA	CCTTCCTCCGGTT	IGTCACCG	307
Query	241	GCAGTCACCTTA	GAGTGCCCAACTG	AATGCTGGCAACTA	AGATCAAGGGTTG	CGCTCGTT	300
Sbjct	308	GCAGTCACCTTA	GAGTGCCCAACTG	AATGCTGGCAACTA	AGATCAAGGGTTG	CGCTCGTT	367
Query	301	GCGGGACTTAAC	CCAACATCTCACG			CCTGTCAC	360
Sbjct	368	GCGGGACTTAAC	CCAACATCTCACG	ACACGAGCTGACGA	CAACCATGCACCA	CCTGTCAC	427
Query	361	TCTGCCCCCGAA	GGGGAAGCCCTAT		GAGGATGTCAAGA	CCTGGTAA	420
Sbjct	428	TCTGCCCCCGAA	GGGGAAGCCCTAT	CTCTAGGGTTGTCA	GAGGATGTCAAGA	CCTGGTAA	487
Query	421	GGTTCTTCGCGT	TGCTTCGAATTAA	ACCACATGCTCCAC	CGCTTGTGCGGGC	CCCCGTCA	480
Sbjct	488	GGTTCTTCGCGT	TGCTTCGAATTAA	ACCACATGCTCCAC	CGCTTGTGCGGGC	CCCCGTCA	547
Query	481	ATTCCTTTGAGT	TTCAGTCTTGCGA		CGGAGTGCTTAAT	GCGTTTGC	540
Sbjct	548	ATTCCTTTGAGT	TTCAGTCTTGCGA	CCGTACTCCCCAGG	CGGAGTGCTTAAT	GCGTTTGC	607
Query	541	TGCAGCACTAAA	GGGCGGAAACCCT	CTAACACTTAGCAC		CGTGGACT	<mark>600</mark>
Sbjct	608	TGCAGCACTAAA	GGGCGGAAACCCT	CTAACACTTAGCAC	ICATCGTTTACGG	CGTGGACT	667
Query	601	ACCAGGGTATCT.	AATCCTGTTCGCT	CCCACGCTTTCGCC	CCTCAGCGTCAG	TTACAGAC	660
Sbjct	668	ACCAGGGTATCT.	AATCCTGTTCGCT	CCCCACGCTTTCGCC	SCCTCAGCGTCAG	TTACAGAC	727
Query	661	CAGAGAGTCGCC	TTCGCCACTGGTG	TCCTCCACATCTC	TACGCATTTCACC	GCTACACG	720
Sbjct	728	CAGAGAGTCGCC	TTCGCCACTGGTG	TTCCTCCACATCTC	TACGCATTTCACC	GCTACACG	787
Query	721	TGGAATTCCACT	CTCCTCTTCTGCAG	CTCAAGTTCCCCAG	TTTCCAATGACCC	ICCCCGGT	780
Sbjct	788	TGGAATTCCACT	CTCCTCTTCTGCAC	CTCAAGTTCCCCAG	TTTCCAATGACCC	ICCCCGGT	847
Query	781	TGAGCCGGGGGC	TTTCACATCAAACT	TAAGAAACCGCCT	GCGCGCGCTTTAC	GCCCAATA	840
Sbjct	848	TGAGCCGGGGGC	TTTCACATCAGACI	TAAGAAACCGCCT	GCGCGCGCTTTAC	GCCCAATA	907
Query	841	ATTCCGGACAAC	GCTTGCCACCTAC	GTATTACCGCGGCT(GCTGGCACGTAAT	TAGCCGTG	900
Sbjct	908	ATTCCGGACAAC	GCTTGCCACCTAC	GTATTACCGCGGCT	GCTGGCACGTAAT	FAGCCGTG	967
Query	901	GCTTTCTGGTTA	GGTACCGTCAAGG		AACGGGAC 947		
Sbjct	968	GCTTTCTGGTTA	GGTACCGTCAAGGI	FACCGCCCTATTCG	AACGGGAC 101	4	

LellQuery_195375	
Bacilius lichenitormi	s stram KP062r 10S nbosomal KNA gene, partial sequence Bacillus licheniformis strain FO-074b 16S ribosomal RNA zene, natial sequence
	GBacillus licheniformis strain 30N1-10 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain Nam2-5 165 ribosomal RNA gene, partial sequence
	Bacillus inchemitormis strain GFL2 105 mosconal RNA4 gene, partial sequence
	Bacillus sonorenis strain ZY-908 165 riboromal RNA eree, partial sequence
	Bacillus sp. RKNM-0105 partial 16S rRNA gene, isolate RKNM-0105
	Bacillus sp. BAB-5095 16S ribosomal RNA gene, partial sequence
	Bacinis inchemitormis strain Q05 105 Hoosomal KVA gene, partai sequence
	Bacillus licheniformis stain RCOStr 16S ribosomal RNA spene, partial sequence
	Bacillus licheniformis strain KP081r 16S ribosomal RNA gene, partial sequence
	Bacillus sp. KP097r 16S ribosomal RNA gene, partial sequence
	Bacilius 59, KP1297 ToS noosomai RNA gene, partial sequence
	Bacillus inchemiormis w x-02 genome Bacillus inchemiormis antial 165 rRNA gene strain IROBAS19
	Bacillus sp. BAB-5495 16S ribosomal RNA gene, partial sequence
	Bacterium AKb15 16S ribosomal KNA gene, partial sequence
	Bacillus lichemitorinis strain CL-3-5 IOS indosomai RNA gene, partial sequence
	Bachilla Si Icheniformis strain NCIN42471 ISS ribosomat RNA gene, partial sequence
	Bacillus licheniformis strain HTP01 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain JS-1 16S nbosomal RNA gene, partial sequence
	Bacillus lichenitormis strain FI44 105 mbosomai KNA gene, partial sequence
	Placific in Figure 1 and the second s
	Bacillus sp. C-3-11 16S ribosomal RNA gene, partial sequence
	Bacillus inchemiformis gene for 16S ribosomal RNA, partial sequence, isolate: MS 5705
	Bacillus inchemitormis gene for 105 riposomal KVA, partai sequence, isolate: MS 8519
	Bacillus sp. ADMK1 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain HRBL-15TD17, complete genome
	Bacillus sp. Bacios his ribosomal KNA gene, partial sequence
	Bacilla se Los Anna RNA sene narial sequence
	Bacillus licheniformis gene for 16S ribosomal RNA, partial sequence, isolate: MS 8516
	Bacillus licheniformis gene for 165 ribosomal RNA, partial sequence, isolate: MS 5716
	Bacillus sonorensis partial 105 rAVA gene, ISO218 M35 Bacillus en BL-208 165 Indecom IRM gene partial semance
	Bacillus sp. Dr 27/2015) 16S ribosomal RNA gene, partial sequence
	Bacillus licheniformis strain F147 16S ribosomal RNA gene, partial sequence
10.0003	Bacillus licheniformis strain F142 165 ribosomal RNA gene, partial sequence
0.0003	Charlen in JUSA (vi 1 DUS-8 105 noosonal RAVA gene, partal sequence
1 1	firmicutes [3] leaves
	Bacillus sp. Bac49R 16S ribosomal RNA gene, partial sequence

9- The phylogenetic analysis of bacteria isolated from upper surfaces of water taps samples

Staphylococcus epidermidis KT887972.1

Staphylococcus epidermidis strain LIPOA/UEL_42 16S ribosomal RNA gene, partial sequence Sequence ID: <a href="https://doi.org/10.1016/journal.partial-sequence-id-adda-sequence-

Range 1: 3 to 1051 GenBank Graphics Vext Match 🔺 Previous Match							
Score 1810 bits	s(2006)	Expect 0.0	Identities 1033/1049(98%)	Gaps 3/1049(0%)	Strand Plus/Plus	
Query	1	GCTCCTCTGAC	GTTAGCGGCGGACG	GGTGAGTAA	CACGTGGA	TAACCTACCTATAAGACT	60
Sbjct	3	GCTCCTCTGAC	GTTAGCGGCGGACG	GGTGAGTAA	CACGTGGA	TAACCTACCTATAAGACT	62
Query	61	GGGATAACTTC	GGGAAACCGGAGCI	AATACCGGA	TATATAT	TGAACCGCATGGTTCAAT	120
Sbjct	63	GGGATAACTTC	GGGAAACCGGAGCI	'AATACCGGA'	PATATAT	IGAACCGCATGGTTCAAT	122
Query	121	AGTGAAAGACO	GTTTTGCTGTCACI	TATAGATGG	ATCCGCGC	CGCATTAGCTAGTTGGTA	180
Sbjct	123	AGTGAAAGACO	GTTTTGCTGTCACI	TATAGATGG	ATCCGCGC	CGCATTAGCTAGTTGGTA	182
Query	181	AGGTAACGGCI	TACCAAGGCAACGA	TGCGTAGCC	GACCTGAG	AGGGTGATCGGCCACACT	240
Sbjct	183	AGGTAACGGCI	TACCAAGGCAACGA	TGCGTAGCC	GACCTGAG	AGGGTGATCGGCCACACT	242
Query	241	GGAACTGAGAG	ACGGTCCAGACTCC	TACGGGAGG	CAGCAGTA	GGGAATCTTCCGCAATGG	300
Sbjct	243	GGAACTGAGAG	ACGGTCCAGACTCC	TACGGGAGG	CAGCAGTA	GGGAATCTTCCGCAATGG	302
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Sbjct	303	GCGAAAGCCTG	ACGGAGCAACGCCG	CGTGAGTGA	IIIIIII IGAAGGTC	TTCGGATCGTAAAACTCT	362
Query	361	GTTATTAGGG	AGAACAAATGTGTA	AGTAACTAT	GCACGTCT	TGACGGTACCTAATCAGA	420
Sbjct	363	GTTATTAGGG	AGAACAAATGTGTA	AGTAACTAT	GCACGTCT	TGACGGTACCTAATCAGA	422
Query	421	AAGCCACGGCI	AACTACGTGCCAGC	AGCCGCGGT	ATACGTA	GGTGGCAAGCGTTATCCG	480
Sbjct	423	AAGCCACGGCI	AACTACGTGCCAGC	AGCCGCGGT	ATACGTA	GGTGGCAAGCGTTATCCG	482
Query	481	GAATTATTGGG	CGTAAAGCGCGCGT	AGGCGGTTT	TTAAGTC	IGATGTGAAAGCCCACGG	540
Sbjct	483	GAATTATTGGG	CGTAAAGCGCGCGCG	AGGCGGTTT	IIIIIII TTAAGTC	TGATGTGAAAGCCCACGG	542
Query	541	CTCAACCGTGG	AGGGTCATTGGAA	CTGGAAAAC	TTGAGTGC	AGAAGAGGAAAGTGGAAT	600
Sbjct	543	CTCAACCGTGG	AGGGTCATTGGAAZ	CTGGAAAAC	TTGAGTGC	AGAAGAGGAAAGTGGAAT	602
Query	601	TCCATGTGTAG	CGGTGAAATGCGC	GAGATATGG	AGGAACAC	CAGTGGCGAAGGCGACTT	660
Sbjct	603	TCCATGTGTAG	CGGTGAAATGCGCZ	GAGATATGG.	AGGAACAC	CAGTGGCGAAGGCGACTT	662
Query	661	TCTGGTCTGT	ACTGACGCTGATG	GCGAAAGCG	IGGGGATC	AAACAGGATTAGATACCC	720
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Query	721	TGGTAGTCCAC	GCCGTAAACGATGA	GTGCTAAGT	GTTAGGGG	GTTTCCGCCCCTTAGTGC	780
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Query	781	TGCAGCTAACG	CATTAAGCACTCC	CCTGGGGAG	TACGACCG	CAAGGTTGAAACTCAAAG	840
Sbjct	783	TGCAGCTAACO	GCATTAAGCACTCCC	CCTGGGGAG	TACGACCG	CAAGGTTGAAACTCAAAG	842
Query	841	GAATTGACGGG	GACCCGCACAAGC	GTGGAGCAT	GTGGTTTA	ATTCGAAGCAACGCGAAA	900
Sbjct	843	GAATTGACGGG	GACCCGCACAAGC	GTGGAGCAT	GTGGTTTA	ATTCGAAGCAACGCGAAG	902
Query	901	AACCTTACCAZ	ATCTTGACTTCCTC	TGACCCCTC	ТАААААТА 	AAGTTTTTCCCCTTCGGGG	960
Sbjct	903	AACCTTACCAZ	ATCTTGACATCCTC	TGACCCCTC	TAGAGATA	GAGTTTTCCCCTTCGGGG	962
Query	961	GAAAAAATGAG	AGG-GGGGCATGG-	TGTCGTCAG	CTCGGGTC	GGGAAATGTTGGGTTAAG	1018
Sbjct	963	GACAGAGTGAC	CAGGTGGTGCATGG	TGTCGTCAG	CTCGTGTC	GTGAGATGTTGGGTTAAG	1022
Query	1019	TCCC-CCACGZ	AGCGCAACCCTTAAG	CTTA 104	6		
Sbjct	1023	TCCCGCAACGA	GCGCAACCCTTAAG	CTTA 105	1		

	Childury 122183 Constraints NLA F at 1667 165 eitheomet BNA constraints and the constraints
	Checkham NLAE/2F708/105/10050mai RNA gene, partial sequence Uncultured bacterium clone nck307a03c1 16S ribosomal RNA gene, partial sequence
	Cincultured bacterium clone nbw1080g0521 16S ribosomal RNA gene, partial sequence Concentration of the second se
	Succerta + Reaves Succerta + Reaves
	Oncultured bacterium clone nbw1117e02c1 16S ribosomal RNA gene, partial sequence of the second
	 Uncultured bacterium clone nbw 1049h07c1 16S ribosomal RNA gene, partial sequ
	Uncultured bacterium clone ncd2635g07c1 16S ribosomal RNA gene, partial sequ
	Uncultured bacterium clone nbu314e00c1 165 ribosomal RNA gene, partial sequence
	Uncultured acterium clone ncd26/sa122/165 ribosonal RAA gene, partial sequence
	Staphylococcus epidermilit strain SEI, complete genome
	 Uncultured bacterium clone nck305a05c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck307a05c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck308/03c1 165 ribosomal RNA gene, partial sequence
	 Uncultured observation contents of 100xe1 to 5 mosconia RNA gene partial sequence Uncultured bacterium clone nex7774/661 (168 mbscome) ERA gene partial sequence
	Uncultured bacterium clone nek277/bloc 1 65 ribosonal RNA gene, partial sequence
	Uncultured bacterium clone nck277d09c1 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone nck277c11c1 165 ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone nck77/g0/21 10s inbosomal RNA gene, partial sequence Uncultured bacterium along and 2727/0421 10s inbosomal RNA gene, partial sequence
	 Uncultured bacterium concentration of the statistical sequences Incultured bacterium concentration of 126 schoomed 180A onen period
	Uncultured bacterium clone nck278/08c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck278g11c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck279h02c1 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone nck280g09c1 165 ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone nck281e11c1165 rhosomal RNA gene, partial sequence Doublured bacterium clone nck28207021162 chocomal RNA gene, partial sequence
04	 Uncultured bacterian concerned concerned to a monomal RNA gene, partial sequence Uncultured bacterian clone net/282007c1 165 ritosomal RNA energy and a sequence
	 Uncultured bacterium clone nck289/2121 16S ribosomal RNA gene, partial sequence
-	Uncultured bacterium clone nck290g11c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck290g06c1 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone net/294g01c1 165 ribosonal RNA gene, partial sequence Uncultured bacterium clone net/2020/161 165 chosenal RNA gene, partial sequence
	Uncultured bacteriani content network/2017c1 for 5 mostoral RNA series partial sequence Uncultured bacteriani classe net/2017c1 for 5 mostoral RNA series narrial semience
	 Uncultured bacterium clone nck304d12c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck305h06c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck306e02c1 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone nck307/d06c1 165 rbosomal RNA gene, partial sequence Uncultured bacterium clone nck3074 104 165 rbosomal RNA gene, partial sequence
	 The culture determine one net S080/67 (15) shows many seven partial sequence In culture determine one exist 3080/67 (15) shows many seven partial sequence
	Uncultured bacterium clone nck323c02c1 IOS ribosomal RNA seene, narial sequence
	Uncultured bacterium clone ncm66c07c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncm75a09c1 16S ribosomal RNA gene, partial sequence
	 Oncultured bacterium clone nck34311c1 165 nbosomal RNA gene, partial sequence
	 Staphylococcus sp. AMILSU(14) 105 mosomai KAA gene, partai sequence Staphylococcus and amilia territo (CCC 1004.156 cites) and partial sequence
	Starby records and provide star and the star of the st
	Staphylococcus epidermidis strain PBR-19 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. CIFE_HT11 16S ribosomal RNA gene, partial sequence
	 Staphylococcus sp. PJSI46 16S ribosomal RNA gene, partial sequence
	 Stapitylococcus sp. IZ/ 105 mosomai KNA gene, partai sequence Stapitylococcus sp. IZZ/165 mosomai KNA gene, partai sequence
	Starby locacias ender miles train As-3 los ritoronal reverse partial sequence
	Staphylococcus epidermidis strain C6/2 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain GKY 4/1A 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain P 1/1 16S ribosomal RNA gene, partial sequence
	• Stabilización solitaria y $M = 10^{-10}$ (100 m molecular de la comparación de la
04	Starby tooccus endermitis strain GVV is 11 bits for boots and Nov gene, partial sequence Analysis strain GVV is 11 bits risks and Nov gene, partial sequence
	Bacterium FM-2014-25 senomic TNA containing 16S-23S intervenic spacer region, isolate 25
	Uncultured bacterium eene for 165 rRNA, partial sequence, clone: smkt Fir 002 001
	Uncultured bacterium gene for 16S rRNA, partial sequence, clone: smkt Fir 002 002
	Uncultured Staphylococcus sp. clone 12L_33 16S ribosomal RNA gene, partial sequence
	Staphylococcus epidermidis strain CIFRI D-TSB-84 16S ribosomal RNA gene, partial sequence
	 Staphylecoccus epidermidis strain MB2MH4 16S ribosomal RNA gene, partial sequence
	 Staphylococcus epidermains strain KSS111/105 InfoSomal KNA gene, partial sequence Staphylococcus epidermidis errini FSZ12111 (SE Visconal PIXA acone partial acoustros
	 Sublycoccus endermidi strain DFGFT 2755 165 (Shoonal RNA one partial sequence
	Samby lococcus enidermilis strain 252 Cost loss of the New Entry Strain a sequence
	Staphylococcus epidermidis strain ZJY-109 16S ribosomal RNA gene, partial sequence
	Apteryx australis mantelli genome assembly AptMant0, scaffold scaffold32554
	Staphylococcus epidermidis strain TWSL_16 16S ribosomal RNA gene, partial sequence
	Staphylococcus sp. TT31 I6S ribosomal RNA gene, partial sequence
	 Sackernum msi i-1 los ribosomal KNA gene, partial sequence Sackernum gene (2017) El schargene (2014)
	 stamptynocecus sp. 5407 105 mosomaa Kryw gene, partaa sequence Stamptynocecus sp. sidermidis certain Restoris VIII Into Schoomaa DRA onto avarial companya
	Supervision and the state of the state
	Staphylococcus epidermidis strain HUC2 27 K 165 ribosomal RNA eene, nartial sequence
	Staphylococcus epidermidis strain LIPOAUEL 42 16S ribosomal RNA gene, partial sequence
	🜷 Uncultured bacterium clone nck292/09c1 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone nck290h07c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck280b0/c1 165 ribosomai RNA gene, partial sequence
	 Uncultured bacterium clone nck278b1c1 165 ribosomal RNA gene, partial sequence Uncultured bacterium clone nck278b1c1 165 ribosomal RNA gene, partial sequence
	Uncultured between addentian close teck/strivel to structorian KNA gene, partial sequence ■ Incultured between used technical sequences ■ Incultured between technical sequences ■ I
	Uncultured bacteriant const texts a source 1 to 100000mul RNA egget, partial sequence Uncultured bacteriant const texts a source 1 to 100000mul RNA egget a ratial sequence
	Uncultured bacterium clone nck259/11/21/65/fibosonal RNA ene, partial sequence
	Uncultured bacterium clone nck187h10c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck186a05c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck185h04c1 16S ribosomal RNA gene, partial sequence
1	firmicutes 2 leaves

Rothia amarae NR_029045.1

Rothia amarae strain J18 16S ribosomal RNA gene, partial sequence Sequence ID: <u>ref[NR_029045.1]</u> Length: 1467 Number of Matches: 1 <u>See 1 more title(s)</u>

Range 1: 98 to 1028 GenBank Graphics Vext Match 🔺 Previous Match					
Score 1644 bit	s(1822)	Expect Identities 0.0 924/931(99%)	Gaps 1/931(0%)	Strand Plus/Plus	
Query	1	GAACGGGTGAGTAATACGTGAGTAA	ACCTGCCTTTGACTC	TGGGATAAGCCTGGGAAACTG	60
Sbjct	98	GAACGGGTGAGTAATACGTGAGTA	ACCTGCCTTTGACTC	TGGGATAAGCCTGGGAAACTG	157
Query	61	GGTCTAATACCGGATATGACAAGGA	ACCGCATGGTTTTT	TGTGGAAAGGGTTTGTACTGG	120
Sbjct	158	GGTCTAATACCGGATATGACAAGGA	ACCGCATGGTTTTI	TGTGGAAAGGGTTTGTACTGG	217
Query	121	TTTTAGATGGGCTCACGGCCTATC	AGCTTGTTGGTGGGG	TAATGGCTCACCAAGGCGACG	180
Sbjct	218	TTTTAGATGGGCTCACGGCCTATCA	AGCTTGTTGGTGGGG	TAATGGCTCACCAAGGCGACG	277
Query	181	ACGGGTAGCCGGCCTGAGAGGGTGA		ACTGAGACACGGCCCAGACTC	240
Sbjct	278	ACGGGTAGCCGGCCTGAGAGGGTGA	ACCGGCCACACTGGG	ACTGAGACACGGCCCAGACTC	337
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Query	301	GCGTGAGGGATGACGGCCTTCGGG	TGTAAACCTCTTTC	AGCAGGGAAGAAGCGAAAGTG	360
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Sbjct	578	CTGTGAAAGACCGGGGCTTAACCCC	CGGTATTGCAGTGGG	TACGGGCAGACTAGAGTGCAG	637
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Query	601	ATGGCGAAGGCAGGTCTCTGGGCTG	TAACTGACGCTGAG	AAGCGAAAGCATGGGGAGCGA	660
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Query	721	TTCCACGTTTTTCCGCGCCGTAGCTA	ACGCATTAAGTGCC	CCGCCTGGGGAGTACGGCCGC	: 780
Sbjct	818	TTCCACGTTTTCCGCGCCGTAGCTA	ACGCATTAAGTGCC	CCGCCTGGGGGAGTACGGCCGC	877
Query	781	AAGGCTAAAACTCAAAGGAATTGAC	GGGGGGCCCGCACAA	GCGGCGCACCATGCGGATTAA	840
Sbjct	878	AAGGCTAAAACTCAAAGGAATTGAC	GGGGGGCCCGCACAA	GCGGCGGAGCATGCGGATTAA	937
Query	841	TTCGATGCAACGCGAAAAACCTTAC	CAAGGCTTGACATA	TACTGGACCGCCTCAAAAATG	; 900
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Query	901	GGGTTTCCCTTC-GGGCTGGTATAC	AGGGGG 930		
Sbjct	998	GGGTTTCCCTTCGGGGCTGGTATAC	AGGTGG 1028		

	Uncultured bacterium clone nck132g02c1 16S ribosomal RNA gene, partial sequence
5	Rothia amarae strain J18 16S ribosomal RNA gene, nartial sequence
	Uncultured bacterium clone ncd401f11c1 16S ribosomal RNA gene, partial sequence
	Bacterium 3D705 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone ncd1418c10c1 16S ribosomal RNA gene, partial sequence
	abteria 8 leaves
	Bacterium RUSp01-4 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck32h02c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck30h04c1 16S ribosomal RNA gene, partial sequence
	Rothia amarae partial 16S rRNA gene, strain R-36507
	Uncultured bacterium clone ncd1336a11c1 16S ribosomal RNA gene, partial sequence
	Rothia sp. RV13 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncd401g09c1 16S ribosomal RNA gene, partial sequence
	• Icl[Query 200873
	Uncultured Alteromonadaceae bacterium clone WEIA1 16S ribosomal RNA gene, partial sequence
	Kocuria sp. Am 16 partial 16 16S rRNA gene, isolate Am 16
	Bacterium WP2ISO15 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nck30e06c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncd402c01c1 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone ncd401c01c1 I6S ribosomal RNA gene, partial sequence
	Bacterium RUSp03-2 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncd1398604c1 16S ribosomal RNA gene, partial sequence
	Rothia amarae partial 16S rRNA gene, strain 0410ARD15N_5
	bacteria 29 leaves
	Rothia mucilaginosa strain CMC18 16S ribosomal RNA gene, partial sequence
	Rothia sp. 6813c 16S ribosomal RNA gene, partial sequence
	A bacteria 3 leaves
	 Uncultured bacterium clone ncd1208a07c1 16S ribosomal RNA gene, partial sequence
	Rothia sp. BBH4 partial 16S rRNA gene, strain BBH4
0.002	bacteria 34 leaves

Delftia lacustris KT958881.1

Delftia lacustris strain BMCH-IB-C 142 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KT958881.1|</u> Length: 1360 Number of Matches: 1

Range 1: 355 to 1340 GenBank Graphics V Next Match 🛦 Previous Match						
Score 1745 bit	s(1934)	Expect 0.0	Identities 980/986(99%)	Gaps 3/986(0%)	Strand Plus/Minus	
Query	1	CCTACTTCTGGC	GAGACCCGCTCCCAI	IGGTGTGACG	GGCGGTGTGTACAAGACCCGGGAA	60
Sbjct	1340	CCTACTTCTGGC	CGAGACCCGCTCCCAT	GGTGTGACG	GCCGTGTGTACAAGACCCGGGAA	1281
Query	61	CGTATTCACCGC	GGCATGCTGATCCGC	CGATTACTAG	CGATTCCGACTTCACGCAGTCGAG	120
Sbjct	1280	CGTATTCACCGC	GGCATGCTGATCCGC	CGATTACTAG	CGATTCCGACTTCACGCAGTCGAG	1221
Query	121	TTGCAGACTGCG	ATCCGGACTACGACI	GGTTTTATG	GGATTAGCTCCCCCTCGCGGGTTG	180
Sbjct	1220	TTGCAGACTGCG	GATCCGGACTACGACI	GGTTTTATG	GGATTAGCTCCCCCTCGCGGGTTG	1161
Query	181	GCAACCCTCTGT	ACCAGCCATTGTAT	GACGTGTGTA	GCCCCACCTATAAGGGCCATGAGG	240
Sbjct	1160	GCAACCCTCTGI	ACCAGCCATTGTAT	ACGTGTGTA	GCCCCACCTATAAGGGCCATGAGG	1101
Query	241	ACTTGACGTCAT	CCCCACCTTCCTCC	GTTTGTCAC	CGGCAGTCTCATTAGAGTGCTCAA	300
Sbjct	1100	ACTTGACGTCAT	CCCCACCTTCCTCC	GTTTGTCAC	CGGCAGTCTCATTAGAGTGCTCAA	1041
Query	301	CTGAATGTAGCA	ACTAATGACAAGGGI	TGCGCTCGT	TGCGGGACTTAACCCAACATCTCA	360
Sbjct	1040	CTGAATGTAGCA	ACTAATGACAAGGGI	TGCGCTCGT	TGCGGGACTTAACCCAACATCTCA	981
Query	361	CGACACGAGCTO	BACGACAGCCATGCAG	GCACCTGTGT	GCAGGTTCTCTTTCGAGCACGAAT	420
Sbjct	980	CGACACGAGCTO	BACGACAGCCATGCAG	GCACCTGTGT	GCAGGTTCTCTTTCGAGCACGAAT	921
Query	421	CCATCTCTGGAA	ACTTCCTGCCATGTC	CAAAGGTGGG	TAAGGTTTTTTCGCGTTGCATCGAA	480
Sbjct	920	CCATCTCTGGAA	ACTTCCTGCCATGTC	CAAAGGTGGG	TAAGGTTTTTCGCGTTGCATCGAA	861
Query	481	TTAAACCACATO	ATCCACCGCTTGTGC	CGGGTCCCCG	TCAATTCCTTTGAGTTTCAACCTT	540
Sbjct	860	TTAAACCACATO	ATCCACCGCTTGTGC	GGGTCCCCG	TCAATTCCTTTGAGTTTCAACCTT	801
Query	541	GCGGCCGTACTC	CCCAGGCGGTCAACI	TCACGCGTT.	AGCTTCGTTACTGAGAAAACTAAT	600
Sbjct	800	GCGGCCGTACTC	CCCAGGCGGTCAACI	TCACGCGTT.	AGCTTCGTTACTGAGAAAACTAAT	741
Query	601	TCCCAACAACCA	GTTGACATCGTTTAG	GGCGTGGAC	TACCAGGGTATCTAATCCTGTTTG	660
Sbjct	740	TCCCAACAACCA	GTTGACATCGTTTAG	GGCGTGGAC	TACCAGGGTATCTAATCCTGTTTG	681
Query	661	CTCCCCACGCTI	TCGTGCATGAGCGTC	AGTACAGGT	CCAGGGGATTGCCTTCGCCATCGG	720
Sbjct	680	CTCCCCACGCTI	TCGTGCATGAGCGTC	AGTACAGGT	CCAGGGGATTGCCTTCGCCATCGG	621
Query	721	TGTTCCTCCGCA	TATCTACGCATTTCA	CTGCTACAC	GCGGAATTCCATCCCCCTCTACCG	780
Sbjct	620	TGTTCCTCCGCA	TATCTACGCATTTCA	ACTGCTACAC	GCGGAATTCCATCCCCCTCTACCG	561
Query	781	TACTCTAGCCAT	GCAGTCACAAATGCA	GTTCCCAGG	TTGAGCCCGGGGGATTTCACATCTG	840
Sbjct	560	TACTCTAGCCAI	GCAGTCACAAATGCA	GTTCCCAGG	TTGAGCCCGGGGGATTTCACATCTG	501
Query	841	TCTTACATAACC	GCCTGCGCACGCTTI	ACGCCCAGT.	AATTCCGATTAACGCTCGCACCCT	900
Sbjct	500	TCTTACATAACC	GCCTGCGCACGCTTI	ACGCCCAGT.	AATTCCGATTAACGCTCGCACCCT	441
Query	901	ACGTATTACCGC	GGCTGCTGGCACGTA	ATTAGCCGG	GGCTTATTCTTACGGTACCGTCAT	960
Sbjct	440	ACGTATTACCGC	GGCTGCTGGCACGTA	AGTTAGCCGG	TGCTTATTCTTACGGTACCGTCAT	381
Query	961	GGG-CCCCTGTA	ATTAAAGAGCTTT	983		
Sbjct	380	GGGCCCCCTGTA	TTAGAAGGAGCTTT	355		

	• kilouery 4095
0	Dethia sp. PVRh-YHB-9-1 16S ribosomal RNA gene, partial sequence
1	Bacterium RRP2-22 16S rRNA gene
	Delftia sp. AN3 16S ribosomal RNA gene, partial sequence
	Delftia sp. 6.13 16S ribosomal RNA gene, partial sequence
	Buncultured bacterium partial 16S rRNA gene, clone SMQ25
	Delftia sp. BN-HKY2 16S ribosomal RNA gene, partial sequence
	Delfia tsuruhatensis strain MTQ1 16S ribosomal RNA gene, partial sequence
	Delftia sp. JDC-3 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 6 16S ribosomal RNA gene, partial sequence
	b-proteobacteria 3 leaves
	Delftia tsuruhatensis strain P027 16S ribosomal RNA gene, partial sequence
	Delftia tsuruhatensis strain D10 16S ribosomal RNA gene, partial sequence
	Delftia sp. enrichment culture clone 12 16S ribosomal RNA gene, partial sequence
	Delftia sp. SS12.34 16S ribosomal RNA gene, partial sequence
	Delftia tsuruhatensis strain IPPBC R15 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone A5 16S ribosomal RNA gene, partial sequence
	Delftia sp. BGW1 16S ribosomal RNA gene, partial sequence
	Delftia sp. DM101 16S ribosomal RNA gene, partial sequence
	Delftia sp. SFG 16S ribosomal RNA gene, partial sequence
	Bacterium BM0414 16S ribosomal RNA gene, partial sequence
	Delftia lacustris voucher RIFA 1193 16S ribosomal RNA gene, partial sequence
	 Deffia sp. X-a12 16S ribosomal RNA gene, partial sequence
	Delftia tsuruhatensis strain CI-23 16S ribosomal RNA gene, partial sequence
	Bacterium WYLW2-4 16S ribosomal RNA gene, partial sequence
	Uncultured Delftia sp. clone GI5-002-F11 I6S ribosomal RNA gene, partial sequence
	Delitia isuruhatensis strain A90 165 ribosomal RNA gene, partial sequence
	Cincultured bacterium gene for 165 rRNA, partial sequence, clone: SSmCB08-/4
	Delita sp. PHD-6 165 ribosomal RNA gene, partial sequence
	 Defina sp. AEX21 105 mossimal KXA gene, partai sequence
	 Unitational consistence of the State of the
	Che mured organism clone (ELCO) 27-1314-5-NI (ONZO) small subunit nobolmal KNA gene, partial sequence
	Definite actustitis strain BMCH-11B-C 142 105 ribosomai KNA gene, partial sequence
	Bacterium BM0429 10S nbosomal RNA gene, partial sequence
	b postediciental 3 leaves
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	 Definition of the second /li>
	verina isurunalensis strain L/ Tos ribosomal KNA gene, partial sequence
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	 Defina Surunaensis strain 5109 105 mosomai KivA gene, partai sequence
	Circultured bacterium clone BC_B1_121105 ribosomai KivA gene, partial sequence
	Uncultured bacterium gene for 165 rRNA, partial sequence, clone: U-1
	 Delta lacustris stran ABBs012-01 165 nbosonal KNA gene, partial sequence
	Delita sp. 20/2/n-As too mosonial RNA gene, partial sequence
	Definita lacustris strain (AKV-NIAW 1-34 105 tribosomai KNA gene, partial sequence
	a baccrum A (394,2011) 105 mbosonai KNA gene, partial sequence
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	Central sourcialensis strain JPK21 105 (noisontai RNA gene, partai sequence Destaving BM0421 165 cibes and 100Å onto a restil a contrati e
	b meteoristical (Composition RAA gene, partial sequence
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b-proteobacteria | 5 leaves

Pseudomonas aeruginosa KF680991.1

Pseudomonas aeruginosa strain ATHA23 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb[KF680991.1]</u> Length: 1115 Number of Matches: 1

Range 1:	Range 1: 98 to 1040 GenBank Graphics Vext Match 🛦 Previous Match						
Score 1649 bit	s(1828)	Expect 1 0.0	(dentities 933/943(99%)	Gaps 2/943(0%)	Strand Plus/Minus		
Query	1	GTGATTGGCGGT	GTGTACAAGGCCC	GGGAACGTATTCA		ATTCACGAT	60
Sbjct	1040	GTGACGGGCGGT	GTGTACAAGGCCC	GGGAACGTATTCA	CCGTGACATTCTG	ATTCACGAT	981
Query	61	TACTAGCGATTCO	CGACTTCACGCAG	CGAGTTGCAGAC		ACGATCGGT	120
Sbjct	980	TACTAGCGATTCO	CGACTTCACGCAG	ICGAGTTGCAGAC	TGCGATCCGGACT	ACGATCGGT	921
Query	121		GCTCCACCTCGCG	GCTTGGCAACCCT	TTGTACCGACCAT	IGTAGCACG	180
Sbjct	920	TTTATGGGATTAG	GCTCCACCTCGCG	GCTTGGCAACCCT	TTGTACCGACCATI	IGTAGCACG	861
Query	181	TGTGTAGCCCTG	GCCGTAAGGGCCA	IGATGACTTGACG	TCATCCCCACCTT	CCTCCGGTT	240
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Query	241	TGTCACCGGCAG	CTCCTTAGAGTG	CCCACCCGAGGTG	CTGGTAACTAAGG	ACAAGGGTT	300
Sbjct	800	TGTCACCGGCAG	CTCCTTAGAGTG	CCCACCCGAGGTG	CTGGTAACTAAGG	ACAAGGGTT	741
Query	301	GCGCTCGTTACG	GACTTAACCCAA	CATCTCACGACAC	GAGCTGACGACAG	CCATGCAGC	360
Sbjct	740	GCGCTCGTTACG	GACTTAACCCAA	CATCTCACGACAC	GAGCTGACGACAG	CCATGCAGC	681
Query	361	ACCTGTGTCTGAG	GTTCCCGAAGGCA	CCAATCCATCTCT		CATGTCAAG	420
Sbjct	680	ACCTGTGTCTGAC	STTCCCGAAGGCA	CCAATCCATCTCT	GGAAAGTTCTCAG	CATGTCAAG	621
Query	421	GCCAGGTAAGGT	CTTCGCGTTGCT	ICGAATTAAACCA	CATGCTCCACCGC	TGTGCGGG	480
Sbjct	620	GCCAGGTAAGGT	CTTCGCGTTGCT	ICGAATTAAACCA	CATGCTCCACCGCI	TTGTGCGGG	561
Query	481	CCCCCGTCAATTO	CATTTGAGTTTTA	ACCTTGCGGCCGT	ACTCCCCAGGCGG	ICGACTTAT	540
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Query	541	CGCGTTAGCTGCG	CCACTAAGATCT	CAAGGATCCCAAC	GGCTAGTCGACAT	CGTTTACGG	600
Sbjct	500	CGCGTTAGCTGCG	CCACTAAGATCT	CAAGGATCCCAAC	GGCTAGTCGACATO	CGTTTACGG	441
Query	601	CGTGGACTACCAG	GGTATCTAATCC	rgtttgctcccca	CGCTTTCGCACCTC	CAGTGTCAG	660
Sbjct	440	CGTGGACTACCAG	GGTATCTAATCC	IGTTTGCTCCCCA	CGCTTTCGCACCTC	CAGTGTCAG	381
Query	661	TATCAGTCCAGGT	GGTCGCCTTCGC	CACTGGTGTTCCT		CATTTCACC	720
Sbjct	380	TATCAGTCCAGGT	GGTCGCCTTCGC	CACTGGTGTTCCT	TCCTATATCTACGO	CATTTCACC	321
Query	721	GCTACACAGGAAA	TTCCACCACCCT	CTACCGTACTCTA		GATGCAAT	780
Sbjct	320	GCTACACAGGAAA	TTCCACCACCCT	CTACCGTACTCTA	GCTCAGTAGTTTTC	GGATGCAGT	261
Query	781	TCCCAGGTTGAGC	CCGGGGGATTTCA	CATCCAACTTGCT	GAACCACCTACGC	GCGCTTTAC	840
Sbjct	260	TCCCAGGTTGAGC	CCGGGGGATTTCA	CATCCAACTTGCT	GAACCACCTACGC	GCGCTTTAC	201
Query	841	GCCCAGTAATTCC	GATTAACCCTTG	CACCCTTCGTATT	ACCGCGGCTGCTG	GCCCGAAGT	900
Sbjct	200	GCCCAGTAATTCC	GATTAACGCTTG	CACCCTTCGTATT	ACCGCGGCTGCTG	GCACGAAGT	141
Query	901	TACCCGG-GCTTA	ATTCTGTTGG-AAG	CGTCAAAAAGCAA	.GGGAT 941		
Sbjct	140	TAGCCGGTGCTTA	TTCTGTTGGTAA	CGTCAAACAGCAA	GGTAT 98		

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	τφ	Preudomonas acrueinosa strain Carb01 63. complete genome
		Pseudomonas aeruginosa DK1 genome assembly Pseudomonas aeruginosa DK1 substr. NH57388A, chromosome
		Pseudomonas aeruginosa strain TH4 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA107 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA110 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA116 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA129 16S ribosomal RNA gene, partial sequence
		 Pseudomonas aerugmosa strain NA152 105 ribosomal RNA gene, partial sequence Dendemona province strain NA154 165 ribosomal RNA gene, partial sequence
		Pseudomonas aerugmosa strain NA154 105 hibosomal PNA gene, partial sequence
		Preudomonas aeruginosa strain NATSO TOS INOSOnal RNA gene, partial sequence
		Beudomonas aeruginosa strain NA139 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA140 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA141 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA143 16S ribosomal RNA gene, partial sequence
		🛢 Pseudomonas aeruginosa strain NA144 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA145 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain NA146 16S ribosomal RNA gene, partial sequence
		 rseucomonas acruginosa strain NA160/16S ribosomai RNA gene, partial sequence Dandama servição a strain NA160/16S ribosomai RNA gene, partial sequence
		 rseudomonas aerugmoša štrain NA165 165 ribosomai KNA gene, partial sequence Dandom partije sequence
		Preudomonas acroganosa strata (NATOS TOS TOS OBSONIA) KIVA gene, partial sequence
		Preudomonas aerueinosa strain Ps15 16S ribosonal RNA cene, nartial seguence
		Pseudomonas aeruginosa strain Ps8 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain Ps6 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain ANF2 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain PS35 16S ribosomal RNA gene, partial sequence
—		Pseudomonas aeruginosa gene for 16S ribosomal RNA, partial sequence, strain: PD29
		Pseudomonas aeruginosa gene for 16S ribosomal RNA, partial sequence, strain: BSS01
		Pseudomonas aeruginosa strain PKS 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain DASE-WMI 165 ribosomai RNA gene, partial sequence
		PSCUCOMONAS actualitos natival 165 INVA gene, isolate SHO
		Bacterium YY3 165 ribosomal RNA seene, partial sequence
		Preudomonas aerueinosa strain ZM130 (55 ribasomal RNA cene, partial sequence
		Pseudomonas aeruginosa strain S7PS5 16S ribosomal RNA gene, partial sequence
		Staphylococcus sciuri strain SBANHCa14 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain DSM 50071 16S ribosomal RNA gene, partial sequence
		9 Pseudomonas aeruginosa strain 12 6 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 12 2 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain GS-33 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain GS-31 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain JBL4 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain HBUZL-6 I6S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain JBL10 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain JBL8 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain JBL/ 165 nbosomai RNA gene, partai sequence
		Preudomonas aeruginosa suran IDELDSD 105 105500 nar KAVA gene, partan sequence
		Pseudomonas aeruginosa strain DELC TOS niosomai RNA gene, partia sequence
		Pseudomonas aeruginos strain 2.5.5 165 ribosonal RNA gene, partai sequence
		Pseudomonas aeruginosa strain 2-4-b-1 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 1-3-b-2 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 1-4-b-1 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 1-5-b-2-1 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 1-4-b-9 16S ribosomal RNA gene, partial sequence
		Seudomonas aeruginosa strain 3-5-b-5 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 3-4-b-2 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 3-4-b-1 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain 3-3-b-2 16S ribosomal RNA gene, partial sequence
		Pseudomonas aerueinosa strain Z158 16S ribosomal RNA eene, partial sequence
		Pseudomonas sp. VIH2 16S ribosomal RNA gene, partial sequence Pseudomonas apprintere attain ZIX 486 16S ribosomal RNA area pratial sequence
		 Preudomonas acruganosa sirian 23 1 - 480 105 noosomal KNA gene, partial sequence Brandomonas acruganosa etasia (21) \$52 165 siboromal DNA canto partial capation capationes
		Pseudomonas aerueinos strain 221-535 tos moscular RAS ene, partial sequence
		Pseudomonas aeruginosa strain ZIY-522 165 ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain ZJY-428 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain ZJY-959 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain ZJY-226 16S ribosomal RNA gene, partial sequence
		Pseudomonas aeruginosa strain RRLP1 16S ribosomal RNA gene, partial sequence
		 Pseudomonas aeruginosa strain SM-1 16S ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain SA-16 for the sequence
		 Pseudomonas aeruginosa strain SM-1 IOS ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain Sp-A IOS ribosomal RNA gene, partial sequence Pseudomonas aeruginosa ctrain EV IOS ribosomal RNA gene, partial sequence
		Pecudomonas acruginosa strain SNA 165 ribosomal RNA gene, partial sequence Pecudomonas acruginosa strain SPA 165 ribosomal RNA gene, partial sequence Pecudomonas acruginosa strain BEL8 165 ribosomal RNA gene, partial sequence Pecudomonas acruginosa strain EBL8 165 ribosomal RNA gene, partial sequence
		 Pecudomonas aeruginosa strain SM-1165 nhosomal RNA gene, partial sequence Pecudomonas aeruginosa strain EBLA 165 nhosomal RNA gene, partial sequence Pecudomonas aeruginosa strain EBLA 165 nhosomal RNA gene, partial sequence Pecudomonas aeruginosa strain EBLA 165 nhosomal RNA gene, partial sequence Pecudomonas aeruginosa strain EBLA 165 nhosomal RNA gene, partial sequence
		 Pecudomona seruginos strain SM-116S rithosomal RNA gene, partial sequence Pecudomonas aeruginos strain EDLS 16 for thosomal RNA gene, partial sequence Pecudomonas aeruginos strain EDLS 16 for thosomal RNA gene, partial sequence Pecudomonas aeruginos strain EDLS 16 for thosomal RNA gene, partial sequence Pecudomonas aeruginos strain EDLS 16 for thosomal RNA gene, partial sequence Pecudomonas aeruginos strain EDLS 16 for thosomal RNA gene, partial sequence Pecudomonas aeruginos strain EDLS 16 for thosomal RNA gene, partial sequence
		 Pecudomona seruginos strain SM-116S finosomal RNA gene, partial sequence Pecudomonas aeruginos strain ISLA 16 finosomal RNA gene, partial sequence Pecudomonas aeruginos strain IELLS 16 finosomal RNA gene, partial sequence Pecudomonas aeruginos strain IELLS 16 finosomal RNA gene, partial sequence Pecudomonas aeruginos strain IELS 16 finosomal RNA gene, partial sequence Pecudomonas aeruginos strain IELS 16 finosomal RNA gene, partial sequence Pecudomonas aeruginos strain IELS 16 finosomal RNA gene, partial sequence Pecudomonas aeruginos strain IELS 16 finosomal RNA gene, partial sequence
		 Pecudomona seruginos strain SM-116S finosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R6 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R5 hosomal RNA gene, partial sequence Pecudomonas aeruginos strain EJLS 16 R5 hosomal RNA gene, partial sequence
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Bacterium WUI 105 ribosomal RNA gene, partial sequence
 Peadomonas aeruginosa straia ATHA23 165 ribosomal RNA gene, partial sequence
 Peadomonas aeruginosa straia ATA19 105 ribosomal RNA gene, partial sequence
 Peadomonas aeruginosa straia BM0 165 ribosomal RNA gene, partial sequence
 Peadomonas aeruginosa straia BM0 165 ribosomal RNA gene, partial sequence
 Peadomonas aeruginosa straia BM0 165 ribosomal RNA gene, partial sequence
 Peadomonas aeruginosa straia BM0 165 ribosomal RNA gene, partial sequence
 Peadomonas aeruginosa straia BM0 165 ribosomal RNA gene, partial sequence
 Bacterium CD7 165 ribosomal RNA, partial sequence

domonas aeruginosa strain VV01 16S ribosomal RNA gene, partial se

JX090199.1

Range 1: 399 to 1347 GenBank Graphics Next Match Previous Match							
Score 1692 bit	s(1876)	Expect 0.0	Identities 945/949(99%)	Gaps 1/949(0%)	<mark>Strand</mark> Plus/Minus		
Query	1	TCCCATGGTGTC	ACGGGCGGTGT	GTACAAGACCCGG	GAACGTATTCACCO	CGGCATGCTG	60
Sbjct	1347	TCCCATGGTGT	GACGGGCGGTGT	GTACAAGACCCGG	GAACGTATTCACC	CGGCATGCTG	1288
Query	61	ATCCGCGATTA	CTAGCGATTCCG	ACTTCACGCAGTC	GAGTTGCAGACTG	GATCCGGACT	120
Sbjct	1287	ATCCGCGATTA	CTAGCGATTCCG	ACTTCACGCAGTC	GAGTTGCAGACTG	CGATCCGGACT	1228
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Query	661	AGCGTCAGTACA	GGTCCAGGGGA	TTGCCTTCGCCAT	CGGTGTTCCTCCGC	ATATCTACGC	720
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Sbjct	567	AATGCAGTTCCC	AGGTTGAGCCC	GGGATTTCACAT	CTGTCTTACATAAC	CGCCTGCGCA	508
Query	840	CGCTTTACGCCC	AGTAATTCCGAT	TTAACGCTCGCAC	CCTACGTATTACCO	CGGCTGCTGG	899
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Sbjct	447	CACGTARTTAGO	CGGTGCTTATT	CTTACGGTACCGT	CATGGGCCTCTCG	399	

Delftia acidovorans strain Aal-4 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|JX090199.1</u>| Length: 1412 Number of Matches: 1

	Icl Query_1	108535
2	-0	Delftia acidovorans strain AaI-4 16S ribosomal RNA gene, partial sequence Companyonas sp. 16S rRNA gene isolate 158
	Τ	Uncultured bacterium clone rRNA334 16S ribosomal RNA gene, partial sequence
		Uncultured Delfuia sp. clone EMP_K41 16S ribosomal RNA gene, partial sequence
		 Uncultured bacterium clone 16slp116-5a09.p1k 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone 16slp116-5a09.p1k 16S ribosomal RNA gene, partial sequence
		 Uncultured oreanism clone FODP_305_1509D_water_sample_1200mbst_2105 ftbosomat RNA gene, partial sequence Uncultured oreanism clone FLU0008-158-SNI 000061 small subunit ribosomat RNA gene, partial sequence
		Uncultured organism clone ELU0181-T360-S-NIPCRAMgANa_000424 small subunit ribosomal RNA gene, partial sequence
		Uncultured organism clone EL U0034-T174-S-NIPCRAMgANb 000300 small subunit ribosomal RNA gene, partial sequence
		Delfa so PPA S 16S ribosomal RNA gene partial sequence
		Bacterium KR 1996/1 33 16S ribosomal RNA gene, partial sequence
		Bacterium KR 1996/1_185 16S ribosomal RNA gene, partial sequence
		Delftia acidovorans strain A-B13B 16S ribosomal RNA gene, partial sequence Delftia Marcanal RNA gene partial sequence
		Uncultured bacterium gene for 165 rRNA, partial sequence
		Delftia sp. SR27 16S ribosomal RNA gene, partial sequence
		Bacterium KR 1996/1_115 16S ribosomal RNA gene, partial sequence
		Defilitia sp. PP4_S7 16S ribosomal RNA gene, partial sequence d localized arganism clone E1 10034.174.5 NIDCR AM6 AND 000657 small subunit ribosomal RNA gene, partial sequence
		 Uncultured organism clone EL 0003+11+174-S-NI PCRAMgANb 000415 small subunit ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000286 small subunit ribosomal RNA gene, partial sequence
		Duffit loanting the second
		Delfua lacustrs strain 05 105 ribosomat Riva gene, partial sequence
		Uncultured bacterium clone 16slp99-2e12.p1k 16S ribosomal RNA gene, partial sequence
		b-proteobacteria 2 leaves
		Delfita acidovorans strain B201 16S ribosomal RNA gene, partial sequence Delfita so he-1 16S ribosomal RNA gene, partial sequence
0.0008		Bacterium KR 1996/1 116 16S ribosomal RNA gene, partial sequence
		Delftia sp. PP4_S6 16S ribosomal RNA gene, partial sequence
		Delftia sp. BFE58M 16S ribosomal RNA gene, partial sequence
		Delée CR02 (16) Accession (18) Ac
		Bacterium KR 1996/1 115 165 ribosoma KNA gene, partial sequence Bacterium KR 1996/1 115 165 ribosoma IRNA eene, partial sequence
		Delfiia sp. PP4_S7 16S ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000657 small subunit ribosomal RNA gene, partial sequence
		 Uncultured organism clone ELU0034-11/4-S-NIPCRAMgAND (00/415 small subunit ribosomal KNA gene, partial sequence Uncultured organism clone ELU0032-11/4-S-NIPCRAMgAND (00/286 cmall cubunit ribosomal RNA gene, partial sequence
		 Uncultured organism clone EL00157-1387-S-NIP CRAMgAND_00/200 small subunit noisonial RNA gene, partial sequence Uncultured organism clone EL00157-1387-S-NIP CRAMgAND_00/200 small subunit noisonial RNA gene, partial sequence
		Pelfia lacustris strain 6S 16S ribosomal RNA gene, partial sequence
		Delfia sp. R-41383 partial 16S rRNA gene, strain R-41383
		b-protokaterial 2 leaves
		 Delfitia acidovorans strain B201 16S ribosomal RNA gene, partial sequence
		Delfia sp. hg-1 16S ribosomal RNA gene, partial sequence
		Bacterium KR 1996/1_116 16S ribosomal KNA gene, partial sequence Delfai en DPM 56 16S ribosomal RNA gene, partial sequence
		Uncultured beta proteobacterium clone Alchichica AL67 2 1B 104 small subunit ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000365 small subunit ribosomal RNA gene, partial sequence
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		Alcalipenes facelis strain 18A 16S ribosomal RNA gene partial sequence
		bacteria 4 leaves
		Delfia sp. KAR43 16S ribosomal RNA gene, complete sequence
		Delfita isuruhatensis sirain X15 165 ribosomal KNA gene, partial sequence
		Bacterium KR 1996/1 85 165 ribosonal RNA gene, partal sequence
		Delfiia sp. PP4_S3 16S ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000557 small subunit ribosomal RNA gene, partial sequence
		 Uncultured organism crione ELC003+117+5-SUPCKAMIgAND 00/201 small subunit ribosomal RNA gene, partial sequence Uncultured organism crione ELC0182-T110-SNIPCRAMgAND 00/010 small subunit ribosomal RNA gene, partial sequence
		O Locultured organism clone EL L0008-T58-S-NI 000080 small subunit ribosomal RNA gene, partial sequence Delfia acidoagenes et eine EGOS 16S cibesomal ENA agene ential sequence
		Uncultured bacterium clone 16shp123-2b06.w2k165 ribosonal RNA gene, partial sequence
		September 2019 Protection (1) 4 leaves Delftia so, BFE41B 16S ribosomal RNA gene, partial sequence
		Delftia sp. B0.09-91 I6S ribosomal RNA gene, complete sequence
		Pacterium KK 19/9/21_152 105 ribosomal KNA gene, partial sequence Delfia sp. PP4 S5 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium partial I6S rRNA gene, clone AcL 606 Uncultured organism clone FLUX024 T172 & NIDCP AMAXIM (XV0363 cmall subwit elbosomel DNA core partial company
		Uncultured organism clone ELU0034-T174-S-NIPCRAMgAND_000116 small subunit ribosomal RivA gene, partial sequence
		Defin acidoagenes et ini WP421 (Se cibacoma JPAA enere nertial sequence
		Uncultured bacterium clone 16slp96-1a04.plt 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone I6slp108-1f06.p1k 16S ribosomal RNA gene, partial sequence
		Bacterium KR 1979/21 26 16S ribosomal RNA gene, partial sequence
		Delftia sp. BFE44L 16S ribosomal RNA gene, partial sequence
		Delfia lacustris strain RK7 16S ribosomal RNA gene, partial sequence Reterring KR 1996/1 37 16S ribosomal RNA gene, partial sequence
		Delfia sp. PP4 54 165 ribosonal RNA gene, partai sequence Delfia sp. PP4 54 165 ribosonal RNA gene, partai sequence
		Delftia sp. PP4_S1 16S ribosomal RNA gene, partial sequence Uncultured organism clone ELU0034-T174-S-NIPCRAMeANb 000335 small subunit ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000231 small subunit ribosomal RNA gene, partial sequence
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		Uncultured bacterium clone 16sip123-1b01.p1k 16S ribosomal RNA gene, partial sequence
		Bacterium SV70AB2-3 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium gene for 16S rRNA, partial sequence, clone: Ms-14-Fx10-1-076 Delfie acideurement strain S. BTA 16S riberemed DNA case acidial accurace
		Ecinia actiovorans strait S-B /A 105 fibosomal RNA gene, partial sequence Bacterium KR 1979/21_108 16S ribosomal RNA gene, partial sequence
		Delftia sp. S17 partial 16S rRNA gene, strain S17 Delftia sp. S17 partial 16S rRNA gene, strain S17 Delftia sp. S17 partial company of the second spectral company of the sec
		Uncultured organism crone ELC0029+11/9-3-NFC-RAMgAND_00050 small subunit ribosomal RNA gene, partial sequence Uncultured organism clone ELC0034-T174-S-NIPCRAMgAND_000507 small subunit ribosomal RNA gene, partial sequence
		Uncultured organism clone ELU0028-T175-S-NIPCRAMgANa 000421 small subunit ribosomal RNA gene, partial sequence Uncultured organism clone ELU0157-T387-S-NIPCRAMgANb 000334 small subunit ribosomal RNA gene, partial sequence
		Delftia acidovorans strain FGQ5 16S ribosomal RNA gene, partial sequence
		Delftia acidovorans strain CL-9.07 16S ribosomal RNA gene, partial sequence Uncultured bacterium clone 16sh97-1a10.p1k 16S ribosomal RNA gene, partial sequence
		Delfia sp. P-101 partial 16S rRNA gene, strain P-101
		Bacterium C0181 105 ribosomal KNA gene, partial sequence

NR_044399.1

Arthrobacter sanguinis strain 741 16S ribosomal RNA gene, partial sequence Sequence ID: <u>ref[NR_044399.1]</u> Length: 1452 Number of Matches: 1 > See 1 more title(s)

Range 1: 419 to 1390 GenBank Graphics Vext Match A Previous Match						
Score 1727 bit	s(1914)	Expect Identities 0.0 967/972(99%	GapsStrand6)1/972(0%)Plus/Minus			
Query	1	CGGGTGTTACCAACTTTCGTG	ACTTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTC	60		
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Delftia acidovorans strain B208 16S ribosomal RNA gene, partial sequence sequence ID: $\underline{gb[KJ781879.1]}$ Length: 1421 Number of Matches: 1

Range 1: 428 to 1376 GenBank Graphics Next Match Previous Match							
Score 1599 bit	s(1772)	Expect 0.0	Identities 927/950(98%)	Gaps 3/950(0%)	Strand Plus/Minus	_	
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Sbjct	1376	GGCGAGACCCGCTCCCATGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTCAC					
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	• k60mm 214175
	 Uncultured organism clone ELU0036-T191-S-NIPCRAMgANb_000088 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000049 small subunit ribosomal RNA gene, partial sequence
	Delfia sp. SM-1 165 ribosonal RNA gene, partial sequence Delfia sp. SM-1 165 ribosonal RNA gene, partial sequence
	Definition action version strain boost tos rebonnant RAA gene, partai sequence Definition Cut-4 DSS ribosomal RAA comelete scouence
	 Uncultured bacterium clone 3H3C_92 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 1H3C_07 16S ribosomal RNA gene, partial sequence
	Delifia sp. Cs1-4, complete genome
	Deterina sp. K-41380 partial 105 rKNA gene, strain K-41380
	Delftia acidovorans strain Aal-4 16S ribosomal RNA gene, partial sequence
	Uncultured Delftia sp. clone G14 16S ribosomal RNA gene, partial sequence
	Delfia sp. Lp-1 16S ribosomal RNA gene, partial sequence
	 Uncultured bacterium clone 16/h0102 + 16/h0102 + 16/s ribosoma KNA gene, partial sequence Discultured bacterium clone 16/h0102 + 16/h0102 + 16/s ribosoma RNA gene, partial sequence
	 Uncultured bacterium clone 16slp108-1006.p1k 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 16slp116-1e05.p1k 16S ribosomal RNA gene, partial sequence
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	Uncultured bacterium clone [6(b)123-1000 wk 105 ribosomal RVA gene partial sequence Uncultured bacterium clone [6(b)123-100) nk 165 ribosomal RVA gene partial sequence
	Uncultured bacterium clone 16slp96-1a04.p1k 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 16slp96-1b03.w2k 16S ribosomal RNA gene, partial sequence
	 Delitia sp. R-41383 partial 165 rRNA gene, strain R-41383 Delitia spi denorme denie (Z. 0.02165 intermed RNA come anticial commons)
	De linia a convoranti strain C.2.7507 to strobomial RCA gene, partat sequence De linia ao. KN19165 Abosomal RCA cene, cartíal sequence
	 Uncultured Delftia sp. clone IODP_305_1309D_water_sample_1200mbsf_216S ribosomal RNA gene, partial sequence
	 Delftia acidovorans strain FGQ8 16S ribosomal RNA gene, partial sequence
	 Defini acidoverans strain FGQS 165 ribosomal RNA gene, partial sequence Defini hearart a train 65 165 elbacomal RNA gene, partial sequence
1.0007	 Alcaligenes faecalis strain 18A 16S ribosonal RNA gene, partial sequence
	 Delfia acidovorans strain WR42 16S ribosomal RNA gene, partial sequence
	 Delftia acidovorans strain NBRC 14950 16S ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL/0008-T38-S-NI 000061 small subunit ribosomal RNA gene, partial sequence
	 Uncumred organism clone EL 0008-158-5-VL 000089 small subunit fibosomia RNA gene, partial sequence Uncumred organism clone EL 10008-158-SNI (00117 cmall subunit fibosomia RNA gene, partial sequence
	 Uncultured organism clone ELU0008-T58-S-NI 000462 small subunit ribosomal RNA cene, partial sequence
	Uncultured organism clone ELU0024-T375-S-NIPCRAMgANb_000253 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone ELU0157-T387-S-NIPCRAMgANb_000129 small subunit ribosomal RNA gene, partial sequence
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	 Uncutated organism clone EL UD18-1 300-S-NIP, KAMgANa (000424 small subunit ribosomal KNA gene, partial sequence Uncutated organism clone EL UD18-NIP (PA Ma AN) (00110 small adunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone ELU0028-T175-SNIP/CRAMgANa (00421 small subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000286 small subunit ribosomal RNA gene, partial sequence
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	 Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb 000300 small subunit ribosomal RNA gene, partial sequence
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	Uncultured organism (close EL10014-T174-SNIPCRAMeANb 000415 small subusit rebosonal PCA gete, partial sequence
	Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb 000507 small subunit ribosomal RNA gene, partial sequence
	Uncultured organism clone ELU0034-T174-S-NIPCRAMgANb_000554 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EL100014-T174-S-NIPCRAMgANb_000557 small subunit ribosomal RNA gene, partial sequence
	 Uncultured organism clone EU003+1174-S-NIPC KAMgANb (00064) small subunit ribosonial KNA gene, partial sequence Uncultured organism clone EU10014-1174-S-NIPC PAMgANb (000657 nm) down of photomal RNA gene, partial sequence
	Cucultured beta moteorie i concerne a cheficichia a LL67 2. IB 104 small subunit ribosomal RNA eren partia sequence
	Uncultured bacterium partial 16S rRNA gene, clone AcL_G06
	 Delfia sp. PP4 S1 165 ribosomal RNA gene, partial sequence
	 Deffaa sp. PP4 52 105 ribosomal RNA gene, partial sequence Deffaa sp. PD4 52 105 ribosomal RNA gene, partial sequence
	 Lettina 50, 174 53 105 fiboromial RNA gene, partai sequence Pablica p. DD. C 154 fiboromial RNA gene, partai sequence
	Debia ap. P 74_09 105 flowsmal RNA cene, partal sequence Debia ap. P 14 55 165 flowsmal RNA cene, partal sequence
	 Delfia sp. PP4_56 16S ribosomal RNA gene, partial sequence
	 Delfia sp. PP4_S7 16S ribosomal RNA gene, partial sequence
	 Defina sp. 517 partial 165 rKNA gene, strain 517 Backnown VP, DOOR 111 165 rKNA gene, nami partial accuracy
	Data Finan KR 1979/_33 103 Hobomata RAA gene, partai sequence deaterium KR 1996/1 85 165 Hobomata RAA gene, partai sequence
	Bacterium KR 1979/2T_108 16S ribosonul RNA gene, partial sequence
	 Bacterium KR 199671 115 16S ribosomal RNA gene, partial sequence
	 Bacterium KR 19901 110 105 rhosomal RNA gene, partial sequence Bacterium KR 19901 151 165 rhosomal RNA gene, partial sequence
	 Bacterium KR 1996/1 37 165 ribosomal RNA gene, partial sequence
	Bacterium KR 1996/1_185 16S ribosomal RNA gene, partial sequence
	 Delfais sp. 8-10 165 ribosomal RNA gene, partial sequence Delfais burnetic during RVA (Escherand RNA) and anticid assume
	Evina accustrs strain Rev / tos recomma Reve gene, partial sequence Ordefita so, BLOOPA1165 ribosomal RNA eene, connelete sequence
	Delfila sp. hg-1 16S ribosomal RNA gene, partial sequence
	 Delfila sp. SR27 16S ribosomal RNA gene, partial sequence
	 Delfita acidovorans strain S-B7A 165 ribosomal RNA gene, partial sequence Delfica acidovorans strain A 2010 165 ribosomal RNA gene, partial sequence
	 Definitia acculorotario strain A-10-150 100 1000 0000 FCA gene, partial sequence Definiti suandateniois strain X15 16S ribosomal FCA gene, partial sequence
	 Uncultured bacterium gene for 16S rRNA, partial sequence, clone: Ms-14-Fx10-1-076
	Uncultared bacterium gene for 16S rRNA, partial sequence, clone: Ms-14-Fx10-2-060
	 Uncultured bacterium gene for 165 rRNA, partial sequence, clone: Mr-14-Fx11-1-075 Uncultured bacterium gene for 165 rRNA (second bacteria)
	Uncultured backening gene for 165 rR5AN, partial sequence, (cone: Bys1405-5810-013 Backening CO183 1165 observational RNA cone, navial sequence
	4 b-protobacteria 16 kaves

Bacillus cereus GQ344804.1

Bacillus cereus strain DC2 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|GQ344804.1|</u> Length: 1424 Number of Matches: 1

Range 1: 442 to 1382 GenBank Graphics Vext Match 🔺 Previous Match							
Score 1397 bits	s(1548)	Expect 0.0	Identities 883/941(94%)	Gaps 11/941(1%)	Strand Plus/Minus	_	
Query	1	CAAACTCTCG	IGGTGTGACGGGC	GGTGTGTACAAGG	CCCGGGAACGTATTC	ACCGCGGCA	60
Sbjct	1382		IGGTGTGACGGGCC	GTGTGTACAAGG	CCCGGGAACGTATTC	ACCGCGGCA	1323
Query	61	TGCTGATCCG	CGATTACTAGCGAT	TTCCAACTTCAT	TAGGCGAGTTGCAGC	CGACAATCC	120
Sbjct	1322	TGCTGATCCG	CGATTACTAGCGAT	TCCAGCTTCATG	TAGGCGAGTTGCAGC	CTACAATCC	1263
Query	121	GAACTGAGAA	CGGTTTTATGAGA	TAGCTCCACCTC	GCGGTCTTGCAGCTC	TTTGTACCG	180
Sbjct	1262	GAACTGAGAA	CGGTTTTATGAGA	TAGCTCCACCTC	CGCGGTCTTGCAGCTC	TTTGTACCG	1203
Query	181	TCCATTGTAT	CACGTGTGTAGCCC	CAGGTCATAAGGG	GCATGATGATTTGAC	GTCATCCCC	240
Sbjct	1202	TCCATTGTAG	CACGTGTGTAGCCO	CAGGTCATAAGGG	GCATGATGATTTGAC	GTCATCCCC	1143
Query	241	ACCTTCCTCC	GGTTTGTCACCGGC	CAGTCACCTTAGA	GTGCCCAACTTAATG	ATGGCAACT	300
Sbjct	1142	ACCTTCCTCC	GTTTGTCACCGGC	CAGTCACCTTAGA	AGTGCCCAACTTAATG	ATGGCAACT	1083
Query	301	AGAATCAAGGO	GTTGCGCTCGTTGC	CGGGACTTAACCO	CAACATCTCACGACAC	GATCTGAAC	360
Sbjct	1082	AAGATCAAGGO	GTTGCGCTCGTTGC	CGGGACTTAACCO	CAACATCTCACGACAC	GAGCTGACG	1023
Query	361	ACAACCATGC	ACCACCTGTCACTO	CTGCTCCCGAAGG	ACAAACCCTATCTCT.	AGGGTTTTC	420
Sbjct	1022	ACAACCATGC	ACCACCTGTCACTO	CTGCTCCCGAAGG	GAGAAGCCCTATCTCT.	AGGGTTTTC	963
Query	421	A-AGGATGTC	AAGACCTGGTAAGO	GTTCTTCGCGTTG	CTTCGAATTAAACCA	CATGCTCCA	479
Sbjct	962	AGAGGATGTC	AAGACCTGGTAAGO	GTTCTTCGCGTTG	GCTTCGAATTAAACCA	CATGCTCCA	903
Query	480	CCGCTTGTGC	GGCCCCCGTCAA	TCCTTTGAGTTT	CAGCCTTGCGGCCGT	ACTCCCCAG	<mark>539</mark>
Sbjct	902	CCGCTTGTGC	GGCCCCCGTCAA	TCCTTTGAGTTT	CAGCCTTGCGGCCGT	ACTCCCCAG	843
Query	540	GCGGAGTGCT	TAATGCGTTAACT	ICAGCACTAAAGG	GCGGAAACCCTCTAA	CACTTATCA	599
Sbjct	842	GCGGAGTGCT	TAATGCGTTAACT	FCAGCACTAAAGG	GCGGAAACCCTCTAA	CACTTAGCA	783
Query	600	CTCATCGTTT	ACGGCGTGGACTAC	CCGGGATATCTAA	ATCGTGTTTGCTCCCC.	ACTCTTTCG	659
Sbjct	782	CTCATCGTTT	ACGGCGTGGACTA	CCAGGGTATCTA	ATCCTGTTTGCTCCCC.	ACGCTTTCG	723
Query	660	CGCCTCAGTG	ICAATTACGGACCA	A-AAAGTCCCCTI	CCCCACTGGTGTTCC	TCCATATCT	718
Sbjct	722	CGCCTCAGTG	ICAGTTACAGACC	AGAAAGTCGCCTI	CGCCACTGGTGTTCC	TCCATATCT	663
Query	719	CTACCCTTTT	CACCGCTACACAT	GAAATT-CACTTI	CCTCTTCTGCACTCA	AGTCTCCCA	777
Sbjct	662	CTACGCATTT	CACCGCTACACAT	GAATTCCACTTI	CCTCTTCTGCACTCA	AGTCTCCCA	603
Query	778	GTTTCCAATG	ACCC-CCACGGTT	SACCCG-GGGTTI	TCACTTCAAACTTAA	-AAACC-CC	833
Sbjct	602	GTTTCCAATG	ACCCTCCACGGTT	SAGCCGTGGGCTI	TCACATCAGACTTAA	GAAACCACC	543
Query	834	GGCCCGCCCT	TTAC-CCCAAT-AT		TTGCCCCCTACTTAT	TACC-Cggg	890
Sbjct	542	TGCGCGCGCGCT	TTACGCCCAATAA	TTCCGGATAACGC	TTGCCACCTACGTAT	TACCGCGGC	483
Query	891	gggggggCCCG2			GAACC 930		
Sbjct	482	TGCTGGCACG	TAGTTAGCCGTGG	CTTTTCTGGTTAG	GTACC 442		



10- The phylogenetic analysis of bacteria isolated from used toothbrushes

samples

Roseomonas mucosa KF247232.1

Roseomonas mucosa strain SM14032013 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KF247232.1</u>| Length: 1455 Number of Matches: 1

Range 1: 102 to 983 GenBank Graphics Vext Match A Previous Match						
Score 1572 bits	s(1742)	Expect 0.0	Identities 879/883(99%)	Gaps 1/883(0%)	Strand Plus/Plus	
Query	1	GGACGGGTGAGTA	ACGCGTAGGAACGT	GTCCTGAGGTGGG	GGACAACCCCGGGAAACTGG	60
Sbjct	102	GGACGGGTGAGTA	ACGCGTAGGAACGT	GTCCTGAGGTGGG	GGACAACCCCGGGAAACTGG	161
Query	61	GGCTAATACCGCA	TATGGGCTGAGGCC	CAAAGCCGAGAGG	CGCCTTTGGAGCGGCCTGCG	120
Sbjct	162	GGCTAATACCGCA	TATGGGCTGAGGCC	CAAAGCCGAGAGG	CGCCTTTGGAGCGGCCTGCG	221
Query	121	TCCGATTAGGTAG	TTGGTGGGGTAAAG	GCCTACCAAGCCT	GCGATCGGTAGCTGGTCTGA	180
Sbjct	222	TCCGATTAGGTAG	TTGGTGGGGTAAAG	GCCTACCAAGCCT	GCGATCGGTAGCTGGTCTGA	281
Query	181	GAGGACGACCAGC	CACACTGGGACTGA	GACACGGCCCAGA	CTCCTACGGGAGGCAGCAGT	240
Sbjct	282	GAGGACGACCAGC	CACACTGGGACTGA	GACACGGCCCAGA	CTCCTACGGGAGGCAGCAGT	341
Query	241	GGGGAATATTGGA	CAATGGGCGAAAGC	CTGATCCAGCAAT	GCCGCGTGGGTGAAGAAGGT	300
Sbjct	342	GGGGAATATTGGA	CAATGGGCGAAAGC	CTGATCCAGCAAT	GCCGCGTGGGTGAAGAAGGT	401
Query	301	CTTCGGATCGTAA	AGCCCTTTCGACGG	GGACGATGATGAC	GGTACCCGTAGAAGAAGCCC	360
Sbjct	402	CTTCGGATCGTAA	AGCCCTTTCGACGG	GGACGATGATGAC	GGTACCCGTAGAAGAAGCCC	461
Query	361	CGGCTAACTTCGT	GCCAGCAGCCGCGG	TAATACGAAGGGG	GCTAGCGTTGCTCGGAATTA	420
Sbjct	462	CGGCTAACTTCGT	GCCAGCAGCCGCGG	TAATACGAAGGGG	GCTAGCGTTGCTCGGAATTA	521
Query	421	CTGGGCGTAAAGG	GCGCGTAGGCGGCG	GCCCAAGTCAGGC	GTGAAATTCCTGGGCTCAAC	480
Sbjct	522	CTGGGCGTAAAGG	GCGCGTAGGCGGCG	GCCCAAGTCAGGC	GTGAAATTCCTGGGCTCAAC	581
Query	481	CTGGGGACTGCGC	TTGATACTGGGTTG	CTTGAGGATGGAA	GAGGCTCGTGGAATTCCCAG	540
Sbjct	582	CTGGGGACTGCGC	TTGATACTGGGTTG	CTTGAGGATGGAA	GAGGCTCGTGGAATTCCCAG	641
Query	541	TGTAGAGGTGAAA	TTCGTAGATATTGG	GAAGAACACCGGT	GGCGAAGGCGGCGAGCTGGT	600
Sbjct	642	TGTAGAGGTGAAA	TTCGTAGATATTGG	GAAGAACACCGGT	GGCGAAGGCGGCGAGCTGGT	701
Query	601	CCATTACTGACGC	TGAGGCGCGACAGC	GTGGGGGAGCAAAC	AGGATTAGATACCCTGGTAG 	660
Sbjct	702	CCATTACTGACGC	TGAGGCGCGACAGC	GTGGGGGGGGCAAAC	AGGATTAGATACCCTGGTAG	761
Query	661	TCCACGCCGTAAA	CGATGTGCGCTGGA	TGTTGGGGGCCCAT	AGGGTCTCAGTGTCGTAGCC	720
Sbjct	762	TCCACGCCGTAAA	CGATGTGCGCTGGA	TGTTGGGGGCCCAT	AGGGTCTCAGTGTCGTAGCC	821
Query	721	AACGCGGTAAGCG	CACCGCCTGGGGAG	TACGGCCGCAAGG	TTGAAACTCAAAGGAATTGA	780
Sbjct	822	AACGCGGTAAGCG	CACCGCCTGGGGAG	TACGGCCGCAAGG	TTGAAACTCAAAGGAATTGA	881
Query	781	CGGGGGGCCCGCAC	AAGCGGTGGAGCAT	GTGGTTTAATTCG	AAGCAACGCGCAAAACCCTT	840
Sbjct	882	CGGGGGGCCCGCAC	AAGCGGTGGAGCAT	GTGGTTTAATTCG	AAGCAACGCGCAGAA-CCTT	940
Query	841	ACCAGCCCTTGAC	ATGGTCACGACCGG	TCCAAAAATGGAC	TTT 883	
Sbjct	941	ACCAGCCCTTGAC	ATGGTCACGACCGG	TCCAGAGATGGAC	TTT 983	

lelQuery_10625

uery_10625	
	Uncultured bacterium clone ncd2760c02c1 16S ribosomal RNA gene, partial sequence
	Roseomonas mucosa strain MDA5527 16S ribosomal RNA gene, partial sequence
	Candidatus Roseomonas massiliae 1461A 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncd752e03c1 16S ribosomal RNA gene, partial sequence
	hacteria 2 leaves
	Chateria 5 leaves
	bacteria 2 leaves
	Uncultured bacterium clone ncd648f11c1 16S ribosomal RNA gene, partial sequence
	Roseomonas sp. CB 286483 partial 16S rRNA gene, strain CB 286483
	Phateria 2 leaves
	Roseomonas sp. CMS4Y-2-2 16S ribosomal RNA gene, partial sequence
	stracteria 2 leaves
	Stateria 5 leaves
	Roseomonas mucosa strain EHFS2_S05Hb 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone ncd507e12c1 16S ribosomal RNA gene, partial sequence
	Anderia 5 leaves
	thateria 2 leaves
	Candidatus Roseomonas massiliae 16S ribosomal RNA gene, partial sequence
	obacteria 2 leaves
	Abadenia 5 leaves
	Roseomonas terpenica partial 16S rRNA gene, type strain DT
	Oncultured bacterium clone ncd510x02c1 16S ribosomal RNA gene, partial sequence
	bacteria 5 leaves
	bacteru 2 leaves
	Koseomonas mucosa strain SM14032013 16S ribosomal KNA gene, partial sequence
	onacteru 2 leaves
	There is a series
	Koreomonas mucosa strain ES_21con 165 ribosomal RNA gene, partial sequence
	Uncultured bacterium cione ncd515a06c1 165 nbosomat KiNA gene, partial sequence
	Process of P7414C charged PVA and and a static common
	Koscononias sp. 0244 105 noosoniai KivA gene, pariai sequence
	Tracera 2 leaves
	TAKETA JA KANS

0.0003

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302

Stenotrophomonas maltophilia LN890169.1

Stenotrophomonas maltophilia partial 16S rRNA gene, strain M83 Sequence ID: <u>emb|LN890169.1</u>] Length: 1688 Number of Matches: 1

Range 1: 190 to 1127 GenBank Graphics Vext Match Previous Match							
Score 1665 bit	s(1846)	Expect I 0.0 S	dentities 933/938(99%)	Gaps 1/938(0%)	Strand Plus/Plu	S	
Query	1	TGGGTGGCGAGTG	GCGGACGGGTGA	AGGAATACATCGO	GAATCTACTCT	GTCGTGGGGGATA	60
Sbjct	190	TGGGTGGCGAGTG	GCGGACGGGTG	AGGAATACATCG	GAATCTACTCT	 GTCGTGGGGGATA	249
Query	61	ACGTAGGGAAACT	TACGCTAATACO	GCATACGACCT	ACGGGTGAAAG	CAGGGGATCTTCG	120
Sbjct	250	ACGTAGGGAAACT	TACGCTAATACO	GCATACGACCT	ACGGGTGAAAG	CAGGGGGATCTTCG	309
Query	121	GACCTTGCGCGAT	TGAATGAGCCGA	ATGTCGGATTAG	CTAGTTGGCGG	GGTAAAGGCCCAC	180
Sbjct	310	GACCTTGCGCGAT	IGAATGAGCCGA	ATGTCGGATTAG	CTAGTTGGCGG	GGTAAAGGCCCAC	369
Query	181	CAAGGCGACGATC	CGTAGCTGGTC	GAGAGGATGAT	CAGCCACACTG	GAACTGAGACACG	240
Sbjct	370	CAAGGCGACGATC	CGTAGCTGGTCI	GAGAGGATGAT	CAGCCACACTG	GAACTGAGACACG	429
Query	241	GTCCAGACTCCTA		AGTGGGGGAATATT	IGGACAATGGG	CGCAAGCCTGATC	300
Sbjct	430	GTCCAGACTCCTA	CGGGAGGCAGCA	AGTGGGGGAATATT	IGGACAATGGG	CGCAAGCCTGATC	489
Query	301	CAGCCATACCGCG	TGGGTGAAGAAG	GCCTTCGGGTT	GTAAAGCCCTT	TTGTTGGGAAAGA	360
Sbjct	490	CAGCCATACCGCG	TGGGTGAAGAAG	GCCTTCGGGTT	STAAAGCCCTT	TTGTTGGGAAAGA	549
Query	361	AATCCAGCTGGCT	AATACCCGGTTG	GGATGACGGTA	CCAAAGAATA	AGCACCGGCTAAC	420
Sbjct	550	AATCCAGCTGGCT	AATACCCGGTTC	GGATGACGGTA	CCAAAGAATA	AGCACCGGCTAAC	609
Query	421	TTCGTGCCAGCAG	CCGCGGTAATAC	GAAGGGTGCAAG	GCGTTACTCGG	AATTACTGGGCGT	480
Sbjct	610	TTCGTGCCAGCAG	CCGCGGTAATAC	GAAGGGTGCAAG	GCGTTACTCGG	AATTACTGGGCGT	669
Query	481	AAAGCGTGCGTAG	GTGGTCGTTTA	AGTCCGTTGTGAZ	AGCCCTGGGC	TCAACCTGGGAAC	540
Sbjct	670	AAAGCGTGCGTAG	GTGGTCGTTTA	AGTCCGTTGTGAZ	AGCCCTGGGC	TCAACCTGGGAAC	729
Query	541	TGCAGTGGATACT	GGGCGACTAGAG	TGTGGTAGAGG	GTAGCGGAATT	CCTGGTGTAGCAG	600
Sbjct	730	TGCAGTGGATACT	GGGCGACTAGAG	TGTGGTAGAGG	STAGCGGAATT	CCTGGTGTAGCAG	789
Query	601	TGAAATGCGTAGA	GATCAGGAGGA	ACATCCATGGCGA	AAGGCAGCTAC	CTGGACCAACACT	660
Sbjct	790	TGAAATGCGTAGA	GATCAGGAGGA	ACATCCATGGCG	AAGGCAGCTAC	CTGGACCAACACT	849
Query	661	GACACTGAGGCAC	GAAAGCGTGGGG	GAGCAAACAGGA	TTAGATACCCT	GGTAGTCCACGCC	720
Sbjct	850	GACACTGAGGCAC	GAAAGCGTGGGG	GAGCAAACAGGA	TTAGATACCCT	GGTAGTCCACGCC	909
Query	721	CTAAACGATGCGA	ACTGGATGTTG	GTGCAATTTGG	CACGCAGTATC	GAAGCTAACGCGT	780
Sbjct	910	CTAAACGATGCGA	ACTGGATGTTG	GTGCAATTTGG	CACGCAGTATC	GAAGCTAACGCGT	969
Query	781	TAAGTTCGCCGCC	TGGGGAGTACG	GTCGCAAGACTG	AAACTC-AAGG	AATTGACGGGGGC	839
Sbjct	970	TAAGTTCGCCGCC	TGGGGAGTACG	GTCGCAAGACTG	AAACTCAAAGG	AATTGACGGGGGC	1029
Query	840	CCGCACAAGCGGT	GGAGTATGTGG			ACCTTACCTGGCC	899
Sbjct	1030	CCGCACAAGCGGT	GGAGTATGTGG	TTTAATTCGATG	CAACGCGAAGA	ACCTTACCTGGCC	1089
Query	900	TTGACATGTCGAG			CC 937		
Sbjct	1090	TTGACATGTCGAG	AACTTTCCAGA	GATGGATGGGTG	CC 1127		

Pseudomonas aeruginosa strain Pse12 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|KR815846.1|</u> Length: 1444 Number of Matches: 1

Range 1:	34 to 99	9 GenBank Graphics		•	Next Match 🔺 Previous Match	
Score 1685 bit	s(1868)	Expect 0.0	Identities 955/966(99%)	Gaps 5/966(0%)	Strand Plus/Plus	
Query	1	TGGATTCNGCGG	CGGACGGGTGAGT	ATGCCTAGGAAT	CTGCCTGGTAGTGGGGGGATAAC	c 60
Sbjct	34	TGGATTCAGCGG	CGGACGGGTGAGT	ATGCCTAGGAAT	CTGCCTGGTAGTGGGGGGATAAC	93
Query	61	GTCCGGAAACGG	GCGCTAATACCGCZ	ATACGTCCTGAGG	GAGAAAGTGGGGGGATCTTCGGA	A 120
Sbjct	94	GTCCGGAAACGG	GCGCTAATACCGC	ATACGTCCTGAGG	GAGAAAGTGGGGGGATCTTCGGA	A 153
Query	121	CCTCACGCTATC	AGATGAGCCTAGG	CGGATTAGCTAG	TTGGTGGGGTAAAGGCCTACC	A 180
Sbjct	154	CCTCACGCTATC	AGATGAGCCTAGG	CGGATTAGCTAG	TTGGTGGGGTAAAGGCCTACCA	A 213
Query	181	AGGCGACGATCC	GTAACTGGTCTGA0	AGGATGATCAGT	CACACTGGAACTGAGACACGGI	240
Sbjct	214	AGGCGACGATCC	GTAACTGGTCTGAC	GAGGATGATCAGT	CACACTGGAACTGAGACACGGI	273
Query	241	CCAGACTCCTAC	GGGAGGCAGCAGT	GGGAATATTGGA	CAATGGGCGAAAGCCTGATCC#	A 300
Sbjct	274	CCAGACTCCTAC	GGGAGGCAGCAGT	GGGAATATTGGA	CAATGGGCGAAAGCCTGATCCA	333
Query	301	GCCATGCCGCGT	GTGTGAAGAAGGTC	CTTCGGATTGTAA	AGCACTTTAAGTTGGGAGGAAG	5 360
Sbjct	334	GCCATGCCGCGT	GTGTGAAGAAGGTC	CTTCGGATTGTAA	AGCACTTTAAGTTGGGAGGAAG	393
Query	361	GGCAGTAAGTTA	ATACCTTGCTGTT1	TGACGTTACCAA	CAGAATAAGCACCGGCTAACTI	420
Sbjct	394	GGCAGTAAGTTA	ATACCTTGCTGTT	TGACGTTACCAA	CAGAATAAGCACCGGCTAACTI	453
Query	421	CGTGCCAGCAGC	CGCGGTAATACGAZ	AGGGTGCAAGCGT	TAATCGGAATTACTGGGCGTA	A 480
Sbjct	454	CGTGCCAGCAGC	CGCGGTAATACGAA	AGGGTGCAAGCGT	TAATCGGAATTACTGGGCGTA	A 513
Query	481	AGCGCGCGTAGG	TGGTTCAGCAAGT	GGATGTGAAATC	CCCGGGCTCAACCTGGGAACTC	3 540
Sbjct	514	AGCGCGCGTAGG	TGGTTCAGCAAGT	GGATGTGAAATC	CCCGGGCTCAACCTGGGAACTC	573
Query	541	CATCCAAAACTA	CTGAGCTAGAGTAG	CGGTAGAGGGTGG	TGGAATTTCCTGTGTAGCGGTG	5 <mark>600</mark>
Sbjct	574	CATCCAAAACTA	CTGAGCTAGAGTAG	GGTAGAGGGTGG	TGGAATTTCCTGTGTAGCGGT	633
Query	601	AAATGCGTAGAT.	ATAGGAAGGAACAC		CGACCACCTGGACTGATACTGA	. 660
Sbjct	634	AAATGCGTAGAT.	ATAGGAAGGAACAC	CAGTGGCGAAGG	CGACCACCTGGACTGATACTGA	693
Query	661	CACTGAGGTGCG	AAAGCGTGGGGAGC	:AAACAGGATTAG 	ATACCCTGGTAGTCCACGCCGI	720
Sbjct	694	CACTGAGGTGCG	AAAGCGTGGGGAGC	CAAACAGGATTAG	ATACCCTGGTAGTCCACGCCGI	753
Query	721	AAACGATGTCGA	CTAGCCGTTGGGA1 	CCTTGAGATCTT	AGTGGCGCAGCTAACGCGATAA	1 780
Sbjct	754	AAACGATGTCGA	CTAGCCGTTGGGAI	CCTTGAGATCTT	AGTGGCGCAGCTAACGCGATAA	813
Query	781	GTCGACCGCCTG	GGGAGTACGGCCGC	CAAGGTTAAAACT(CAAATGAATTGACGGGGGCCCCG	; 840
Sbjct	814	GTCGACCGCCTG	GGGAGTACGGCCGC	AAGGTTAAAACT	CAAATGAATTGACGGGGGGCCCG	; 873
Query	841	CACAAGCGGTGG	AGCATGTGGTTTA#	ATTCTAAGCAACG	CGAAAAACCTTACCTGGGCCTT	900
Sbjct	874	CACAAGCGGTGG	AGCATGTGGTTTAA	TTCGAAGCAACG	CGAAAAACCTTACCTGGGCCTT	933
Query	901	GACATGCTGAAA	ACTTTCCAAAAATG	GATTGGGGGCCTT	CGGAACTC-AAACCCAGG 	955
Sbjct	934	GACATGCTGAGA	ACTTTCCAAAGATG	GATTGGTGCCTT	CCGGGAACTCAAAACACAGGGI	993
Query	956	GCTGCA 961				
Sbjct	994	GCTGCA 999				


Leclercia adecarboxylata strain MCCB 331 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KT899848.1</u>| Length: 1323 Number of Matches: 1

Range 1:	1 to 905	5 <u>GenBank</u>	Graphics				🔻 Next M	1atch 🔺 P	revious Mate	ch	
Score 1597 bit	s(1770)		Expect 0.0	Identities 897/905(99%)		Gaps L/905(0%))	Strand Plus/Plu	5		
Query	6	GACGA	GTGGCGGA	CGGGTGAGTAAT	GTCTG	GAAACT	GCCTG	ATGGAG	GGGGATA	ACTAC	65
Sbjct	1	GACGA	GTGGCGGA	CGGGTGAGTAAT.	GTCTG	GAAACT	GCCTG	ATGGAG	GGGGATA	ACTAC	60
Query	66	TGGAA	ACGGTAGC	TAATACCGCATA	AYGTC	GCAAGAC	CAAAG	AGGGGG	ACCTTCO	GGCCT	125
Sbjct	61	TGGAA	ACGGTAGC	TAATACCGCATA	ATGTC	GCAAGAC	CAAAG	AGGGGG	ACCTTCO	GGCCT	120
Query	126	CTTGC	CATCGGAT	GTGCCCAGATGG	GATTA	SCTAGTA	GGTGG	GGTAAT	GGCTCAC	CTAGG	185
Sbjct	121	CTTGC	CATCGGAT	GTGCCCAGATGG	GATTA	GCTAGTA	GGTGG	GGTAAT	GGCTCAC	CTAGG	180
Query	186	CGACG	ATCCCTAG	CTGGTCTGAGAG	GATGA	CAGCCA	CACTG	GAACTG	AGACACO	GTCCA	245
Sbjct	181	CGACG	ATCCCTAG	CTGGTCTGAGAG	GATGA	CAGCCA	CACTG	GAACTG	AGACACG	GTCCA	240
Query	246	GACTC	CTACGGGA	GGCAGCAGTGGG	GAATA	TGCACA	ATGGG	CGCAAG	CCTGATO	CAGCC	305
Sbjct	241	GACTC	CTACGGGA	.GGCAGCAGTGGG	GAATA	TGCACA	ATGGG	CGCAAG	CCTGATO	GCAGCC	300
Query	306	ATGCC	GCGTGTAT	GAAGAAGGCCTT	CGGGT	GTAAAG	TACTT	TCAGCG	AGGAGGA	AGGCA	365
Sbjct	301	ATGCC	GCGTGTAT	GAAGAAGGCCTT	CGGGT	GTAAAG	TACTT	IIIII ICAGCG	AGGAGGA	AGGCG	360
Query	366	TTGTG	GTTAATAA	CCGCAGTGATTG	ACGTT	ACTCGCA	GAAGA	AGCACC	GGCTAAC	TCCGT	425
Sbjct	361	TTGTG	GTTAATAA	CCACAGTGATTO	ACGTT	ACTCGCA	GAAGA	AGCACC	GGCTAAC	TCCGT	420
Query	426	GCCAG		GTAATACGGAGG	GTGCA	GCGTTA	ATCGG	AATTAC	TGGGCGI	AAAGC	485
Sbjct	421	GCCAG	CAGCCGCG	GTAATACGGAGG	GTGCA	AGCGTTA	ATCGG	AATTAC	TGGGCGI	AAAGC	480
Query	486	GCACG	CAGGCGGT	CTGTTAAGTCAG	ATGTG	AATCCC	CGGGC	TCAACC	TGGGAAC	TGCAT	545
Sbjct	481	GCACG	CAGGCGGT	CTGTTAAGTCAG	ATGTG	AATCCC	CGGGC	TCAACC	TGGGAAC	TGCAT	540
Query	546	TTGAA	ACTGGCAG	GCTTGAGTCTTG	TAGAG	GGGGGTA	GAATT	CCAGGT	GTAGCGG	TGAAA	605
Sbjct	541	TTGAA	ACTGGCAG	GCTTGAGTCTTG	TAGAG	GGGGGTA	GAATT	CCAGGT	GTAGCGG	TGAAA	600
Query	606	TGCGT	AGAGATCT	GGAGGAATACCG	GTGGC	GAAGGCG	GCCCC	CTGGAC	AAAGACI	FGACGC	665
Sbjct	601	TGCGT	AGAGATCT	GGAGGAATACCG	GTGGC	SAAGGCG	GCCCC	CTGGAC	AAAGACI	GACGC	660
Query	666	TCAGG	IGCGAAAG	CGTGGGGGAGCAA		ATTAGAT.	ACCCT	GGTAGT	CCACGCO	GTAAA	725
Sbjct	661	TCAGG:	IGCGAAAG	CGTGGGGGAGCAA	ACAGG	ATTAGAT.	ACCCT	GGTAGT	CCACGCO	GTAAA	720
Query	726	CGATG	TCGACTTG	GAGGTTGTGCCC	TTGAG	GCGTGGC	TTCCG	GAGCTA	ACGCGTI	TAAGTC	785
Sbjct	721	CGATG	ICGACTTG	GAGGTTGTGCCC	TTGAG	CGTGGC	TTCCG	GAGCTA	ACGCGTI	TAAGTC	780
Query	786	GACCG	CCTGGGGA	GTACGGCCGCAA	GGTTA	AACTCA	AATGA	ATTGAC	GGGGGCC	CCG-CA	844
Sbjct	781	GACCG	CCTGGGGA	GTACGGCCGCAA	GGTTA	AACTCA	AATGA	ATTGAC	GGGGGCC	CGCCA	840
Query	845	CAAGCO	GGGGGAGC	ATGTGGTTTAAT	TCAAT	GCAACGC	GAAAA	ACCTTA	CCTACTO	CTTGAC	904
Sbjct	841	CAAGC	GGTGGAGC	ATGTGGTTTAAT	TCGAT	GCAACGC	GAAGA	ACCTTA	CCTRCTO	CTTGAC	900
Query	905	ATCCA	909								
Sbjct	901	ATCCA	905								



Enterobacter asburiae EU239468.1

Enterobacter asburiae strain XJUHX-5 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|EU239468.1</u>] Length: 1446 Number of Matches: 1

Range 1:	504 to 13	94 GenBank Graphics	1	V N	ext Match 🔺 Previous M	latch	
Score 1573 bits	s(1744)	Expect 0.0	Identities 884/891(99%)	Gaps 1/891(0%)	Strand Plus/Minus		
Query	1	TTTGCAACCCAC	TCCCATGGTGTGAC	GGCGGTGTGTA		GTATTCAC	60
Sbjct	1394	TTTGCAACCCAC	TCCCATGGTGTGAC	GGCGGTGTGTAC	AAGGCCCGGGAAC	GTATTCAC	1335
Query	61		ATCTACGATTACTAC	CGATTCCGACT	CATGGAGTCGAGT	TGCAGACT	120
Sbjct	1334	CGTAGCATTCTG	ATCTACGATTACTAC	GCGATTCCGACTI	CATGGAGTCGAGT	TGCAGACT	1275
Query	121	CCAATCCGGACT	ACGACGCACTTTATC	BAGGTCCGCTTGC	TCTCGCGAGGTCG	CTTCTCTT	180
Sbjct	1274	CCAATCCGGACT	ACGACGCACTTTATO	SAGGTCCGCTTGC	TCTCGCGAGGTCG	CTTCTCTT	1215
Query	181	TGTATGCGCCAT	TGTAGCACGTGTGTZ	AGCCCTACTCGT		CTTGACGT	240
Sbjct	1214	TGTATGCGCCAT	TGTAGCACGTGTGT	AGCCCTACTCGT	AGGGCCATGATGA	CTTGACGT	1155
Query	241	CATCCCCACCTT	CCTCCAGTTTATCAC	CTGGCAGTCTCCI	TTGAGTTCCCGGC	CTAACCGC	300
Sbjct	1154	CATCCCCACCTT	CCTCCAGTTTATCAC	CTGGCAGTCTCCI	TTGAGTTCCCGGC	CTAACCGC	1095
Query	301	TGGCAACAAAGG	ATAAGGGTTGCGCTC	CGTTGCGGGACTI	AACCCAACATTTC	ACAACACG 	360
Sbjct	1094	TGGCAACAAAGG	ATAAGGGTTGCGCTC	CGTTGCGGGGACTI	AACCCAACATTTC	ACAACACG	1035
Query	361	AGCTGACGACAG	CCATGCAGCACCTG	CTCAGAGTTCCC	GAAGGCACCAAGC	CATCTCTG	420
Sbjct	1034	AGCTGACGACAG	CCATGCAGCACCTG	CTCAGAGTTCCC	GAAGGCACCAATC	CATCTCTG	975
Query	421	GCAAGTTCTCTG	GATGTCAAGAGTAGO	TAAGGTTCTTCG	CGTTGCATCGAAT	TAAACCAC	480
Sbjct	974	GAAAGTTCTCTG	GATGTCAAGAGTAG	STAAGGTTCTTCC	CGTTGCATCGAAT	TAAACCAC	915
Query	481	ATGCTCCACCGC	TTGTGCGGGCCCCCG	STCAATTCATTTC	AGTTTTAACCTTG	CGGCCGTA 	540
Sbjct	914	ATGCTCCACCGC	TTGTGCGGGCCCCCC	STCAATTCATTTC	AGTTTTAACCTTG	CGGCCGTA	855
Query	541	CTCCCCAGGCGG	TCGACTTAACGCGT1 	AGCTCCGGAAGO	CACGCCTCAAGGG	CACAACCT	600
Sbjct	854	CTCCCCAGGCGG	TCGACTTAACGCGTI	PAGCTCCGGAAGC	CACGCCTCAAGGG	CACAACCT	795
Query	601	CCAAGTCGACAT	CGTTTACGGCGTGGA	CTACCAGGGTAT	CTAATCCTGTTTG	CTCCCCAC	660
Sbjct	794	CCAAGTCGACAT	CGTTTACGGCGTGGA	CTACCAGGGTAI	CTAATCCTGTTTG	CTCCCCAC	735
Query	661	GCTTTCGCACCT	GAGCGTCAGTCTTTG	TCCAGGGGGGCCG	CCTTCGCCACCGG	TATTCCTC	720
Sbjct	734	GCTTTCGCACCT	GAGCGTCAGTCTTTG	TCCAGGGGGGCCG	CCTTCGCCACCGG	TATTCCTC	<mark>675</mark>
Query	721	CAGATCTCTACG	CATTTCACCGCTACA	CCTGGAATTCTA		GACTCAAG	780
Sbjct	674	CAGATCTCTACG	CATTTCACCGCTACA	CCTGGAATTCTA	ACCCCCCTCTACAA	GACTCTAG	615
Query	781	CCTGCCAGTTTC	AAATGCAGTTCCC-N	IGTTGAGCCCGGG	GATTTCACATCTG	ACTTAACA	839
Sbjct	614	CCTGCCAGTTTC	GAATGCAGTTCCCAG	GTTGAGCCCGGG	GATTTCACATCTG	ACTTGACA	555
Query	840	GACCGCCTGCGT	GCGCTTTACGCCCAG	TAATTCCGATTA	ACGCTTGCACCC	890	
Sbjct	554	GACCGCCTGCGT	GCGCTTTACGCCCAG	TAATTCCGATTA	ACGCTTGCACCC	504	



Candidatus Roseomonas massiliae 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KT321690.1|</u> Length: 1443 Number of Matches: 1

Range 1:	74 to 97	4 GenBank	<u>Graphics</u>				🔻 Next M	iatch 🔺 Pr	evious Match	
Score 1613 bits	s(1788)		Expect 0.0	Identities 898/901(99%)	((Gaps 0/901(0%))	Strand Plus/Plus		
Query	1	GGACGO	GTGAGTA	ACGCGTAGGAA	CGTGTCC	TGAGGT	GGGGG	ACAACCO	CGGGAAACTGG	60
Sbjct	74	GGACGO	GTGAGTA	ACGCGTAGGAA	CGTGTCC	TGAGGT	GGGGG	ACAACCO	CGGGAAACTGG	133
Query	61	GGCTAA	TACCGCA	ATATGGGCTGAG	GCCCAAA	GCCGAG	AGGCG	CCTTTGO	AGCGGCCTGCG	120
Sbjct	134	GGCTAR	ATACCGCA	ATATGGGCTGAG	GCCCAAA	GCCGAG	AGGCG	CCTTTGG	AGCGGCCTGCG	193
Query	121	TCCGAT	TAGGTAG	TTGGTGGGGTA	AAGGCCI	ACCAAG	CCTGC	GATCGGI	AGCTGGTCTGA	180
Sbjct	194	TCCGAI	TAGGTAG	GTTGGTGGGGTA	AAGGCCI	ACCAAG	CCTGC	GATCGGI	AGCTGGTCTGA	253
Query	181	GAGGAG	GACCAGO	CACACTGGGAC	TGAGACA		AGACTO	CCTACGO	GAGGCAGCAGT	240
Sbjct	254	GAGGAC	GACCAGO	CACACTGGGAC	TGAGACA	CGGCCC	AGACTO	CCTACGO	GAGGCAGCAGT	313
Query	241	GGGGAA	TATTGGA	ACAATGGGCGAA	AGCCTGA	TCCAGC	AATGC	CGCGTGG	GTGAAGAAGGT	300
Sbjct	314	GGGGAA	TATTGGA	ACAATGGGCGAA	AGCCTGA	TCCAGC	AATGC	CGCGTGG	GTGAAGAAGGT	373
Query	301	CTTCGG	ATCGTA	AGCCCTTTCGA	CGGGGA	GATGAT	GACGG	FACCCGI	AGAAGAAGCCC	360
Sbjct	374	CTTCGG	ATCGTAA	AGCCCTTTCGA	CGGGGA	GATGAT	GACGG	FACCCGI	AGAAGAAGCCC	433
Query	361	CGGCTA	ACTTCGI	GCCAGCAGCCG	CGGTAAI	ACGAAG	GGGGC	TAGCGTI	GCTCGGAATTA	420
Sbjct	434	CGGCTA	ACTTCGI	GCCAGCAGCCG	CGGTAAI	ACGAAG	GGGGC:	TAGCGTI	GCTCGGAATTA	493
Query	421	CTGGGG	GTAAAGG	GCGCGTAGGCG	GCGGCCC	AAGTCA	GGCGT	GAAATTO	CTGGGCTCAAC	480
Sbjct	494	CTGGGG	GTAAAGO	GCGCGTAGGCG	GCGGCCC	AAGTCA	GGCGT	GAAATTO	CTGGGCTCAAC	553
Query	481	CTGGGG	ACTGCGC	CTTGATACTGGG	TTGCTT	AGGATG	GAAGA	GCTCGI	GGAATTCCCAG	540
Sbjct	554	CTGGGG	ACTGCGC	CTTGATACTGGG	TTGCTTO	AGGATG	GAAGA	GCTCGI	GGAATTCCCAG	613
Query	541	TGTAGA	GGTGAAA	ATTCGTAGATAT	TGGGAAG	AACACC	GGTGGG	CGAAGGC	GGCGAGCTGGT	600
Sbjct	614	TGTAGA	AGGTGAAA	ATTCGTAGATAT	TGGGAAG	AACACC	GGTGGC	GAAGGO	GGCGAGCTGGT	673
Query	601	CCATTA	ACTGACGO	TGAGGCGCGAC	AGCGTGG	GGAGCA	AACAGO	GATTAGA	TACCCTGGTAG	660
Sbjct	674	CCATTA	ACTGACGO	TGAGGCGCGAC	AGCGTGG	GGAGCA	AACAGO	GATTAGA	TACCCTGGTAG	733
Query	661	TCCACO	CCGTAAA	ACGATGTGCGCT	GGATGTI	GGGGCC	CATAGO	GTCTCA	GTGTCGTAGCC	720
Sbjct	734	TCCACO	CCGTAAA	ACGATGTGCGCT	GGATGTI	GGGGGCC	CATAGO	GTCTCA	GTGTCGTAGCC	793
Query	721	AACGCG	GTAAGCG	GCACCGCCTGGG	GAGTACO	GCCGCA	AGGTTO	GAAACTC	AAAGGAATTGA	780
Sbjct	794	AACGCG	GTAAGCO	GCACCGCCTGGG	GAGTACO	GCCGCA	AGGTTO	GAAACTO	AAAGGAATTGA	853
Query	781	CGGGGG	CCCGCAC	CAAGCGGTGGAG	CATGTGG	TTTAAT	TCGAAG	GCAACGC	GCANAACCTTA	840
Sbjct	854	CGGGGG	CCCGCAC	CAAGCGGTGGAG	CATGTGG	TTTAAT	TCGAAG	GCAACGC	GCAGAACCTTA	913
Query	841	CCAGCO	CTTGACA	TGGTCACGACC	GGTCCAA	AAATGG	ACTTTO	CCTAGCA	ATAGGCGTGAT	900
Sbjct	914	CCAGCO	CTTGACA	ATGGTCACGACC	GGTCCAG	I IIII AGATGG	ACTTT	CCTAGCA	ATAGGCGTGAT	973
Query	901	G 901	L							
Sbjct	974	G 974	1							

с	[Quay_145579	
à	Uncultured bacterium clone ncd2760x02c1 16S ribosomal RNA gene, partial sequence	
	Roseomonas mucosa strain MDA5527 16S ribosomal RNA gene, partial sequence	
	Gandidatus Roseomonas massiliae 1461A 16S ribosomal RNA gene, partial sequence	
	8 Uncultured bacterium clone 16slp96-1b10.p1k 16S ribosomal RNA gene, partial sequence	
	Abacteria 2 leaves	
	Roseomonas mucosa strain ES_21con 16S ribosonal RNA gene, partial sequence	
	Uncultured bacterium clone ncd502g04c1 16S ribosomal RNA gene, partial sequence	
	Abacteria 5 leaves	
	Abacteria 7 leaves	
	Abacteria 7 leaves	
	Autoria / 7 leaves	
	Stacteria 7 leaves	
	g bacteria 7 leaves	
	bacteria 2 leaves	
	Roseomonas sp. CMS4Y-2-2 16S ribosomal RNA gene, partial sequence	
	abacteria 2 leaves	
	Roseomonas terpenica partial 16S rRNA gene, type strain DT	
	Our Concultured bacterium clone ncd191h10c1 16S ribosomal RNA gene, partial sequence	
	Abacteria 5 leaves	
0.0003	Deateria 7 leaves	
1	Abacteria 7 leaves	
	Subacteria 7 leaves	
	*bacteria 2 leaves	
	Candidatus Roseomonas massiliae 16S ribosomal RNA gene, partial sequence	
	Poacteria 2 leaves	
	Roseomonas mucosa strain EHFS2_505Hb 16S ribosonal RNA gene, partial sequence	
	Uncultured bacterium clone ncd191h11c1 16S ribosomal RNA gene, partial sequence	
	fourteria 2 leaves	
	Roscomonas mucosa strain SM14032013 16S ribosomal RNA gene, partial sequence	
	Pacteria 2 leaves	
. 8893	tucteria 2 leaves	
	Moateria 2 leaves	
	Roseomonas sp. BZ44 16S ribosomal RNA gene, partial sequence	
	*bocteria 2 leaves	

Pseudomonas parafulva KT758848.1

Pseudomonas parafulva strain YAB-1 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KT758848.1</u>] Length: 1401 Number of Matches: 1

Range 1: 3	39 to 98	0 GenBank Graphics		🔻 Next Match 🔺 Pr	revious Match	
Score 1651 bits	s(1830)	Expect Identi 0.0 933/9	ties Gaps (42(99%) 2/942(09	6) Strand 6) Plus/Plus	;	
Query	1	AGCGGCGGACGGGTGAGI	TAATGCCTAGGAATCTGCC	TGGTAGTGGGGG	ACAACGTTTCGA	60
Sbjct	39	AGCGGCCGGACGGGTGAGT	TAATGCCTAGGAATCTGCC	TGGTAGTGGGGG	ACAACGTTTCGA	98
Query	61	AAGGAACGCTAATACCGC	CATACGTCCTACGGGAGAA	AGCAGGGGACCTI	ICGGGCCTTGCG	120
Sbjct	99	AAGGAACGCTAATACCGC	CATACGTCCTACGGGAGAA	AGCAGGGGGACCT	ICGGGCCTTGCG	158
Query	121	CTATCAGATGAGCCTAG	TCGGATTAGCTTGTTGGT	GAGGTAATGGCT	CACCAAGGCGAC	180
Sbjct	159	CTATCAGATGAGCCTAG	GTCGGATTAGCTTGTTGGT	GAGGTAATGGCT	CACCAAGGCGAC	218
Query	181	GATCCGTAACTGGTCTGA	AGAGGATGATCAGTCACAC	TGGAACTGAGACA	ACGGTCCAGACT	240
Sbjct	219	GATCCGTAACTGGTCTGA	AGAGGATGATCAGTCACAC	TGGAACTGAGAC	ACGGTCCAGACT	278
Query	241	CCTACGGGAGGCAGCAGI	GGGGAATATTGGACAATG	GGCGAAAGCCTGA	ATCCAGCCATGC	300
Sbjct	279	CCTACGGGAGGCAGCAGI	GGGGAATATTGGACAATG	GGCGAAAGCCTG	ATCCAGCCATGC	338
Query	301	CGCGTGTGTGAAGAAGG	CTTCGGATTGTAAAGCAC	TTTAAGTTGGGAG	GGAAGGGTTGTA	360
Sbjct	339	CGCGTGTGTGAAGAAGGI	CTTCGGATTGTAAAGCAC	TTTAAGTTGGGAG	GGAAGGGTTGTA	398
Query	361	GATTAATACTCTGCAATI	TTGACGTTACCGACAGAA	TAAGCACCGGCT	AACTCTGTGCCA	420
Sbjct	399	GATTAATACTCTGCAATI	TTGACGTTACCGACAGAA	TAAGCACCGGCT	ACTCTGTGCCA	<mark>4</mark> 58
Query	421	GCAGCCGCGGTAATACAG	GAGGGTGCAAGCGTTAATC	GGAATTACTGGGG	CGTAAAGCGCGC	480
Sbjct	459	GCAGCCGCGGTAATACAG	GAGGGTGCAAGCGTTAATC	GGAATTACTGGGG	CGTAAAGCGCGC	518
Query	481	GTAGGTGGTTTGTTAAGT	TGGATGTGAAAGCCCCGG	GCTCAACCTGGG	AACTGCATCCAA	540
Sbjct	519	GTAGGTGGTTTGTTAAGI	TGGATGTGAAAGCCCCGG	GCTCAACCTGGGZ	ACTGCATCCAA	578
Query	541	AACTGGCAAGCTAGAGTA	ACGGTAGAGGGTGGTGGAA		CGGTGAAATGCG	600
Sbjct	579	AACTGGCAAGCTAGAGTA	ACGGTAGAGGGTGGTGGAA	TTTCCTGTGTAG	CGGTGAAATGCG	<mark>6</mark> 38
Query	601	TAGATATAGGAAGGAACA	ACCAGTGGCGAAGGCGACC	ACCTGGACTGAT	ACTGACACTGAG	660
Sbjct	639	TAGATATAGGAAGGAACA	ACCAGTGGCGAAGGCGACC	ACCTGGACTGAT	ACTGACACTGAG	698
Query	661	GTGCGAAAGCGTGGGGAG	CAAACAGGATTAGATACC	CTGGTAGTCCAC	GCCGTAAACGAT	720
Sbjct	699	GTGCGAAAGCGTGGGGGAG	GCAAACAGGATTAGATACC	CTGGTAGTCCAC	GCCGTAAACGAT	758
Query	721	GTCAACTAGCCGTTGGAA	ATCCTTGAGATTTTAGTGG	CGCAGCTAACGC	ATTAAGTTGACC	780
Sbjct	759	GTCAACTAGCCGTTGGAA	ATCCTTGAGATTTTAGTGG	CGCAGCTAACGC	ATTAAGTTGACC	818
Query	781	GCCTGGGGAGTACGGCCG	GCAAGGTT-AAACTCAAAT	GAATTGACGGGGG	GCCCGCACAAGC	839
Sbjct	819	GCCTGGGGGAGTACGGCCG	GCAAGGTTAAAACTCAAAT	GAATTGACGGGG	GCCCGCACAAGC	878
Query	840	GGTGGAGCATGTGGTTTA	ATTCGAAACAACGCGAAA	AACCTTACCAGG	CCTTGACATGCA	899
Sbjct	879	GGTGGAGCATGTGGTTT	ATTCGAAGCAACGCGAAG	AACCTTACCAGG	CCTTGACATGCA	938
Query	900	AAAAACTTTCCAAAAATG	GGATTGGGGGCCTTCGGG-A	CTCTGA 940		
Sbjct	939	GAGAACTTTCCAGAGAT	GATTGGTGCCTTCGGGAA	CTCTGA 980		

	Pseudomonas sp. 4 16S ribosomal RNA gene, partial sequence	
	Pseudomonas parafulva strain AJ 2129 16S ribosomal RNA gene, partial sequence	
	 Pseudomonas paralulva strain AJ 2129 165 ribosomal KNA gene, partial sequence Pseudomonas sp. Fa24 165 ribosomal RNA gene, rartial sequence 	
	Uncultured bacterium clone ned2626a08c1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva strain EX4 16S ribosomal RNA gene, partial sequence	
	Pseudomonas putida strain CLF2014-1 16S ribosonal RNA gene, partial sequence	
	notocharteria 2 leaves	
	Pseudomonas fulva strain 1Y1103 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone nbw621f12c1 16S ribosomal RNA gene, partial sequence	De denne file and 7 MY 1/2 de al DVI and and de
	Broadomonas sp. DP108 16S ribosomal RNA cone, partial sequence	Pseudonionas luiva strain i/_MX 10S nbosomai KNA gene, partial sequence
	Pseudomonas sp. NCCP-562 gene for 16S ribosomal RNA, partial sequence	
	Abacteria 2 leaves	
	Pseudomonas sp. NCCP-553 gene for 168 ribosomal RNA, partial sequence Discributed bacterisism close abd 18(11) 168 ribosomal RNA some methal companies.	
	Constant of the main test and the rest of the second states from particular subjects to	Deudomonas fulva strain i13_MX 16S ribosomal RNA gene, partial sequence
	-proteobacteria 4 keaves	
	Pseudomonas fulva gene for 16S rRNA, partial sequence, strain: NBRC 16638 Providence folio 67 16S relevand DNA entry partial sequences	
	President of the statement of the second of	Uncultured bacterium clone nck137a07c1 16S ribosomal RNA gene, partial sequence
	Endophytic bacterium SV807 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. NCCP-571 gene for 16S ribosomal RNA, partial sequence	
	secondinatas sp. N. W-1 105 filosofiai KNA gene, partai sequence	Uncultured bacterium clone ncd2661h11c1 16S ribosomal RNA gene, partial sequence
	Pseudomonas fulva strain SW32 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva strain iMX35 16S ribosomal RNA gene, partial sequence	
	Circumited bacterium cione (554955 105 ribosonia) KNA gene, partial sequence	Pseudomonas parafulya strain L4-302 16S ribosomal RNA cone martial concerns
		Pseudomonas fulva strain AKB22 16S ribosomal RNA gene, partial sequence
		Pseudomonas fulva strain ANA72 16S ribosomal RNA gene, partial sequence
	2	Pseudomonas sp. ANA/1105 ribosomal KNA gene, partial sequence Uncultured bacterium close ned905h11c116S ribosomal RNA cene, partial sequence.
1001		Beudomonas fulva partial 165 rRNA gene, strain YMC09/4/B4619
	Pseudomonas parafulva strain NBRC 16636 16S ribosomal RNA cene, partial sequence	101 222
	Uncultured bacterium clone ncd2803c07c1 16S ribosomal RNA gene, partial sequence	
	\$	Pseudomonas sp. MH-135 16S rRNA gene, strain MH-135
	Pseudomonas fulva strain BSW6 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. enrichment culture clone 8 16S ribosomal RNA gene, partial sequence	
	Seudomonas sp. 3zhy partial 16S rRNA gene, strain 3zhy	The short he station close as (NG401a) 165 shareand DVA case and al conserva-
	Devadaments on MG 2011.7. Bl nartial 16S (DNA cone. strain 7 isolate BL	 Cneutarea bacteriam cione neuzosti i rei 105 mosoniai Kov gene, partai sequenci
	Uncultured bacterium clone nbt22b10 16S ribosomal RNA gene, partial sequence	
	Second se	Pseudomonas fulva strain iP1 MX 16S ribosomal RNA gene, partial sequence
	Pseudomonas sp. DP109B 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. NCCP-561 gene for 16S ribosomal RNA, partial sequence	
	Pseudomonas sp. 2B5B 16S ribosomal RNA gene, partial sequence	
	the barther in the shift with the shared BM and a shift and	Oncultured bacterium clone MWL-15 16S ribosomal RNA gene, partial sequence
	Cheulared nacienam cione next3 /d0/201105 ribosomai KiNA gene, partial sequence	
	Breddomonus sp. UKSIOLI w K12:111 genome assenioly Tellemolgz, enromosome : T	Brandomonos parafules strain (PS01.1, complete apromo.
		Predomonas spiraturva strain CR501-1, complete genome
	ò	Pseudomonas sp. LB-S-GYM-2 16S ribosomal RNA gene, partial sequence
		Uncultured bacterium clone ned2686h02c1 16S ribosomal RNA gene, partial sequence
	Pseudomonas fulva strain BSW3 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. B-6 16S ribosomal RNA gene, partial sequence	
	Preudomonas sp. H10zhy partial 16S rRNA gene, strain H10zhy	A11
	- neutodouteria 6 louves	 Uncummed bacterium cione ncd361g02c1 16S ribosomal KNA gene, partial sequence
	Prodomonas fulva strain SCRA109.165 ribosomal RNA sone portial common	
	Uncultured bucterium clone ncd2598/06c1 165 ribosomal RNA cene, nartial sequence	
	A second s	Pseudomonas fulva strain QC07 16S ribosomal RNA gene, partial sequence
	Pseudomonas fulva strain BSW1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva 16S ribosomal RNA gene, partial sequence	
	Beudomonas fulva partial 16S rRNA gene, strain Z67zhy	
		Oncultured bacterium clone ncd908e02c1 16S ribosomal RNA gene, partial sequence
	Pseudomonas fulva gene for 165 rRNA, partial sequence, strain: NBRC 16639	
	Cincultured bacterium cione nowbesevici i tos noosomai KNA gene, partial sequence	O Demologicano false strain 110 MY 165 ribosom I DNA conc. partial common
	Budging AM0215 165 channel DVA and addition	 Pseudonionas nuva sirain 117_NIA 105 noosonai KNA gene, partai sequence
	bacerium Abio515 105 notsoniai KivA gene, partiai sequence	
	Pseudomonas sp. NCCP-567 gene for 16S ribosomal RNA, partial sequence	
	Pseudomonas sp. 471-1 16S ribosomal RNA gene, partial sequence	
	A support of the second	OF Incultured hysterium close and VC000641 16S also could PNA gene method courses
	Benderson and Antonia 17 1231 1/C - Annual DV1 and and a	 OR many overman concrete to a nonomal root gare, paras sequence
	Pseudomonas punda strain JT-K21 16S ribosomal KNA gene, partial sequence	
	Uncultured bacterium partial 16S rRNA gene, clone Ca07 74	
	neutasharteria 4 leaves	
	A Decemental a rearch	
	re-proteobacteria 3 leaves	
	Pseudomonas fulva strain BSW8 16S ribosomal RNA gene, partial sequence	
	Perudomonas fulva strain CI-11 16S ribocomil PNA cone, nutrial comence.	
	De la contra sur la contra sur contra de la	
	Beudomonas punda strain F29 165 ribosomal RNA gene, partial sequence	
		¹⁰ Uncultured bacterium clone ncd2625e06c1 16S ribosomal RNA gene, partial sequence

Bacillus licheniformis KU314515.1

Bacillus licheniformis strain HS10 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|KU314515.1</u>| Length: 1514 Number of Matches: 1

Range 1:	491 to 14	41 GenBank Graph	ics		🔻 Next Match 🔺 Previous	Match	
Score 1710 bits	s(1896)	Expect 0.0	Identities 950/951(99%)	Gaps 0/951(0%)	Strand Plus/Minus		
Query	1	CACCGACTTCG	GGTGTTACAAACT	CTCGTGGTGTGA	CGGGCGGTGTGTACAZ	AGGCCCGGG	60
Sbjct	1441	CACCGACTTCG	GGTGTTACAAACT		CGGGCGGTGTGTACA	Feeccceee	1382
Query	61	AACGTATTCAC	CGCGGCATGCTGA	ICCGCGATTACT	AGCGATTCCAGCTTC	ACGCAGTCG	120
Sbjct	1381	AACGTATTCAC	CGCGGCATGCTGA	ICCGCGATTACT	AGCGATTCCAGCTTC	ACGCAGTCG	1322
Query	121	AGTTGCAGACT	GCGATCCGAACTG	AGAACAGATTTG	TGGGATTGGCTTAGC	CTCGCGGCT	180
Sbjct	1321	AGTTGCAGACT	GCGATCCGAACTG	AGAACAGATTTG	TGGGATTGGCTTAGC	TCGCGGCT	1262
Query	181	TCGCTGCCCTT	TGTTCTGCCCATT	GTAGCACGTGTG	TAGCCCAGGTCATAAG	GGGCATGA	240
Sbjct	1261	TCGCTGCCCTT	TGTTCTGCCCATT	GTAGCACGTGTG	TAGCCCAGGTCATAAG	GGGCATGA	1202
Query	241	TGATTTGACGT	CATCCCCACCTTC	CTCCGGTTTGTC	ACCGGCAGTCACCTT	AGAGTGCCC	300
Sbjct	1201	TGATTTGACGT	CATCCCCACCTTC	CTCCGGTTTGTC	ACCGGCAGTCACCTT	AGAGTGCCC	1142
Query	301	AACTGAATGCT	GGCAACTAAGATCA	AAGGGTTGCGCT	CGTTGCGGGACTTAAC	CCCAACATC	360
Sbjct	1141	AACTGAATGCT	GGCAACTAAGATCA	AAGGGTTGCGCT	CGTTGCGGGACTTAAC	CCCAACATC	1082
Query	361	TCACGACACGA	GCTGACGACAACCA	ATGCACCACCTG	TCACTCTGCCCCCGAZ	AGGGGAAGC	420
Sbjct	1081	TCACGACACGA	GCTGACGACAACCA	ATGCACCACCTG	TCACTCTGCCCCCGA	AGGGGAAGC	1022
Query	421	CCTATCTCTAG	GGTTGTCAGAGGA	IGTCAAGACCTG	GTAAGGTTCTTCGCG	TGCTTCGA	480
Sbjct	1021	CCTATCTCTAG	GGTTGTCAGAGGA	IGTCAAGACCTG	GTAAGGTTCTTCGCG	TGCTTCGA	962
Query	481	ATTAAACCACA	TGCTCCACCGCTT	STGCGGGCCCCC	GTCAATTCCTTTGAG	TTCAGTCT	540
Sbjct	961	ATTAAACCACA	TGCTCCACCGCTT(GTGCGGGCCCCC	GTCAATTCCTTTGAG	TTCAGTCT	902
Query	541	TGCGACCGTAC	TCCCCAGGCGGAG	IGCTTAATGCGT	TTGCTGCAGCACTAA	AGGGCGGAA	600
Sbjct	901	TGCGACCGTAC	TCCCCAGGCGGAG	IGCTTAATGCGT	TTGCTGCAGCACTAA	AGGGCGGAA	842
Query	601	ACCCTCTAACA	CTTAGCACTCATCO	TTTACGGCGTG	GACTACCAGGGTATCT	AATCCTGT	660
Sbjct	841	ACCCTCTAACA	CTTAGCACTCATCO	TTTACGGCGTG	GACTACCAGGGTATCT	AATCCTGT	782
Query	661	TCGCTCCCCAC	GCTTTCGCGCCTC	AGCGTCAGTTAC.	AGACCAGAGAGTCGCC	TTCGCCAC	720
Sbjct	781	TCGCTCCCCAC	GCTTTCGCGCCTC	AGCGTCAGTTAC.	AGACCAGAGAGTCGCC	TTCGCCAC	722
Query	721	TGGTGTTCCTC	CACATCTCTACGCA	ATTTCACCGCTA	CACGTGGAATTCCACT	CTCCTCTT	780
Sbjct	721	TGGTGTTCCTC	CACATCTCTACGC	ATTTCACCGCTA	CACGTGGAATTCCACT	CTCCTCTT	662
Query	781	CTGCACTCAAG	TTCCCCAGTTTCCZ	ATGACCCTCCC	CGGTTGAGCCGGGGGC	TTTCACAT	840
Sbjct	661	CTGCACTCAAG	TTCCCCAGTTTCC	ATGACCCTCCC	CGGTTGAGCCGGGGGC	TTTCACAT	602
Query	841	CAAACTTAAGA	AACCGCCTGCGCGC	GCTTTACGCCC.	AATAATTCCGGACAAC	GCTTGCCA	900
Sbjct	601	II IIIIIII CAGACTTAAGA	AACCGCCTGCGCGC	GCTTTACGCCC.	AATAATTCCGGACAAC	GCTTGCCA	542
Query	901	CCTACGTATTA	CCGCGGCTGCTGG	CACGTAGTTAGC	CGTGGCTTTCTGGTT	951	
Sbjct	541	CCTACGTATTA	CCGCGGGCTGCTGGC	CACGTAGTTAGC	CGTGGCTTTCTGGTT	491	

Bacillus sp. BAB-5485 16S ribosomal RNA gene, partial sequence Bacillus cp. BK IDB (018 partial 16S rBNA gene, isolate BK IDB (018)	
Bacillus sp. RKNM-0105 partial 16S rRNA gene, isolate RKNM-0105	
Bacillus licheniformis strain KP050r 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain KP052r 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain HS10 16S ribosomal RNA gene, partial sequence	
Bacillus sp. BAB-5636 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain SIB_Zn_R4 16S ribosomal RNA gene, partial sequent	ce
Bacillus licheniformis strain FJAT-29133 16S ribosomal RNA gene, partial sequen	ice
Bacillus sp. S94(2016) 16S ribosomal RNA gene, partial sequence Bacillus sp. S37 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain VTB12 16S ribosomal RNA gene, partial sequence	
Bacillus inchemiormis strain CQN-22 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain CQN-12 I6S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain CQN-11 105 ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain CY2-24 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain CY1-20 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain Y8 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain KTNB0010 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain SMR8 16S ribosomal RNA gene, partial sequence	ce
Bacillus licheniformis strain SMR7 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain EVR2 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain RA5UN 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain RA13UN 16S ribosomal RNA gene, partial sequence	
Uncultured bacterium clone 958-15 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis 16S ribosomal RNA gene, partial sequence Bacillus sp. C-3-24 16S ribosomal RNA gene, partial sequence	
Bacillus sp. C-3-11 16S ribosomal RNA gene, partial sequence	
Bacillus sp. C-3-3 16S ribosonal RNA gene, partial sequence	
Bacillus sp. B-3-7 16S ribosomal RNA gene, partial sequence Bacillus sp. A-3-12 16S ribosomal RNA gene, partial sequence	
Bacillus sp. A-3-5 16S ribosomal RNA gene, partial sequence	
Bacillus sp. A-2-29 IoS ribosomal RNA gene, partial sequence Bacillus sp. A-2-18 IoS ribosomal RNA gene, partial sequence	
Bacillus sp. A-2-13 16S ribosomal RNA gene, partial sequence Bacillus op C-130 16S ribosomal RNA gene partial sequence	
Bacillus sp. C-1-20 165 ribosomal RNA gene, partial sequence	
Bacillus sp. C-1-3 IoS ribosomal RNA gene, partial sequence Bacillus sp. B-1-40 I6S ribosomal RNA gene, partial sequence	
Bacillus sp. B-1-16 16S ribosomal RNA gene, partial sequence Bacillus sp. B-1-15 16S ribosomal RNA gene, partial sequence	
Bacillus sp. B31(2015) 16S ribosomal RNA gene, partial sequence Bacillus sp. B27(2015) 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain F1159 16S ribosomal RNA cene, partial sequence	
Bacillus licheniformis strain FI152 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain FI47 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain FI44 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain I-A-E-34 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain IS-1 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain DMB31 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain NCIM2405 165 ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain NCIM2471 165 ribosomal RNA gene, partial sequence	
Bacillus sp. NG4-2 16S ribosomal RNA gene, partial sequence Bacillus sp. NL3-1 16S ribosomal RNA gene, partial sequence	
 Bacillus sp. NZ3-2 16S ribosomal RNA gene, partial sequence Bacillus sp. NZ3-1 16S ribosomal RNA gene, partial sequence 	
Bacillus flexus strain NZ2-2 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain 103D-012 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain N9 16S ribosomal RNA gene, partial sequence Bacterium ARb15 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain CY-012 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain IHBB 11006 16S ribosomal RNA gene, partial sequence	
Bacillus sp. BAB-5495 16S ribosomal RNA gene, partial sequence	
Bacillus sp. N33621VB01 1492r 16S ribosomal RNA gene, partial sequence	
Bacillus sp. FJAT-22511 16S ribosonal RNA gene, partial sequence	
Bacillus licheniformis partial 165 rKNA gene, strain IRQBAS20 Bacillus licheniformis partial 165 rRNA gene, strain IRQBAS19	
Bacillus licheniformis partial 16S rRNA gene, strain IRQBAS18 Bacillus sp. NCIM 2131 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain KAR76 16S ribosomal RNA gene, complete sequence Bacillus licheniformis WX-02 genome	
Bacillus sp. KP138r 16S ribosomal RNA gene, partial sequence Bacillus sp. KP129r 16S ribosomal RNA gene, partial sequence	
Bacillus sp. KP126r 16S ribosomal RNA gene, pantial sequence	
Bacillus accentionnis strain KY1207 105 ribosomai KNA gene, partial sequence Bacillus sp. KP097r 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain KP093r 16S ribosomal RNA gene, partial sequence Bacillus licheniformis strain KP091r 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain KP081r 16S ribosomal RNA gene, partial sequence	
Bacillus sp. KP0/1r 16S nbosomal KNA gene, partial sequence Bacillus sp. KP057r 16S ribosomal RNA gene, partial sequence	
Bacillus licheniformis strain KP056r 16S ribosomal RNA gene, partial sequence Decilius on KP055r 16S chaptered RNA sequence partial sequence	
working syn revolt too noosonia rivy gene, panar sequence	lclQuery 71993
Bacillus sp. KP053r 16S ribosomal RNA gene, partial sequence Bacillus sp. H15-1 16S ribosomal RNA gene, partial sequence	

Pseudomonas aeruginosa KF680991.1

Pseudomonas aeruginosa strain ATHA23 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KF680991.1|</u> Length: 1115 Number of Matches: 1

Range 1: 8	86 to 106	5 GenBank Graphic	5		V Next Match 🔺 Previous N	1atch	
Score 1763 bits	s(1954)	Expect 0.0	Identities 979/980(99%)	Gaps 0/980(0%)	Strand P <mark>lu</mark> s/Minus		
Query	1	TTCTGGAGCAA	CCCACTCCCATG	GTGTGACGGGCG	GTGTGTACAAGGCCCGG	GAACGTAT	60
Sbjct	1065	TTCTGGAGCAA	CCCACTCCCATG	GTGTGACGGGCG	JIIIIIIIIIIIIIIIIIII GTGTGTACAAGGCCCGG	 GAACGTAT	1006
Query	61	TCACCGTGACA	TTCTGATTCACG	ATTACTAGCGAT:	ICCGACTTCACGCAGTC	GAGTTGCA	120
Sbjct	1005	TCACCGTGACA	TTCTGATTCACG	ATTACTAGCGAT	ICCGACTTCACGCAGTC	GAGTTGCA	946
Query	121	GACTGCGATCC	GGACTACGATCG	GTTTTATGGGAT:	FAGCTCCACCTCGCGGC	TTGGCAAC	180
Sbjct	945	GACTGCGATCC	GGACTACGATCG	GTTTTATGGGAT	IAGCTCCACCTCGCGGC	TTGGCAAC	886
Query	181	CCTTTGTACCG	ACCATTGTAGCA	CGTGTGTAGCCC	IGGCCGTAAGGGCCATG	ATGACTTG	240
Sbjct	885	CCTTTGTACCG	ACCATTGTAGCA	CGTGTGTAGCCC	IGGCCGTAAGGGCCATG	ATGACTTG	826
Query	241	ACGTCATCCCC	ACCTTCCTCCGG	TTTGTCACCGGC	AGTCTCCTTAGAGTGCC	CACCCGAG	300
Sbjct	825	ACGTCATCCCC	ACCTTCCTCCGG	TTGTCACCGGC	AGTCTCCTTAGAGTGCC	CACCCGAG	766
Query	301	GTGCTGGTAAC	TAAGGACAAGGG'	TTGCGCTCGTTA	CGGGACTTAACCCAACA	TCTCACGA	360
Sbjct	765	GTGCTGGTAAC	TAAGGACAAGGG	TTGCGCTCGTTA	CGGGACTTAACCCAACA	TCTCACGA	706
Query	361	CACGAGCTGAC	GACAGCCATGCA	GCACCTGTGTCT	GAGTTCCCGAAGGCACC	AATCCATC	420
Sbjct	705	CACGAGCTGAC	GACAGCCATGCA	GCACCTGTGTCT	GAGTTCCCGAAGGCACC	AATCCATC	646
Query	421	TCTGGAAAGTT	CTCAGCATGTCA	AGGCCAGGTAAG	GTTCTTCGCGTTGCTTC	GAATTAAA	480
Sbjct	645	TCTGGAAAGTT	CTCAGCATGTCA	AGGCCAGGTAAG	JIIIIIIIIIIIIIIIIIIII GTTCTTCGCGTTGCTTC	GAATTAAA	586
Query	481	CCACATGCTCC	ACCGCTTGTGCG	GGCCCCCGTCAA	ITCATTTGAGTTTTAAC	CTTGCGGC	540
Sbjct	585	CCACATGCTCC	ACCGCTTGTGCG	GGCCCCCGTCAA	IIIIIIIIIIIIIIIIIIIII TTCATTTGAGTTTTAAC	CTTGCGGC	526
Query	541	CGTACTCCCCA	GGCGGTCGACTT	ATCGCGTTAGCT	GCGCCACTAAGATCTCA	AGGATCCC	600
Sbjct	525	CGTACTCCCCA	GGCGGTCGACTT	ATCGCGTTAGCT	GCCACTAAGATCTCA	AGGATCCC	466
Query	601	AACGGCTAGTC	GACATCGTTTAC	GGCGTGGACTAC	CAGGGTATCTAATCCTG	TTTGCTCC	660
Sbjct	465	AACGGCTAGTC	GACATCGTTTAC	GGCGTGGACTAC	CAGGGTATCTAATCCTG	TTTGCTCC	406
Query	661	CCACGCTTTCG	CACCTCAGTGTC	AGTATCAGTCCA	GGTGGTCGCCTTCGCCA	CTGGTGTT	720
Sbjct	405	CCACGCTTTCG	CACCTCAGTGTC	AGTATCAGTCCA	GTGGTCGCCTTCGCCA	CTGGTGTT	346
Query	721	CCTTCCTATAT	CTACGCATTTCA	CCGCTACACAGG	AAATTCCACCACCCTCT	ACCGTACT	780
Sbjct	345	CCTTCCTATAT	CTACGCATTTCA	CCGCTACACAGG	AAATTCCACCACCCTCT	ACCGTACT	286
Query	781	CTAGCTCAGTA	GTTTTGGATGCA	GTTCCCAGGTTG		TCCAACTT	840
Sbjct	285	CTAGCTCAGTA	GTTTTGGATGCA	GTTCCCAGGTTG	AGCCCGGGGGATTTCACA	TCCAACTT	226
Query	841	GCTGAACCACC	TACGCGCGCTTT.	ACGCCCAGTAAT	ICCGATTAACGCTTGCA	CCCTTCGT	900
Sbjct	225	GCTGAACCACC	TACGCGCGCTTT	ACGCCCAGTAAT	ICCGATTAACGCTTGCA	CCCTTCGT	166
Query	901	ATTACCGCGGC	TGCTGGCACGAA	GTTAGCCGGGGC'		TCAAACAG	960
Sbjct	165	ATTACCGCGGC	TGCTGGCACGAA	GTTAGCCGGTGC'	TATTCTGTTGGTAACG	TCAAACAG	106
Query	961	CAAGGTATTAA	CTTACTGCC 9	80			
Sbjct	105	CAAGGTATTAA	CTTACTGCC 8	6			

 Icl[Query 70005]
 Pseudomönas aeruginosa SJTD-1, complete genome
 Bacterium NV2A 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain RS1 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain CAU342A 16S ribosomal RNA gene, partial sequence
 Pseudomonas senzuginosa strain PS1 16S ribosomal RNA gene, partial sequence
 Pseudomonas senzuginosa strain PS5 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain PS5 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain FX 18S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain FX 18S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain FX 18S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain PS4 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain FX 18S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain PS4 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain PS4 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain PS4 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain ATCC 27853, complete genome
 Pseudomonas aeruginosa strain P9670, complete genome
 Pseudomonas aeruginosa strain V3662, complete genome
 Pseudomonas aeruginosa strain V36968, complete genome
 Pseudomonas aeruginosa strain H27930, complete genome
 Pseudomonas aeruginosa strain W36968, complete genome
 Pseudomonas aeruginosa strain W36978, complete genome
 Pseudomonas aeruginosa strain W36978, complete genome
 Pseudomonas aeruginosa strain W36978, complete genome Pseudomonas aeruginosa strain S680/8, complete genome
 Pseudomonas aeruginosa strain S680/8, complete genome
 Pseudomonas aeruginosa strain S680/8, complete genome
 Pseudomonas aeruginosa strain F23197, complete genome
 Pseudomonas aeruginosa strain N15-01092, complete genome
 Pseudomonas aeruginosa strain N15-01092, complete genome
 Pseudomonas aeruginosa strain N15-01092, complete genome
 Pseudomonas aeruginosa strain T52373, complete genome
 Pseudomonas aeruginosa strain T52373, complete genome
 Pseudomonas aeruginosa strain T52373, complete genome
 Pseudomonas aeruginosa strain R269 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain R269 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain R269 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain R269 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain R265 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain R263 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain R263 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain AM-1-1-1 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain I-M-2-2-3 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain I-M-2-2-1 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain G-R-5-3-1 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain ASA 102 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain ASA 102 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain ASA 102 16S ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain MSA 164S 165 ribosomal RNA gene, partial sequence< Peedomonas aeruginosa strain BBSR1 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain BBSR1 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain BBPO16 165 ribosomal RNA gene, partial sequence Uncultured bacterium clone PLB97 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DMB41 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM141 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM14 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM14 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM14 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM14 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM11 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM11 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM11 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM11 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM11 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain DM11 165 ribosomal RNA gene, partial sequence Viscultured bacterium partial 165 rRNA cene, clone 16ms 16-2e111 p1k Pseudomonas aeruginosa strain DM2 16S ribosomal RNA gene, partial sequence Pseudomonas aeruginosa strain VA-134, complete genome Pseudomonas aeruginosa strain VA-134, complete genome Pseudomonas aeruginosa strain UKMP14 16S ribosonial RNA gene, partial sequence Pseudomonas aeruginosa strain VKR05 16S ribosonial RNA gene, partial sequence Pseudomonas aeruginosa strain VKR05 16S ribosonial RNA gene, partial sequence Bacterium GX7 165 ribosonal RNA gene, partial sequence Proteobacteria | 11 leaves
 Pseudomonas aeruginosa strain HMT 7 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain HMT 1 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain A1 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain A1 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain NBAII AFP-6 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain NBAII AFP-3 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain NBAII AFP-3 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain NBAII AFP-3 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain NBAII AFP-3 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain n II 105 ribosomal RNA, partial sequence
 Pseudomonas aeruginosa strain SAII ND-41 LART-B 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain n SAII ND-41 ART-B 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain n SAII ND-41 ART-B 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain n SAII ND-41 AS ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain n SAI 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain AI N2 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain BM6 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain BM6 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain AI THA23 165 ribosomal RNA gene, partial sequence
 Pseudomonas aeruginosa strain AI THA23 165 ribosomal RNA gene, partial sequence Pseudomonas aeruginosa AES-IR, complete genome

318

Agrobacterium larrymoorei strain 2R46 16S ribosomal RNA gene, partial sequence sequence ID: <u>gb|EF178437.1|</u> Length: 1414 Number of Matches: 1

Range 1:	398 to 13	29 GenBank Graphics			🗸 Next Match 🔺 Previous Match	
Score 1676 bit	s(1858)	Expect Id 0.0 93	entities 1/932(99%)	Gaps 0/932(0%)	Strand Plus/Minus	
Ouerv	1	GGGTAAAACCAACI	CCCATGGTGTG	ACGGGCGGTGT	GTACAAGGCCCGGGAACGTATTCA	60
Sbjct	1329	 GGGTAAAACCAACI	 CCCATGGTGTG	ACGGGCGGTGT		1270
Query	61	CCGCAGCATGCTGA	TCTGCGATTAC	TAGCGATTCCA	ACTTCATGCACTCGAGTTGCAGAG	120
Sbjct	1269	CCGCAGCATGCTGA	TCTGCGATTAC	TAGCGATTCCA	ACTTCATGCACTCGAGTTGCAGAG	1210
Query	121	TGCAATCCGAACTG	AGATGGCTTTT	GGAGATTAGCT	CGACATCGCTGTCTCGCTGCCCAC	180
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Query	181	TGTCACCACCATTO	TAGCACGTGTG	TAGCCCAGCCC	STAAGGGCCATGAGGACTTGACGT	240
Sbjct	1149	TGTCACCACCATTO	TAGCACGTGTG	TAGCCCAGCCC	GTAAGGGCCATGAGGACTTGACGT	1090
Query	241	CATCCCCACCTTCC	TCTCGGCTTAT	CACCGGCAGTC	CCCTTAGAGTGCCCAACCAAATGC	300
Sbjct	1089	CATCCCCACCTTCC	TCTCGGCTTAT	CACCGGCAGTC	CCCTTAGAGTGCCCAACCAAATGC	1030
Query	301	TGGCAACTAAGGGC	GAGGGTTGCGC	TCGTTGCGGGA	CTTAACCCAACATCTCACGACACG	360
Sbjct	1029	TGGCAACTAAGGGC	GAGGGTTGCGC	TCGTTGCGGGA	CTTAACCCAACATCTCACGACACG	970
Query	361	AGCTGACGACAGCO	ATGCAGCACCT	GTTCTAGGGCC	AGCCGAACTGAAGGTCATCGTCTC	420
Sbjct	969	AGCTGACGACAGCO	ATGCAGCACCT	GTTCTAGGGCC	AGCCGAACTGAAGGTCATCGTCTC	910
Query	421	CAATGACCATACCO	CGAATGTCAAG	AGCTGGTAAGG	ITCTGCGCGTTGCTTCGAATTAAA	480
Sbjct	909	CAATGACCATACCO	CGAATGTCAAG	AGCTGGTAAGG	ITCTGCGCGTTGCTTCGAATTAAA	850
Query	481	CCACATGCTCCACC	GCTTGTGCGGG	CCCCCGTCAAT	ICCTTTGAGTTTTAATCTTGCGAC	540
Sbjct	849	CCACATGCTCCACC	GCTTGTGCGGG	CCCCCGTCAAT	ICCTTTGAGTTTTAATCTTGCGAC	790
Query	541		GGAATGTTTAA	TGCGTTAGCTG	CGCCACCGAACAGTATACTGCCCG	600
nchinlmn	ih.gov/Bla	st.cai#dlaDwnl 1228924	77 AATGTTTAA	TGCGTTAGCTG	CGCCACCGAACAGTATACTGCCCG	730
Query	601	ACGGCTAACATTCA	TCGTTTACGGC	GTGGACTACCA(GGTATCTAATCCTGTTTGCTCCC	660
Sbjct	729	ACGGCTAACATTCA	TCGTTTACGGC	GTGGACTACCA	GGGTATCTAATCCTGTTTGCTCCC	670
Query	661	CACGCTTTCGCACC	TCAGCGTCAGT	AATGGACCAGTZ	AGCCGCCTTCGCCACTGGTGTTC	720
Sbjct	669	CACGCTTTCGCACC	TCAGCGTCAGT	AATGGACCAGT	AAGCCGCCTTCGCCACTGGTGTTC	610
Query	721	CTGCGAATATCTAC	GAATTTCACCT	CTACACTCGCA	ATTCCACTTACCTCTTCCATACTC	780
Sbjct	609	CTGCGAATATCTAC	GAATTTCACCT	CTACACTCGCA	ATTCCACTTACCTCTTCCATACTC	550
Query	781	AAGATACCCAGTAT	CAAAGGCAGTT		CTCTGGGATTTCACCCCTGACTTA	840
Sbjct	549	AAGATACCCAGTAT	CAAAGGCAGTT	CCAGAGTTGAG	CTCTGGGATTTCACCCCTGACTTA	490
Query	841	AATATCCGCCTACG	TGCGCTTTACG	CCCAGTAATTCO		900
Sbjct	489	AATATCCGCCTACG	TGCGCTTTACG	CCCAGTAATTCO	CGAACAACGCTAGCCCCCTTCGTA	430
Query	901	TTACCGCGGCTGCT	GGCACGAAGTT	AGCCGGG 932	2	
Sbjct	429	TTACCGCGGCTGCT	GGCACGAAGTT	AGCCGGG 398	3	



Pantoea septica KF475883.1

Pantoea septica strain IHB B 1545 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KF475883.1</u>| Length: 1508 Number of Matches: 1

Range 1:	68 to 97	5 GenBank Graphics			Next Match 🔺 Previous Match	
Score 1599 bits	s(1772)	Expect 0.0	Identities 900/908(99%)	Gaps 1/908(0%)	Strand Plus/Plus	
Query	1	ACAGAAGAGCTTG	CTCTTTGGGTGGC	GAGTGGCGGACGG	GTGAGTAATGTCTGGGAAAC	г 60
Sbjct	<mark>6</mark> 8	ACAGAAGAGCTTG	CTCTTTGGGTGGC	GAGTGGCGGACGG	GTGAGTAATGTCTGGGAAAC	I F 127
Query	61	GCCCGATGGAGGG	GGATAACTACTGG	AAACGGTAGCTAA	TACCGCATAACGTCGCAAGA	c 120
Sbjct	128	GCCCGATGGAGGG	GGATAACTACTGG	AAACGGTAGCTAA	TACCGCATAACGTCGCAAGA	C 187
Query	121	CAAAGTGGGGGGAC	CTTCGGGCCTCAC	ACCATCGGATGTG	CCCAGATGGGATTAGCTAGT.	A 180
Sbjct	188	CAAAGTGGGGGGAC	CTTCGGGCCTCAC.	ACCATCGGATGTG	CCCAGATGGGATTAGCTAGT.	A 247
Query	181	GGTGGGGTAACGG	CTCACCTAGGCGA	CGATCCCTAGCTG	GTCTGAGAGGATGACCAGCC.	A 240
Sbjct	248	GGTGGGGTAACGG	CTCACCTAGGCGA	CGATCCCTAGCTG	GTCTGAGAGGATGACCAGCC.	A 307
Query	241	CACTGGAACTGAG	ACACGGTCCAGAC	TCCTACGGGAGGC	AGCAGTGGGGAATATTGCAC.	A 300
Sbjct	308	CACTGGAACTGAG	ACACGGTCCAGAC	TCCTACGGGAGGC	AGCAGTGGGGGAATATTGCAC	A 367
Query	301	ATGGGCGCAAGCC	TGATGCAGCCATG	CCGCGTGTATGAA	GAAGGCCTTCGGGTTGTAAA	G 360
Sbjct	368	ATGGGCGCAAGCC	TGATGCAGCCATG	CCGCGTGTATGAA	GAAGGCCTTCGGGTTGTAAA	G 427
Query	361	TACTTTCAGCGGG	GAGGAAGGCGACG	CGGTTAATAACCG	CGTCGATTGACGTTACCCGC.	A 420
Sbjct	428	TACTTTCAGCGGG	GAGGAAGGCGACG	CGGTTAATAGCCG	CGTCGATTGACGTTACCCGC.	A 487
Query	421	GAAGAAGCACCGG	CTAACTCCGTGCC.	AGCAGCCGCGGTA	ATACGGAGGGTGCAAGCGTT.	A 480
Sbjct	488	GAAGAAGCACCGG	CTAACTCCGTGCC.	AGCAGCCGCGGTA	ATACGGAGGGTGCAAGCGTT.	A 547
Query	481	ATCGGAATTACTG	GGCGTAAAGCGCA	CGCAGGCGGTCTG	TCAAGTCGGATGTGAAATCC	C 540
Sbjct	548	ATCGGAATTACTG	GGCGTAAAGCGCA	CGCAGGCGGTCTG	TCAAGTCGGATGTGAAATCC	c 607
Query	541	CGGGCTTAACCTG	GGAACTGCATTCG	AAACTGGCAGGCT	AGAGTCTTGTAGAGGGGGGGT.	A 600
Sbjct	608	CGGGCTTAACCTG	GGAACTGCATTCG	AAACTGGCAGGCT	AGAGTCTTGTAGAGGGGGGT.	A 667
Query	601	GAATTCCAGGTGT	AGCGGTGAAATGC	GTAGAGATCTGGA	GGAATACCGGTGGCGAAGGCG	660
Sbjct	668	GAATTCCAGGTGT	AGCGGTGAAATGC	GTAGAGATCTGGA	GGAATACCGGTGGCGAAGGCG	727
Query	661	GCCCCCTGGACAA	AGACTGACGCTCA	GGTGCGAAAGCGT	GGGAGCAAACAGGATTAGAT	720
Sbjct	728	GCCCCCTGGACAA	AGACTGACGCTCA	GGTGCGAAAGCGT	GGGGAGCAAACAGGATTAGAT	787
Query	721	ACCCTGGTAGTCC	ACGCCGTAAACGA	IGTCGACTTGGAG	GCTGTTCCCCTGAGGAGTGGC	780
Sbjct	788	ACCCTGGTAGTCC	ACGCCGTAAACGA	IGTCGACTTGGAG	GCTGTTCCCTTGAGAAGTGGC	847
Query	781	TTCCGGAGCTAAC	GCGTTAAGTCGAC	CGCCTGGGGAGTA	CGGGCGCAAGGTTAAAACTCA	840
Sbjct	848	TTCCGGAGCTAAC	GCGTTAAGTCGAC	CGCCTGGGGAGTA	CGGCCGCAAGGTTAAAACTCA	907
Query	841	AATGAATTGACGG		CGGGGGGAGCATGT	GGTTTAATTCGATGC-ACGCG	899
Sbjct	908	AATGAATTGACGG	GGGCCCGCACAAG	CGGTGGAGCATGT	GGTTTAATTCGATGCAACGCG	967
Query	900	AAAAACCT 907				
Sbjct	968	AAGAACCT 975				



Stenotrophomomnas rhizophila KP050794.1

Stenotrophomonas rhizophila strain HT12-MRL 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb|KP318062.1</u>| Length: 1295 Number of Matches: 1

Range 1:	Range 1: 59 to 491 GenBank Graphics			🔻 Next Match 🔺 Previous Match			
Score		Expect	Identities	Gaps	Strand		
760 bits	(842)	0.0	429/433(99%)	1/433(0%)	Plus/Plus		
Query	37	GGGTGAGGAA-A	ACATCGTTGTCTACC	TTTTCGTGGGGGGAT.	AACGTAGGGAAACTTACGC	г 95 I	
Sbjct	59	GGGTGAGGAAT	ACATCGGAATCTACCI	TTTTCGTGGGGGGAT.	AACGTAGGGAAACTTACGC	, r 118	
Query	96		CGACCTTCGGGTGAA	AGCAGGGGGACCTTC	GGGCCTTGCGCGGATAGAT	G 155 I	
Sbjct	119	AATACCGCATA	CGACCTTCGGGTGAA	AGCAGGGGGACCTTC	GGGCCTTGCGCGGATAGAT	G 178	
Query	156	AGCCGATGTCG	GATTAGCTAGTTGGCC	GGGTAAAGGCCCA	CCAAGGCGACGATCCGTAG	C 215	
Sbjct	179	AGCCGATGTCG	GATTAGCTAGTTGGCC	GGGTAAAGGCCCA	CCAAGGCGACGATCCGTAG	238	
Query	216	TGGTCTGAGAG	GATGATCAGCCACACI	GGAACTGAGACAC	GGTCCAGACTCCTACGGGA	G 275	
Sbjct	239	TGGTCTGAGAG	GATGATCAGCCACACI	IGGAACTGAGACAC	GGTCCAGACTCCTACGGGA	G 298	
Query	276	GCAGCAGTGGG	GAATATTGGACAATGO	GCGCAAGCCTGAT	CCAGCCATACCGCGTGGGT	G 335	
Sbjct	299	GCAGCAGTGGG	GAATATTGGACAATGO	GCGCAAGCCTGAT	CCAGCCATACCGCGTGGGT	, 3 358	
Query	336	AAGAAGGCCTT	CGGGTTGTAAAGCCCT	TTTTGTTGGGAAAG	AAAAGCAGTCGATTAATAC	r 395	
Sbjct	359	AAGAAGGCCTT	CGGGTTGTAAAGCCCI	TTTTGTTGGGAAAG.	AAAAGCAGTCGATTAATAC	r 418	
Query	396	CGGTTGTTCTG	ACGGTACCCAAAGAAT	TAAGCACCGGCTAA	CTTCGTGCCAGCAGCCGCG	G 455	
Sbjct	419	CGGTTGTTCTG	ACGGTACCCAAAGAAT	TAAGCACCGGCTAA	CTTCGTGCCAGCAGCCGCG	G 478	
Query	456	TAATACGAAGG	GT 468				
Sbjct	479	TAATACGAAGG	GT 491				

		Clouery_10905	A Deservation of a D 1/P channel BMA and a series and a language
			 Scholoppionionals sp. P 105 mossonial KXA gene, partial sequence Uncultured Stenotrophomonas sp. clone CSC13DXR2A 44 16S ribosomal RNA gene, partial sequence
			⁴ Stenotrophomonas sp. partial 16S rRNA gene, isolate BA-91-09
			Stenotrophomonas rhizophila partial 16S rRNA gene, isolate P4D11
			Stenetrophomonas rhizophila partial 165 rRNA gene, isolate P3G11 Stenetrophomonas rhizophila partial 165 rRNA gene, isolate P3G11
			Stentonophoninas mizophila partal 105 rRNA eene, isolate P5/12 Stentonophononas rhizophila partal 105 rRNA eene, isolate P5/7
		Ť l	Stenotrophomonas sp. DS-8PS-13 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. DS-8PS-10 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. DS-7PS-8 16S ribosomal RNA gene, partial sequence
			Stenetrophomonas sp. DS-3PS-10-16S ribosomal RNA gene, partial sequence Stenetrophomonas en DS-SIS 0-16S ribosomal RNA gene, partial sequence
			Stentorophonanas sp. D5-75-7 105 nosonia RAA gene, partal sequence Stentorophonanas sp. D5-75-6 16S ribosonia RAA gene, partal sequence
			Stenotrophomonas sp. DS-3PS-12 16S ribosomal RNA cene, partial sequence
			Stenotrophomonas rhizophila strain P8 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. KC4-MRL 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. 3Z7 16S ribosomal RNA gene, partial sequence
			Stenetrophomonas sp. DF9D 16S ribosomal RNA gene, partial sequence Stenetrophomonas sp. EECC 551 16S ribosomal RNA gene, partial sequence
			Stendorophomonas sp. EECC-551 to 5 monoma RVA gene, partal sequence Stendorophomonas sp. EECC-521 IOS monoma RVA gene, partal sequence
			Uncultured bacterium clone 4II.10H 16S ribosomal RNA gene, partial sequence
			Output de la clerium clone 4II.29 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. LVo-amp_7-5 16S ribosomal RNA gene, partial sequence
			Stanctorophomonas sp. REp-tel_144 105 nbosonal KNA gene, partai sequence Stanctorophomonas ribitzabila cartial 165 RENA gene etapia G15
			Stenotophoninas in Zophia partain OB-TRAA gene, suan OLD
			Stenotrophomonas sp. SOZ2-4071 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. SOZI-7104 16S ribosomal RNA gene, partial sequence
			Uncultured Stenotrophomonas sp. clone CTL-92 16S ribosomal RNA gene, partial sequence
			Stenetrophomonas mizophia strain Bacteria 188 105 ntosomai RNA gene, partiai sequence Stenetrophomonas rhizophia strain Bacteria 187 105 ribosomai BNA gone partiai sequence
			Stentorophononas inzophia starii Bactera_167 105 Hosoniai ROA gene nariisleenee
			Stenotrophomonas sp. S169-III-5 partial 16S rRNA gene, isolate S169-III-5
			Stenotrophomonas rhizophila strain AB11 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. BSw21910 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. DF17TA 16S ribosomal RNA gene, partial sequence OFTPM UNLOG OFTPMU ICO OFTPU ICO OFTPU ICO OFTPMU ICO OFTPU
			Stenetrophomonas sp. GCDP10_III 105 nbosomai KNA gene, partial sequence Stenetrophomonas rhizorbila strain KC0 165 cibocoma IRNA gene, partial sequence
			Stentorophomonas imzopina suam (C) 105 novsonat KVA gene, partial sequence Stentorophomonas n. PDD-321-30 165 ribosonal RVA gene, partial sequence
			Stenotrophomonas maltophilia strain SGb417 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas maltophilia strain SGb391 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas rhizophila strain PCA_13 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas maltophilia gene for 16S rRNA, partial sequence, strain: MUJ
			Cincutared bacterium clone N-32 165 nbosomal RNA gene, partial sequence Standbommans of S2D32(011) 165 thoograph DNA gene partial sequence
			Stentrophonomass, Statistics in Strophone BA-145
			Stenotrophomonas sp. partial 16S rRNA gene, isolate BA-146
			Stenotrophomonas maltophilia strain D46 Z-22-2 I6S ribosomal RNA gene, partial sequence
			Schotrophomonas sp. Dra15 16S ribosomal RNA gene, partial sequence
			Senotrophomonas sp. Dra20 16S ribosomal RNA gene, partial sequence
			Schotrophomonas sp. Dra23 105 ribosomal KNA gene, partial sequence
			Scenotrophomonas sp. Oral 5 105 ribosomal RNA gene, partial sequence
			Scholophonikolas Sp. Aladz 105 mossonial RNA gene, partai sequence
			Schotrophomonas sp. Adap 165 fibosomal RNA gene, partai sequence
			Standardhomonas sp. (255) 105 Hotsonial RivA gene, partial aquence
			Stendrophonical as sp. Awar 105 mossimal RNA gene, partial sequence
			Senterophononas sp. Awal 4 ISS ribosomal RVA gene, partial sequence
			Standardshanasas hizohlia ttain LV3 165 ribosona RVA gene, panar sequence
			Stendrophomonas trizophila strain LK25 165 Bosonia RVA gene, partial sequence
			Senotrophomonas rhizophila strain PN8 16S ribosomal RNA eene, nartial sequence
			Stenotrophomonas rhizophila strain SK16 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas rhizophila strain SN1 16S ribosomal RNA gene, partial sequence
			Bacterium amp-w-7 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas rhizophila partial 16S rRNA gene, isolate U10
			Stenotrophomonas rhizophila strain P-21 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas rhizophila partial 16S rRNA gene, strain S8
			Stenotrophomonas sp. partial 16S rRNA gene, isolate BD17-E03
			Stenotrophomonas rhizophila partial 16S rRNA gene, isolate BD17-E04
			Xanthomonadaceae bacterium BvORR135 gene for 16S rRNA, partial sequence
			Stenotrophomonas sp. MM65May 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. PA21Nov 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. PA59Nov 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. PA53Nov 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas sp. PA67Nov 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas rhizophila strain DSM14405 genome
			Stenotrophomonas rhizophila strain FS16-1 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas maltophilia strain FGS4-1 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas rhizophila strain DKB1 16S ribosomal RNA gene, partial sequence
			Stenotrophomonas mizophila strain e-p10 165 nbosomal RNA gene, complete sequence Spentification and Cliffic de all and a strain a strain and a strain a strain and a strain a strain and
			 Menotrophomonas sp. G1_0 165 ribosomal RNA gene, partial sequence
			Stenstrophomonas sp. Y4 288 6 105 ribosomal RNA gene, partial sequence
			Stendorophomonas sp. 74–507 1 16S nbosomal KNA gene, partial sequence
			Stendorophomonas sp. 14 507 1 16S nbosomal KNA gene, partial sequence
			Techorophomonas sp. 14 - 0.22 105 ribosomal RNA gene, partial sequênce
			Techorophomonas sp. 14–231_3 IOS fibosomal RNA gene, partial sequênce
			Technologinomonas sp. 14 252 2 105 mossonial Kiva gene, partial sequence
			Stendorophomonas sp. 14 _2.5 _3 105 mosomal RNA gene, partial sequence Standrophomonas en BLTBLIAD IAS thosomal DNA once and partial sequence
			Standarophomona sp. D 3D 147 105 D050mal KNA gene, partial sequence
			Stendardynomias attraphila statil 1007-105 trosoniar Kiva gelle, partial sequence
			Totelouophonanaa sp. ress. partai 105 IKiva gene, isolate Pesso a Daonbascillus en Dast (f) naria 165 ePA aona isolata Dast f()
			reactions in us ap. rest to partial ros risks gene, isolate rest to
			Stendrophomonas in NCCP-549 one for 16S ribosomal RNA, partial sources
			Proteins on NCCP-655 one for 16S ribosomal RNA partial sequence
			Stendarohomonas en AlsPAR ISS ribosomal RNA, panar Sequence
			Stendtophomonas p. 710 7D 105 10500ma RNA guie, partial sequence
0.0008	1		Stendtrohomonas p. 460/51 los ritosoma RNA gene, partal sequence
	1		Bacterium C0088 16S ribosomal RNA gene, partial sequence
			Stenotophomonas thizophila strain HT12-MRL 165 ribosomal RNA cone, partial sequence
			sector operations of the plant south for the operation in the gene, partial sequence

Citrobacter freundii CP007557

Citrobacter freundii CFNIH1, complete genome Sequence ID: <u>gb|CP007557.1|</u> Length: 5099034 Number of Matches: 1

Range 1:	5039222 to 5039628 GenBank Graphics Vext Match 🔺 Previous Match						
Score		Expect	Identities	Gaps	Strand		
477 bits(528)	6e-131	353/408(87%)	3/408(0%)	Plus/Plus		
Features:	<u>hypothetical pr</u> phospho-2-deh	r <u>otein</u> 1ydro-3-deoxy	heptonate aldolase				
Query	1	TTTTTGC	ATGATGGTGATCCTG	TTTAGCTCGTTTGC	GATAGTTGATCCT	CAGCGAGGAGG	60
Sbjct	5039222	TTTTTGC	ATGATGGTGATCCTG	TTTAGCTCGTTTGC	GATAGTTGATCCT	CAGCGAGGAGG	5039281
Query	61	AAATAAC	GATACCATAACAGGT	AAAGATTTCAATCO	CACAATCCGTAAAT	TTTAATTTACA	120
Sbjct	5039282	AAATAAC	GATATCACAACAGGT	AAAGATTTCAATCO	CATAATCCGTAAAA	IIIIIIIIIII TTTAATTTACA	5039341
Query	121	CAGTGTT	ATTTTAAGGCAAAAA	TAGCCCTT-ATAAA	ATGTACACTTAAAT	TTACACCA-CG	178
Sbjct	5039342	TCGAGCT	'ATTA-ATCACTAAAT	AAGGCCTACATGAG	TGTATAGTTAAAT	TTACACAATCC	5039400
Query	179	AATTTTC	AGATCCGCTATGCTT	aaaaaaCAAGGGA	AGCACAGGCAATGA	AGCAACTCATC	238
Sbjct	5039401	TGATATC	AGATCCACTATGCTT	AAAAAAACGCGGGA	AGCACAGACAATGA	AGCAACTCATC	5039460
Query	239	AGCATCT	TATTTCTATTCCTTC	TTAGCGGATGTCAG	ATAGATCCCTACA	CCCATGCCCCT	298
Sbjct	5039461	AGCACCT	TATTAATACTACTTC	TTAGCGGATGTCAG	GATAGACCCCTATA	CTCACGCCCCT	5039520
Query	299	ACCTGGA	CTGGTACCGACTGGT	ATGACGCCGGGATA	CAAGATGCCATTT	CGGGCTATGCC	358
Sbjct	5039521	ACCTGGA	CTGGCACCGACTGGT	ACGATGCCGGCATA	ACGCGATGCCATTT	CAGGCTATGCG	5039580
Query	359	GTTAAAG	ATAATGAAACTCTTG	CCGACAACTTCAAT	GATCCCGAAGTC	406	
Sbjct	5039581	GTTAAAG	ATAATGAAATTCTTG	CCGACAATTACAAT	GATCCAGAAGTC	5039628	



Pseudomonas frederiksbergensis EU373369.1

Pseudomonas frederiksbergensis strain PR18 16S ribosomal RNA gene, partial sequence Sequence ID: <u>gb[KJ870030.1]</u> Length: 1449 Number of Matches: 1

Range 1:	832 to 13	350 <u>GenBank</u> <u>Gra</u>	<u>phics</u>	V	Next Match 🔺 Previous N	Match	
Score	(1018)	Expect	Identities	Gaps	Strand Plus/Minus		
919 DIG	(1010)	0.0	515/515(55%)	0/313(070)	Flus/Pillus		
Query	6	GTGTGTAAAA	TGCGCGGGGAACGTAT	TCACCGCGACATI	CTGATTCGCGATTA	CTAGCGAT	65
Sbjct	1350	GTGTGTACAA	GGCCCGGGAACGTA	TTCACCGCGACATI	CTGATTCGCGATTA	CTAGCGAT	1291
Query	66	TCCGACTTCA	CGCAGTCGAGTTGC	AGACTGCGATCCGG	ACTACGATCGGTTT	TATGGGAT	125
Sbjct	1290	TCCGACTTCA	CGCAGTCGAGTTGC	AGACTGCGATCCGG	ACTACGATCGGTTT	TCTGGGAT	1231
Query	126	TAGCTCCACO	TCGCGGCTTGGCAA	CCTCTGTACCGAC	CATTGTAGCACGTG	TGTAGCCC	185
Sbjct	1230	TAGCTCCACC	TCGCGGCTTGGCAA	CCTCTGTACCGAC	CATTGTAGCACGTG	TGTAGCCC	1171
Query	186	AGGCCGTAAG	GGCCATGATGACTTO	GACGTCATCCCCAC	CTTCCTCCGGTTTG	TCACCGGC	245
Sbjct	1170	AGGCCGTAAG	GGCCATGATGACTT	GACGTCATCCCCAC	CTTCCTCCGGTTTG	TCACCGGC	1111
Query	246	AGTCTCCTTA	GAGTGCCCACCATT	ACGTGCTGGTAACI	AAGGACAAGGGTTG	CGCTCGTT	305
Sbjct	1110	AGTCTCCTTA	GAGTGCCCACCATT	ACGTGCTGGTAACI	AAGGACAAGGGTTG	CGCTCGTT	1051
Query	306	ACGGGACTTA	ACCCAACATCTCAC	GACACGAGCTGACG	ACAGCCATGCAGCA	CCTGTCTC	365
Sbjct	1050	ACGGGACTTA	ACCCAACATCTCAC	GACACGAGCTGACG	ACAGCCATGCAGCA	CCTGTCTC	991
Query	366	AATGTTCCCG	AAGGCACCAATCCA	CTCTGGAAAGTTC	ATTGGATGTCAAGG	CCTGGTAA	425
Sbjct	990	AATGTTCCCG	AAGGCACCAATCCA	CTCTGGAAAGTTC	ATTGGATGTCAAGG	CCTGGTAA	931
Query	426	GGTTCTTCGC	GTTGCTTCGAATTA	ACCACATGCTCCA	CCGCTTGTGCGGGC	CCCCGTCA	485
Sbjct	930	GGTTCTTCGC	GTTGCTTCGAATTA	ACCACATGCTCCA		CCCCGTCA	871
Query	486	ATTCATTTGA	GTTTTAACCTTGCGG	SCCGTACTCCCCCA	G 524		
Sbjct	870	ATTCATTTGA	GTTTTAACCTTGCG	GCCGTACTCCCCCA	I AG 832		

9 kilQuery_233167	
Viseudomonas sp. HNR15 165 ribosomai KNA gene, partiai sequence	Pseudomonas vancouverensis strain BDd4 16S ribosomal RNA gene, partial sequen
	Uncultured bacterium clone B12 16S ribosomal RNA gene, partial sequence
•	g-proteobacteria 4 leaves
	Pseudomonas putida strain DNRS04 16S ribosomal RNA gene, partial sequence
Pseudomonas sp. s1p25 165 nbosomal RNA gene, partial sequence Pseudomonas sp. s2p12 165 ribosomal RNA gene, partial sequence	
Pseudomonas sp. s2p21 165 ribosomal RNA gene, partial sequence	
Pseudomonas sp. s2p23 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. s3p21 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. s4p22 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. BSP27 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. PT19 16S ribosomai KNA gene, partial sequence Disautomonas en SCALW0301 16S ribosomal DNA gana matial sequence	
Pseudomonas sp. 5C/COC0501 ToS Tibosonial RVA gene, partial sequence	
Pseudomonas sp. P32(2013) 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. QW11 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone CD36 16S ribosomal RNA gene, partial sequence	
Pseudomonas chlororaphis partial 16S rRNA gene, isolate ToZa7	
 Uncultured Pseudomonas sp. clone BJPS522-001 165 ribosomal RNA gene, partial sequence Pseudomonas cadrina strain LCO.5 165 ribosomal RNA gana partial sequence 	
Preudomonas en RA-20 16S ribosomal RNA cone nartial sequence	
Pseudomonas sp. S10114 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. SB442 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. XBBSA2 16S ribosomal RNA gene, partial sequence	
Pseudomonas extremorientalis strain EB-171 16S ribosomal RNA gene, partial sequence	
Pseudomonas vancouverensis strain Amp46 16S ribosomal RNA gene, partial sequence	
 Preudomonas extremorientalis strain NSPIBS/02 165 ribosomal KNA gene, partial sequence Preudomonas extremorientalis strain NSPIBs/02 165 ribosomal DNA anna partial sequence 	
Pseudomonas executionentais suatu (VSF10X/2-105-1100sonia) revy gene, partial sequence Pseudomonas sp. P2-1 16S ribosonal RNA gene, nartial sequence	
Pseudomonas chlororaphis strain 7,3B 16S ribosomal RNA gene, nartial sequence	
Pseudomonas sp. C34 16S ribosomal RNA gene, partial sequence	
Endophytic bacterium 10 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IN2CAET02BYJJ0 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. ctone IE34LQE05GD48X 16S ribosomal RNA gene, partial sequence	
 Uncultured Pseudomonas sp. ctone IE:54LQE/04EFH0G 16S ribosomal KNA gene, partial sequence Uncultured Pseudomonas en clone IE:341 OF/07L7/C1 16S ribosomal RNA gene, partial environmente 	
 Uncultured Pseudomonas sp. cone fi2942.0E07IDF3Y 16S ribosomal RNA gene, partial sequence Uncultured Pseudomonas sp. clone IE34LOE07IDF3Y 16S ribosomal RNA gene rartial sequence 	
Uncultured Pseudomonas sp. clone IE34LQE07IKPDF 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IE34LQE08JTYDU 16S ribosomal RNA gene, partial sequence	
⁹ Uncultured bacterium clone LY62 16S ribosomal RNA gene, partial sequence	
⁹ Uncultured bacterium clone HZ37 16S ribosomal RNA gene, partial sequence	
 Pseudomonas sp. QJX-1 IoS ribosomal RNA gene, partial sequence Braudomonas on AD157 IoS ribosomal RNA gene, partial sequence 	
Preudomonas sp. AD157 105 Horsonial ROA gene, partial sequence Preudomonas azotoformans strain BG. D. 16S ribosomal RNA gene nartial sequence	
Pseudomonas azonoromans scain Do D fos friosomal RNA gene, partial sequence	
Pseudomonas azotoformans strain Sn16 16S ribosomal RNA gene, partial sequence	
Pseudomonas azotoformans strain Sn22 16S ribosomal RNA gene, partial sequence	
Pseudomonas azotoformans strain Sn47 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. DR 1-03 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. GR 7-06 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. NR 6-08 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. U 1 3-02 165 ribosomal KNA gene, partial sequence	
Uncultured Pseudomonas sp. clone RHDI WG191 105 ribosomal RNA gene	partial sequence
Uncultured Pseudomonas sp. clone RHD1 w0205 105 fibosomal RNA gene.	partial sequence
Uncultured Pseudomonas sp. clone RHD1 wG209 105 H00somal RNA gene,	partial sequence
Dreudomonas sp. COR-2P 16S ribosomal PNA cene partial sequence	partial sequence
Rhodococcus sp. COB-21 105 Hossonial RIVA gene, partial sequence	
Preudomonas marginalis strain CTE722-C 16S ribosomal RNA gene, partial	sequence
Pseudomonas sp. ACP 01 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seg M-R15 16SR 16S ribosomal RNA eene. no	artial sequence
Vucultured prokaryote clone sea M-R17 16SR 16S ribosomal RNA eene, p	artial sequence
Quncultured prokaryote clone seg M-R43 16SR 16S ribosomal RNA gene, by	artial sequence
Uncultured prokaryote clone seq M-R48 16SR 16S ribosomal RNA gene, p	artial sequence
Uncultured prokaryote clone seq_M-R49_16SR 16S ribosomal RNA gene, p.	artial sequence
Uncultured prokaryote clone seq_M-S13_16SR 16S ribosomal RNA gene, page 100 PM and 10	utial sequence
Uncultured prokaryote clone seq_M-S15_16SR 16S ribosomal RNA gene, page 10 Provide the set of th	irtial sequence
Uncultured prokaryote clone seq_M-S19_16SR 16S ribosomal RNA gene, page 100 PM and 10	utial sequence
 Uncultured prokaryote clone seq_M-S22_16SR 16S ribosomal RNA gene, page 	utial sequence
 Uncultured prokaryote clone seq_M-S25_16SR 16S ribosomal RNA gene, pa 	utial sequence
Uncultured prokaryote clone seq_M-S29_16SR 16S ribosomal RNA gene, page 100 PM and 10	utial sequence
Uncultured prokaryote clone seq_M-S3_16SR 16S ribosomal RNA gene, par Uncultured prokaryote clone seq_M-S3_16SR 16S ribosomal RNA gene, par	tial sequence
 Uncultured prokaryote clone seq_M-S30_16SR 16S ribosomal RNA gene, pa Unaultured prokaryote clone seq_M-S30_16SR 16S ribosomal RNA gene, pa 	irtial sequence
Uncutatived prokaryote clone seq_M-531_165K 165 ribosomal RNA gene, pa	iriai sequence
Uncultured prokaryote clone seq_M-S35_105K 16S ribosomal RNA gene, pa Uncultured prokaryote clone seq_M_S35_16SB 16S ribosomal RNA gene, pa Uncultured prokaryote clone seq_M_S35_16SB 16S ribosomal RNA gene, pa	irtai sequence
 Uncultured prokaryote clone seq_NI-53/_105K 105 ribosomal KNA gene, pa Uncultured prokaryote clone seq_MI-247_16SD 16S ribosomal DNA sono model 	urial sequence
Uncultured prokaryote clone seq_Nr-54/_105K 105 ribosonial KINA gene, pa	tial sequence
 Uncultured prokaryote clone seq_51-55_105K 105 fibosomal KNA gene, par Uncultured prokaryote clone seq_M_S56_16SR 16S ribosomal DNA gene, par 	intial sequence
Uncultured protections seq_01-5.50_105K 105 ribosonial KIVA gene, pa	tial sequence
Uncultured prokaryote clone seq_NI-59_105K 105 fibosomal RNA gene, par	rtial sequence
Uncultured prokaryote clone seq 1*R10_105R 105 10050ma1 RNA gene, pa	rtial sequence
Uncultured prokaryote clone sed T_R34_16SR 16S ribosomal RNA gene, pa	rtial sequence
Uncultured prokaryote clone seq T-R47 16SR 16S ribosomal RNA gene, pa	rtial sequence
Uncultured prokatyote clone seq_1-ret/_rosk ros noosonal RivA gene, pa Uncultured prokatyote clone seq_T_S43_16SR 16S ribosonal RNA gene, pa	rtial sequence
Uncultured prokaryote clone sea T-S46 16SR 16S ribosomal RNA gene, pa	rtial sequence
Uncultured protaryote clone seq T-S9 I6SR 16S ribosonial RNA gene, part	ial sequence
Uncultured prokaryote clone seg C04 Z1224 Z014717a W-X2 16SR 16S	ibosomal RNA gene, partial sequence
Vucultured prokaryote clone sea C05 Z1224 Z014718a W-X3 16SR 16S	ibosomal RNA gene, partial sequence
Uncultured prokaryote clone sea C06 Z1224 Z014719a W-X6 16SR 16S	ibosomal RNA gene, partial sequence
Uncultured prokaryote clone seg H08 Z118570 Z079335a W-X1 16SR 16	S ribosomal RNA gene, partial sequence
Uncultured prokaryote clone seq_H09_Z118570_Z079336a_W-X9_16SR 16	S ribosomal RNA gene, partial sequence
g-proteobacteria 4 leaves	